Final risk analysis report for the release of *Stomphastis* *thraustica* for the biological control of *Jatropha gossypiifolia*

April 2022



© Commonwealth of Australia 2022

**Ownership of intellectual property rights**

Unless otherwise noted, copyright (and any other intellectual property rights, if any) in this publication is owned by the Commonwealth of Australia (referred to as the Commonwealth).

**Creative Commons licence**

All material in this publication is licensed under a [Creative Commons Attribution 4.0 International Licence](https://creativecommons.org/licenses/by/4.0/legalcode), except content supplied by third parties, logos and the Commonwealth Coat of Arms.

Inquiries about the licence and any use of this document should be emailed to copyright@awe.gov.au.



**Cataloguing data**

This publication (and any material sourced from it) should be attributed as: DAWE 2022, *Final risk analysis report for the release of* Stomphastis thraustica *for the biological control of* Jatropha gossypiifolia, Department of Agriculture, Water and the Environment, Canberra. CC BY 4.0.

This publication is available at [awe.gov.au/biosecurity-trade/policy/risk-analysis/biological-control-agents](http://www.awe.gov.au/biosecurity-trade/policy/risk-analysis/biological-control-agents).

Department of Agriculture, Water and the Environment
GPO Box 858 Canberra ACT 2601

Telephone 1800 900 090

Web [awe.gov.au](https://www.awe.gov.au/)

Email plantstakeholders@awe.gov.au

**Liability**

The Australian Government acting through the Department of Agriculture, Water and the Environment has exercised due care and skill in preparing and compiling the information in this publication. Notwithstanding, the Department of Agriculture, Water and the Environment, its employees and advisers disclaim all liability, including liability for negligence and for any loss, damage, injury, expense or cost incurred by any person as a result of accessing, using or relying upon any of the information or data in this publication to the maximum extent permitted by law.

Contents

Acronyms and abbreviations v

Summary 1

1 Introduction 2

1.1 Australia’s biosecurity policy framework 2

1.2 This risk analysis 2

2 Assessment of off-target risks 5

2.1 Stage 1: Initiation 6

2.2 Stage 2: Risk assessment 6

3 Recommendation on release 12

4 Attachment 1 12

5 Stakeholder responses to draft risk analysis report 13

Appendix A: Method for pest risk analysis 15

Stage 1 Initiation 15

Stage 2 Pest risk assessment 16

Stage 3 Pest risk management 23

Glossary 25

References 29

Tables

Table 2.1 Risk estimation matrix. 5

Maps

Map 1 Map of Australia iv

Map 2 A guide to Australia’s bio-climatic zones iv

Map 1 Map of Australia



Map 2 A guide to Australia’s bio-climatic zones



Acronyms and abbreviations

| Term or abbreviation | Definition |
| --- | --- |
| ACT | Australian Capital Territory |
| ALOP | Appropriate level of protection |
| BA | Biosecurity Advice |
| BCA | Biological Control Agent |
| BICON | The Australian Department of Agriculture, Water and the Environment Biosecurity Import Conditions database |
| BIRA | Biosecurity Import Risk Analysis |
| CSIRO | Commonwealth Scientific and Industrial Research Organisation |
| FAO | Food and Agriculture Organization of the United Nations |
| IPC | International Phytosanitary Certificate |
| IPPC | International Plant Protection Convention |
| ISPM | International Standard for Phytosanitary Measures |
| NSW | New South Wales |
| NPPO | National Plant Protection Organisation |
| NT | Northern Territory |
| PRA | Pest risk assessment |
| Qld | Queensland |
| SA | South Australia |
| SPS Agreement | WTO Agreement on the Application of Sanitary and Phytosanitary Measures |
| Tas. | Tasmania |
| the department | The Australian Government Department of Agriculture, Water and the Environment |
| Vic. | Victoria |
| WA | Western Australia |
| WTO | World Trade Organization |

Summary

The Department of Agriculture, Water and the Environment has prepared this final report to assess the proposal by the Queensland Department of Agriculture and Fisheries (QDAF) to release *Stomphastis* *thraustica* for the biological control of *Jatropha gossypiifolia* in Australia.

This final report recommends that the release of *S.* *thraustica* should be permitted, subject to standard quarantine conditions associated with the import and release of exotic biological control agents.

This final report has determined the overall risk associated with the release of *S.* *thraustica* to be Negligible. A risk estimate of Negligible achieves Australia’s appropriate level of protection (ALOP).

The assessment of risk to off-target plants included consideration of the testing methodology used and the plant species test list, including non-target species tested in described experiments. The biology of *S.* *thraustica* was also considered. Following the publication of the draft report for stakeholder consultation in November 2021, the previously named *Stomphastis* sp. nov. was identified as *Stomphastis thraustica*. This final report reflects the species level identification.

This final report also contains details of the risk assessment process used for consideration of potential off-target effects associated with the proposed release of *S.* *thraustica* into the Australian environment.

There is also an approval process for the import and release of biological control agents under the *Environment Protection and Biodiversity Conservation Act 1999* within the department. The approval process under that Act will commence upon finalisation of this risk analysis process.

Two submissions were received from stakeholders during the consultation period for the draft report. These submissions were taken into consideration in preparing the final report.

The application from QDAF that was provided to the department has been included with this final report (Attachment 1).

# Introduction

## Australia’s biosecurity policy framework

Australia's biosecurity policies aim to protect Australia against the risks that may arise from exotic pests entering, establishing and spreading in Australia, thereby threatening Australia's unique flora and fauna, as well as those agricultural industries that are relatively free from serious pests.

The risk analysis process is an important part of Australia’s biosecurity policies. It enables the Australian Government to formally consider the level of biosecurity risk that may be associated with proposals to import goods or biological materials into Australia. If the biosecurity risks do not achieve the appropriate level of protection (ALOP) for Australia, risk management measures are proposed to reduce the risks to an acceptable level. If the risks cannot be reduced to an acceptable level, the goods or biological materials will not be imported into Australia until suitable measures are identified.

Successive Australian Governments have maintained a stringent, but not a zero risk, approach to the management of biosecurity risks. This approach is expressed in terms of the ALOP for Australia, which is defined in the *Biosecurity Act 2015* as providing a high level of protection aimed at reducing risk to a very low level, but not to zero.

Australia’s risk analyses are undertaken by the Department of Agriculture, Water and the Environment using technical and scientific experts in relevant fields and involve consultation with stakeholders at various stages during the process.

Further information about Australia’s biosecurity framework is provided in the *Biosecurity* *Import Risk Analysis Guidelines 2016* located on the [Australian Government Department of Agriculture, Water and the Environment website](https://www.awe.gov.au/biosecurity-trade/policy/risk-analysis/guidelines).

## This risk analysis

### Background

An application has been submitted by QDAF to release a biological control agent (Attachment 1). The identified biological control agent, *S.* *thraustica* is a leaf-mining moth proposed for the biological control of *J. gossypiifolia* (bellyache bush). The applicant has followed the steps outlined in the [Biosecurity Guidelines](https://www.awe.gov.au/biosecurity-trade/policy/risk-analysis/biological-control-agents/protocol_for_biological_control_agents) for the Introduction of Exotic Biological Control Agents for the Control of Weeds and Plant Pests.

*Jatropha gossypiifolia* is a woody perennial shrub, commonly known as bellyache bush. It is native to Central and South America. In Australia, it is a Weed of National Significance (WONS) found in rangelands in tropical areas, particularly in riparian zones. *Jatropha gossypiifolia* has naturalised throughout many areas of Queensland, Western Australia and the Northern Territory (Attachment 1).

*Jatropha gossypiifolia* is a major environmental and agricultural weed in Australia that is difficult and costly to control. It is toxic to both livestock and humans and forms monocultures, preventing growth of pasture species. It also reduces biodiversity and can cause soil erosion and destabilisation of river banks (Attachment 1).

*Stomphastis* *thraustica* is a small leaf-mining moth previously only recorded from Africa and Asia. The population reported on in the application was collected from Peru; this is the first known record for a *Stomphastis* species from South America. The most damaging life stage to *J. gossypiifolia* are the larvae, which have the potential to rapidly destroy leaves via their mines. Adult moths have a high fecundity and short generation time (Attachment 1).

A colony of *S.* *thraustica* was established in quarantine in Australia using individuals imported from Peru and it was this colony that was used in the host specificity testing reported in the application. If release is approved this colony will be used for mass-rearing and release. Since the publication of the draft report, *Stomphastis* sp. nov. has been identified as *Stomphastis thraustica*. This final report has been updated with the correct species name. A full genome sequence has also been completed and has been submitted to GenBank. It is noted that the *S. thraustica* colony from Peru with which host specificity testing was conducted is genetically distinct from *S. thraustica* populations found in Africa. However, this genetic difference is not considered sufficient to consider the population held in quarantine to be a different species (see Attachment 1, page 7 and Appendix 1 for further information).

### Scope

The scope of this risk analysis is to consider the biosecurity risk that may be associated with the release of an exotic biological control agent into the Australian environment (excluding its external territories). The primary risk associated with a release of this nature is the possibility of unwanted off-target effects on other species already present in Australia. The Department of Agriculture, Water and the Environment assesses the risk under the *Biosecurity Act 2015*. There is also an approval process within the department under the *Environment Protection and Biodiversity Conservation Act 1999*. Under section 303EE(4) of the *Environment Protection and Biodiversity Conservation Act 1999*, risk analysis reports prepared by the department may be used by the Minister for the Environment in making a determination to include the species on the *List of specimens taken to be suitable for live import*.

Plants that are considered weeds are sometimes also considered to have value, for example, for purposes such as ornamental display, traditional medicine, feed for stock, etc. Considerations of the benefits, and therefore any associated concerns about eradication of the target weed species are out of the scope of this analysis.

The Department of Agriculture, Water and the Environment will not commence an assessment to release a biological control agent unless the target has been approved by an appropriate government body. *Jatropha gossypiifolia* was approved by the Standing Committee on Agriculture and Resource Management in 1999.

### Associated pests

There are pests that may arrive with an imported exotic biological control agent. Section 9 of the *Biosecurity Act 2015* defines a pest as ‘a species, strain or biotype of plant or animal, or a disease agent, that has the potential to cause, either directly or indirectly, harm to: human, animal or plant health; or the environment.’ These pests may include, for example, parasitoids, mites or fungi. Should an application to release a biological control agent be approved, these pests will be addressed by existing operational procedures that apply to the importation and final release of the agent. These procedures include detailed examination of imported material, confirmation of identity, and breeding under containment conditions before release. For this reason, associated pests are not further considered in this risk analysis.

### Consultation

In August 2021, a preliminary draft of this report was distributed to state and territory departments of primary industry via the Plant Health Committee and the Commonwealth Scientific and Industrial Research Organisation (CSIRO). Comments were considered and some minor changes were made to the application and draft report as a result. There was no change to the off-target risk estimate of Negligible and no objections to the proposed recommendation to release *Stomphastis* sp. nov. (now identified as *S. thraustica*) for the biological control of the weed *J. gossypiifolia*.

In November 2021, Biosecurity Advice 2021-P13 informed stakeholders of the release of a draft risk analysis report for the release of *Stomphastis* sp. nov. (now identified as *S. thraustica*) for the biological control of the weed *J. gossypiifolia*. The draft report was released for a 30 day stakeholder consultation period that closed on 20 December 2021. Two stakeholder submissions were received during the consultation period. One submission supported the proposed recommendation to permit release of the biological control agent. The second submission did not support release, the response to this submission is detailed in section 5 of this report. It is noted that the second submission was primarily concerned with the lack of a species level identification for the biological control agent. As the species has now been identified as *S. thraustica*, the concerns raised have been resolved.

### Next Steps

Publication of the final report represents the end of the risk analysis process. There is also an approval process within the department under the *Environment Protection and Biodiversity Conservation Act 1999*. Risk analysis reports may be used by the Minister for the Environment in making a determination to include the species on the *List of specimens taken to be suitable for live import* (the Live Import List). This approval process will now take place. If the Department of Agriculture, Water and the Environment approves release of the biological control agent and the Live Import List is amended to include the agent, a letter will be sent to the applicant providing conditions of release.

# Assessment of off-target risks

This section sets out the assessment of off-target risks that could be associated with the release of the biological control agent. Where appropriate, the methods followed those used for pest risk analysis (PRA) by the Department of Agriculture, Water and the Environment in accordance with the International Standards for Phytosanitary Measures (ISPMs), including ISPM 2: *Framework for pest risk analysis* (FAO 2019a), ISPM 3: *Guidelines for the export, shipment, import and release of biological control agents and other beneficial organisms* (FAO 2017) and ISPM 11: *Pest risk analysis for quarantine pests* (FAO 2019c) that have been developed under the World Trade Organization (WTO) Agreement on the Application of Sanitary and Phytosanitary Measures (SPS Agreement) (WTO 1995). The methodology for a commodity-based PRA is provided in Appendix A.

The SPS Agreement defines the concept of an ‘appropriate level of sanitary or phytosanitary protection (ALOP)’ as the level of protection deemed appropriate by the WTO Member establishing a sanitary or phytosanitary measure to protect human, animal or plant life or health within its territory.

Like many other countries, Australia expresses its ALOP in qualitative terms. The ALOP for Australia, which reflects community expectations through government policy, is currently expressed as providing a high level of sanitary or phytosanitary protection aimed at reducing risk to a very low level, but not to zero. The band of cells in Table 2.1, marked ‘very low risk’, represents the upper boundary of the ALOP for Australia.

The risk associated with the release of a biological control agent is a combination of the estimates of likelihood of off-target effects and the potential consequences of any off-target effects. A risk estimation matrix (Table 2.1) is used to combine these estimates.

Table 2.1 Risk estimation matrix.

|  |  |
| --- | --- |
| Likelihood of off-target effects | Consequences of off-target effects |
| Negligible  | Very low | Low  | Moderate | High | Extreme  |
| High  | Negligible risk | Very low risk | Low risk | Moderate risk | High risk | Extreme risk |
| Moderate | Negligible risk | Very low risk | Low risk | Moderate risk | High risk | Extreme risk |
| Low | Negligible risk | Negligible risk | Very low risk | Low risk | Moderate risk | High risk |
| Very low | Negligible risk | Negligible risk | Negligible risk | Very low risk | Low risk | Moderate risk |
| Extremely low | Negligible risk | Negligible risk | Negligible risk | Negligible risk | Very low risk | Low risk |
| Negligible  | Negligible risk | Negligible risk | Negligible risk | Negligible risk | Negligible risk | Very low risk |

###

## Stage 1: Initiation

Initiation commences when an applicant provides a submission proposing the release of a biological control agent. The Department of Agriculture, Water and the Environment will not commence an assessment to release a biological control agent unless the target pest in the submission has been approved as a biological control target by an appropriate government body.

The risk analysis area is defined as all of Australia (excluding its external territories), given that once released there will be no control of spread of the agent other than environmental constraints related to the biology of the organism.

## Stage 2: Risk assessment

This assessment evaluates the likelihood of off-target effects and the potential economic and environmental consequences of any such effects.

The risk assessment is based primarily on consideration of the information provided by the applicant in the application package, including the results of host specificity testing, and current information in the scientific literature, where this is available. Given that the proposal is for deliberate release, the likelihood of entry, establishment and spread is assumed to be certain, and therefore the assessment relates to the host specificity of the proposed agent.

A likelihood is assigned to the estimate of occurrence of off-target effects. Six descriptors are used: high; moderate; low; very low; extremely low; and negligible. Definitions for these descriptors and their indicative ranges are given in Appendix A, Table 1.

### Host specificity testing methodology

The following summarised information regarding host specificity testing has been sourced from the application provided by QDAF (Attachment 1). For further details please refer to the application.

In order to determine whether any non-target species would be at risk from the candidate agent, host specificity tests were conducted with *S.* *thraustica* under quarantine containment conditions in Australia. The applicant conducted host specificity tests on a total of 50 non-target plant species (Attachment 1, Table 5). The standard phylogenetic approach for test list species selection, where closely related species within the target species’ family are tested, was followed (Briese 2005). *Jatropha gossypiifolia* belongs to the subfamily Crotonoideae and family Euphorbiaceae. There are four introduced *Jatropha* species, including the target weed, in Australia and these species were all included in the testing. There are no native *Jatropha* species present in Australia. Representatives from both the family and subfamily, including native and exotic species, were included in testing. Five unrelated plant species were also tested as they are attacked by other closely related *Stomphastis* species.

Non-target plant species were obtained as seeds, cuttings or whole plants from a range of locations across Australia.

Host specificity testing in this application involved several experimental methods. Testing consisted of no-choice tests, paired-choice comparison tests, choice oviposition tests and multiple-choice oviposition tests, in the absence of *J. gossypiifolia*. Paired-choice comparison testing was only carried out on non-target species on which larval development was recorded in no-choice trials (*Jatropha curcas* only).

All testing was carried out in quarantine glasshouses maintained at 30°C and 65% relative humidity during the day and 20°C and 55% relative humidity at night for the length of the project on a 12:12 hour cycle.

#### No-choice tests

Twenty newly emerged unsexed *S.* *thraustica* adult moths were released into a gauze covered cage (45 x 45 x 90cm) containing one potted test plant. Sugar solution was provided for sustenance. Plants were checked periodically for eggs and larval mines, and finally when all adults had died. All test plants with eggs and larval mines were monitored until adults had emerged on the control *J. gossypiifolia* plant. If there was any evidence of live larvae on non-target plants at this stage, the test plants were monitored until no live *S.* *thraustica* were present.

*Camellia sinensis* and *Coffea arabica* were tested at a later date. For these no-choice tests only 10 newly emerged unsexed adults were used.

Ten newly emerged unsexed adults were used for the *J. gossypiifolia* controls. The applicant states that this was so as not to overburden the plants.

Non-target test plant species were subjected to a minimum of 5 replicates (exceptions were *Ricinocarpos pinifolius* and *Euphorbia plumerioides* which were subjected to 3 replicates).

#### Paired-choice comparison tests

Only *J. curcas* was tested using this method as this species was the only test plant species that supported larval development. Ten newly emerged unsexed *S.* *thraustica* adult moths were released into a gauze covered cage (100 x 45 x 90cm) containing one *J. curcas* potted plant and one *J. gossypiifolia* potted plant. Adults were removed after one day and egg numbers on each plant counted. Plants were monitored and the number and duration of each life stage (larvae, adults and pupae) were recorded. Eight replicates were completed.

#### Choice oviposition tests

Choice oviposition tests were carried out on all non-target plant species on which eggs were laid and first instar larvae hatched during no-choice testing. *Jatropha curcas* was not tested during these trials as this species had already been tested using paired-choice comparison tests.

A preliminary trial was conducted with single plants of *Croton verreauxii*, *Baloghia inophylla* and *Aleurites moluccanus* and a single *J. gossypiifolia* plant using 10 newly emerged unsexed adults in a gauze covered cage (100 x 45 x 90cm). Plants were checked weekly and numbers of eggs and mines counted. Five replicates were completed.

Following the preliminary trial, choice oviposition tests were conducted using three newly emerged sexed adult male/female pairs for all non-target plants tested during no-choice trials that had sustained egg laying and first instar larval hatching. Some species tested during no-choice trials were unable to be sourced for the choice oviposition tests. The same methods as the preliminary trial were used however a larger cage size (215 x 140 x 210cm) was used for all species except for *Camellia sinensis* and *Coffea arabica* (100 x 45 x 90cm). The larger cage size provided adult moths more space to facilitate choice in their egg laying.

#### Multiple-choice oviposition tests, in the absence of *J. gossypiifolia*

This test was conducted with six non-target plant species. The species were chosen due to the high egg lay sustained during no-choice testing. Species tested were *Alchornea ilicifolia*, *Antidesma bunis*, *Baloghia inophylla*, *Bridelia exaltata*, *Croton insularis*, and *Omphalea celata*. Three newly emerged sexed adult male/female pairs were used in the larger cage size used in choice oviposition tests. The cage contained a single potted plant of each non-target species. When all adults had died the plants were checked for eggs, and then also checked again several days later. If eggs were found, plants were monitored until no live *S.* *thraustica* were present. Five replicates were completed.

### Host specificity testing results

#### No-choice tests

No-choice testing resulted in eggs being laid on 35 of the 50 non-target plant species tested. Egg hatch occurred on 28 of those 35 plant species. Development through to the adult stage was only recorded on the target plant and *J. curcas*. Damage to leaves caused by larval mining was extensive on both *J. gossypiifolia* and *J. curcas*.

For all other test plants, first instar larvae attempted to feed, resulting in exploratory mines, but were unable to develop and died shortly after emergence. Exploratory mines were small and only affected the surface layers of leaves. No leaf drop or death was recorded as a result of these mines.

It is noted that the standard errors for the mean results for egg lay and hatching were quite variable. This is assumed to be attributed to the variable numbers of female *S.* *thraustica* moths that would have been present during each replicate as the moths used during testing were unsexed.

#### Paired-choice comparison tests

During paired-choice comparison trials, eggs were laid on both *J. gossypiifolia* and *J. curcas*, with no significant difference between the two species. Development of eggs through to the adult stage was also recorded, also resulting in no significant difference between the two species.

#### Choice oviposition tests

During choice oviposition trials, minimal numbers of eggs were laid on non-target plant species. Eggs were consistently laid on the target plant in all replicates for all trials. During preliminary trials an average of 89.9% of eggs were laid on the target plant, which was significantly greater than the percentage of eggs laid on the non-target plant species tested. During all remaining trials, significantly greater quantities of eggs were laid on the target weed than the non-target plants tested. No eggs were laid on the majority of non-target plant species tested. The maximum mean number of eggs recorded on a non-target plant was 6 eggs laid on *Jatropha multifida*.

During all choice oviposition trials there was no larval development on any non-target plant species. There was also minimal first instar feeding recorded. On the non-target plants where exploratory mines were recorded, these were small and only affected the surface layers of leaves. No leaf drop or death was recorded as a result of these mines.

#### Multiple-choice oviposition tests, in the absence of *J. gossypiifolia*

During the multiple-choice oviposition tests, in the absence of *J. gossypiifolia*, no eggs were laid, across all replicates, on any of the non-target plant species tested.

### Comments on host specificity testing

By testing closely related non-target plant species, using the methodology outlined above, and in the application (Attachment 1), the applicant is considered to have satisfactorily assessed the likelihood of off-target effects occurring in the Australian environment.

The reported results of the host specificity testing consistently showed lifecycle completion and damage to the target weed, *J. gossypiifolia* and the closely related species, *J. curcas* by *S.* *thraustica* within all replicates across all tests. The applicant noted that the complete development recorded on *J. curcas* was not unexpected as *S.* *thraustica* was also found on *J. curcas* during field surveys. It is noted that *J. curcas* is also an approved target for biological control in Australia. The other closely related *Jatropha* species tested (*J. podagrica* and *J. multifida*) were unable to support larval development beyond the first instar.

Eggs were laid and exploratory mines were recorded on many non-target test plant species during no-choice testing, however no further development occurred beyond this stage. It is noted that exploratory mines only caused superficial damage and did not impact plant health. Further choice testing showed minimal egg lay on non-target plant species in the presence of the target weed and in larger cage environments where the moths were in less confined conditions with non-target species.

### Likelihood of off-target effects

The likelihood of off-target effects is estimated on the basis of the outcomes of host specificity testing and other relevant information presented in the application (Attachment 1).

Host specificity testing results indicate that *S.* *thraustica* is host specific to two species of *Jatropha*: the target weed, *J. gossypiifolia*, and the closely related weed *J. curcas*. With the exception of *J. curcas*, no other plant species tested are expected to be at risk of off-target impacts from *S.* *thraustica*. As *J. curcas* is expected to sustain damage where it co-occurs with *J. gossypiifolia* in northern Australia, off-target effects will be highly likely to occur.

On the basis of the results of host specificity testing reported in this application, it is concluded that the likelihood of occurrence of off-target effects in Australia is **High**.

### Assessment of potential consequences of off-target effects

The potential consequences of the off-target effects of this biological control agent have been assessed using the same methodology (Appendix A) as used in the import risk analysis process for pests associated with imported fresh produce.

|  |  |
| --- | --- |
| Criterion | Estimate and rationale |
| **Direct** |
| Plant life or health | A—indiscernibleIt is not anticipated that there will be any direct negative consequences of off-target effects on plant life or health. Host specificity testing has demonstrated that *S.* *thraustica* is only expected to impact the target weed and the closely related *J. curcas*. *Jatropha curcas* is a declared introduced weed species in NT and WA and is an approved target for biological control. Therefore, any off-target impacts sustained on this species are expected to be beneficial. |
| Other aspects of the environment | A— indiscernibleThere is no evidence that the introduction of *S. thraustica* would have any negative effects on any other aspects of the environment. Control of bellyache bush (*J. gossypiifolia*) would be beneficial due to the impact of this weed across its range in Australia.  |
| Indirect |
| Eradication, control | A—indiscernible*Stomphastis* *thraustica* is a biological control agent proposed for release for the biological control of *J. gossypiifolia*, a Weed of National Significance (WONS). As the only anticipated off-target impacts are expected to occur on *J. curcas*, also a weed and an approved target for biological control, *S.* *thraustica* is very unlikely to require attempted eradication or control. |
| Domestic trade | A—indiscernible*Stomphastis* *thraustica* is a biological control agent proposed for release for the biological control of *J. gossypiifolia*, a Weed of National Significance (WONS). The results of host specificity testing indicate that the only species likely to be damaged by *S.* *thraustica* is *J. curcas*, also a weed and an approved target for biological control. Therefore, no impacts on domestic trade are anticipated. |
| International trade | A—indiscernible*Stomphastis* *thraustica* is a biological control agent proposed for release for the biological control of *J. gossypiifolia*, a Weed of National Significance (WONS). No off-target impacts are expected to occur on any plant species of significance to international trade. |
| Environmental and non-commercial | A—indiscernible*Jatropha gossypiifolia* and *J. curcas* (the only species anticipated to be impacted by *S.* *thraustica*) are both introduced and invasive weeds in Australia. No indirect environmental or non-commercial impacts are expected if *S.* *thraustica* is successful in reducing the prevalence of these species in the environment. |

Based on this assessment the potential consequences of off-target effects are assessed as: **Negligible**.

### Off-target risk estimate

Unrestricted risk is the result of combining the likelihood of off-target effects with the outcome of potential consequences. Off-target effects and consequences are combined using the risk estimation matrix shown in Table 2.1.

|  |
| --- |
| Risk estimate for *S. thraustica* |
| Likelihood of off-target effects | High |
| Consequences | Negligible |
| Risk | Negligible |

As indicated, the risk estimate for release of *S. thraustica* has been assessed as ‘Negligible’, which achieves the appropriate level of protection (ALOP) for Australia.

# Recommendation on release

The overall risk estimate for release of *S.* *thraustica* has been assessed as Negligible, which achieves the ALOP for Australia. Therefore, it is recommended that this biological control agent be permitted to be released, subject to standard import and release conditions to ensure that the released material is free of other organisms.

This recommendation is made on the basis of the high level of host specificity demonstrated by the biological control agent on *J. gossypiifolia*, and is based on currently available information.

There may be differences in host specificity of *S. thraustica* between African and Peruvian populations, therefore, release in Australia has only been sought for *S. thraustica* from Peru. The current recommendation only applies to *S. thraustica* from Peru.

# Attachment 1

‘Application to release *Stomphastis* *thraustica* (Peru population), an agent for the biological control of *Jatropha gossypiifolia* in Australia’

# Stakeholder responses to draft risk analysis report

This section summarises stakeholder comments raised during consultation on the draft report, and the response. Two submissions were received during the consultation period for the draft risk analysis report. One submission, from the Western Australian Department of Primary Industries and Regional Development (DPIRD) supported the draft recommendation to permit release of the biological control agent. One submission (see below), from Dr Penelope Greenslade, raised some concerns with the proposed recommendations in the draft report.

**Dr Penelope Greenslade submission:**

**“This application is highly competent and thorough. It clearly has cost a considerable sum. So I am amazed that clearly no money has been spent on describing the new species that is being recommended for importation. Never should any species be considered let alone recommended for importation for any reason what so ever if it is undescribed. Also, as far as I can see, there is no suggestion that specimens be deposited in museums for future reference nor is it to be barcoded . So field workers will not with certainty be able to identify any specimens they find. It is one thing to test potential biological control agents in the laboratory but quite another in the field where unpredictable outcomes may occur. Once in a new environment, species can behave in unpredictable ways and there are examples of this from the past. I strongly recommend that this application be rejected until 1. The new species is described by an experienced taxonomist of Lepidoptera; 2. Specimens be bar coded and the code store on Genbank, and 3. Specimens of all life stages be deposited in at least two Australian Museums. Also authorities should be included for all scientific names quoted even to family and genus.”**

Dr Greenslade’s comments are addressed on the basis of advice provided by the applicant (QDAF) as follows:

**Issue 1: New species description by an experienced Lepidopteran taxonomist**

Importing undescribed species into quarantine for host specificity testing and biological studies is not uncommon. Potential agents are often collected from parts of the world where relatively little research has been conducted into local fauna and flora.

For many years Gracillariid expert Dr Jurate De Prins considered this insect to be a species new to science (as reflected in the release application, which was for *Stomphastis* species nova). There have been delays with completing the identification of the insect due to unforeseen circumstances. However advice has now been received from Dr De Prins that, based on the species description, the insect is in fact *Stomphastis thraustica* (Meyrick, 1908).

*Stomphastis thraustica* is known to occur in Africa, and Asia. The Global Taxonomic Database of Gracillariidae (<http://www.gracillariidae.net/>) list the hosts of *Stomphastis thraustica* to be *Jatropha gossypiifolia, J. curcas* and *Microstachys chamalea* (the website also lists *Sebastiana* *chamalea,* which is a synonym of *Microstachys chamalea)*.

According to Dr De Prins, *Stomphastis thraustica* is a morphologically diverse taxon, but the recognised morphological variations are not sufficient to delimit them as distinct species. Samples from Peru (the quarantine colony) are genetically distinct from African populations, but such variations/difference are not sufficient to place them into a new species. The population of *S. thraustica* from Peru is both genetically distinct from other African species and highly host specific, as demonstrated by quarantine host specificity testing.

**Issue 2: Bar-coding of specimens and storage on Genbank**

As per page 7 of the ‘Application to release *Stomphastis* *thraustica*, an agent for the biological control of *Jatropha gossypiifolia* in Australia’, a full genome sequence of this species has been completed and has been submitted to GenBank. The GenBank code is "BankIt2504776 Stomphastis\_sp\_nov\_mitogenome OK376599”. This code is not active online yet.

**Issue 3: Specimens of all life stages should be deposited in at least two Australian museums**

It is a condition of import permits that voucher specimens be submitted to the Australian National Insect Collection (ANIC). Prior to release of biocontrol agents from quarantine, additional voucher specimens will be lodged with ANIC or an Australian State/Territory agricultural collection or herbarium, labelled with the following information:

* Binomial name and authority
* Country of origin
* Host
* Date collected
* Collection location
* Target species
* Source of identification
* Contact officer for biological control program

In this instance it is intended to lodge specimens both with ANIC and the Queensland Government Department of Agriculture and Fisheries Insect Collection.

Appendix A: Method for pest risk analysis

This chapter sets out the method used for the pest risk analysis (PRA) in this report. The Department of Agriculture, Water and the Environment has conducted this PRA in accordance with the International Standards for Phytosanitary Measures (ISPMs), including ISPM 2: *Framework for pest risk analysis* (FAO, 2019a) and ISPM 11: *Pest risk analysis for quarantine pests* (FAO, 2019c) that have been developed under the SPS Agreement (WTO, 1995).

A PRA is ‘the process of evaluating biological or other scientific and economic evidence to determine whether an organism is a pest, whether it should be regulated, and the strength of any phytosanitary measures to be taken against it’ (FAO, 2019b). A pest is ‘any species, strain or biotype of plant, animal, or pathogenic agent injurious to plants or plant products’ (FAO, 2019b). This definition is also applied in the *Biosecurity Act 2015*.

Biosecurity risk consists of two major components: the likelihood of a pest entering, establishing and spreading in Australia from imports; and the consequences should this happen. These two components are combined to give an overall estimate of the risk.

Unrestricted risk is estimated taking into account the existing commercial production practices of the exporting country and that, on arrival in Australia, the department will verify that the consignment received is as described on the commercial documents and its integrity has been maintained.

Restricted risk is estimated with phytosanitary measure(s) applied. A phytosanitary measure is ‘any legislation, regulation or official procedure having the purpose to prevent the introduction and/or spread of quarantine pests, or to limit the economic impact of regulated non-quarantine pests’ (FAO, 2019b).

A glossary of the terms used in the risk analysis is provided at the end of this report.

The PRAs are conducted in the following three consecutive stages: initiation, pest risk assessment and pest risk management.

Stage 1 Initiation

Initiation identifies the pest(s) and pathway(s) that are of quarantine concern and should be considered for risk analysis in relation to the identified PRA area.

For this risk analysis, the ‘PRA area’ is defined as Australia for pests that are absent, or of limited distribution and under official control. For areas with regional freedom from a pest, the ‘PRA area’ may be defined on the basis of a state or territory of Australia or may be defined as a region of Australia consisting of parts of a state or territory or several states or territories.

For pests that had been considered by the department in other risk assessments and for which import conditions already exist, this risk analysis considered the likelihood of entry of pests on the commodity and whether existing policy is adequate to manage the risks associated with its import. Where appropriate, the previous risk assessment was taken into consideration in this risk analysis.

Stage 2 Pest risk assessment

A pest risk assessment (for quarantine pests) is the ‘evaluation of the probability of the introduction and spread of a pest and of the magnitude of the associated potential economic consequences’ (FAO, 2019b).

The following three, consecutive steps were used in pest risk assessment:

#### Pest categorisation

Pest categorisation identifies which of the pests with the potential to be on the commodity are quarantine pests for Australia and require pest risk assessment. A ‘quarantine pest’ is a pest of potential economic importance to the area endangered thereby and not yet present there, or present but not widely distributed and being officially controlled (FAO, 2019b).

The pests identified in Stage 1 were categorised using the following primary elements to identify the quarantine pests for the commodity being assessed:

* identity of the pest
* presence or absence in the PRA area
* regulatory status
* potential for establishment and spread in the PRA area
* potential for economic consequences (including environmental consequences) in the PRA area.

#### Assessment of the probability of entry, establishment and spread

Details of how to assess the ‘probability of entry’, ‘probability of establishment’ and ‘probability of spread’ of a pest are given in ISPM 11 (FAO, 2019c). The SPS Agreement (WTO 1995) uses the term ‘likelihood’ rather than ‘probability’ for these estimates. In qualitative PRAs, the department uses the term ‘likelihood’ for the descriptors it uses for its estimates of likelihood of entry, establishment and spread. The use of the term ‘probability’ is limited to the direct quotation of ISPM definitions.

A summary of this process is given here, followed by a description of the qualitative methodology used in this risk analysis.

##### Likelihood of entry

The likelihood of entry describes the likelihood that a quarantine pest will enter Australia as a result of trade in a given commodity, be distributed in a viable state in the PRA area and subsequently be transferred to a host. It is based on pathway scenarios depicting necessary steps in the sourcing of the commodity for export, its processing, transport and storage, its use in Australia and the generation and disposal of waste. In particular, the ability of the pest to survive is considered for each of these various stages.

The likelihood of entry estimates for the quarantine pests for a commodity are based on the use of the existing commercial production, packaging and shipping practices of the exporting country. Details of the existing commercial production practices for the commodity are set out in the report. These practices are taken into consideration by the department when estimating the likelihood of entry.

For the purpose of considering the likelihood of entry, the department divides this step into two components:

* **Likelihood of importation**—the likelihood that a pest will arrive in Australia when a given commodity is imported.
* **Likelihood of distribution**— the likelihood that the pest will be distributed, as a result of the processing, sale or disposal of the commodity, in the PRA area and subsequently transfer to a susceptible part of a host.

Factors to be considered in the likelihood of importation may include:

* distribution and incidence of the pest in the source area
* occurrence of the pest in a life-stage that would be associated with the commodity
* mode of trade (for example, bulk, packed)
* volume and frequency of movement of the commodity along each pathway
* seasonal timing of imports
* pest management, cultural and commercial procedures applied at the place of origin
* speed of transport and conditions of storage compared with the duration of the lifecycle of the pest
* vulnerability of the life-stages of the pest during transport or storage
* incidence of the pest likely to be associated with a consignment
* commercial procedures (for example, refrigeration) applied to consignments during transport and storage in the country of origin, and during transport to Australia.

Factors to be considered in the likelihood of distribution may include:

* commercial procedures (for example, refrigeration) applied to consignments during distribution in Australia
* dispersal mechanisms of the pest, including vectors, to allow movement from the pathway to a host
* whether the imported commodity is to be sent to a few or many destination points in the PRA area
* proximity of entry, transit and destination points to hosts
* time of year at which import takes place
* intended use of the commodity (for example, for planting, processing or consumption)
* risks from by-products and waste.

##### Likelihood of establishment

Establishment is defined as the ‘perpetuation for the foreseeable future, of a pest within an area after entry’ (FAO, 2019b). In order to estimate the likelihood of establishment of a pest, reliable biological information (for example, lifecycle, host range, epidemiology, survival) is obtained from the areas where the pest currently occurs. The situation in the PRA area can then be compared with that in the areas where it currently occurs and expert judgement used to assess the likelihood of establishment.

Factors to be considered in the likelihood of establishment in the PRA area may include:

* availability of hosts, alternative hosts and vectors
* suitability of the environment
* reproductive strategy and potential for adaptation
* minimum population needed for establishment
* cultural practices and control measures.

##### Likelihood of spread

Spread is defined as ‘the expansion of the geographical distribution of a pest within an area’ (FAO, 2019b). The likelihood of spread considers the factors relevant to the movement of the pest, after establishment on a host plant or plants, to other susceptible host plants of the same or different species in other areas. In order to estimate the likelihood of spread of the pest, reliable biological information is obtained from areas where the pest currently occurs. The situation in the PRA area is then carefully compared with that in the areas where the pest currently occurs and expert judgement used to assess the likelihood of spread.

Factors to be considered in the likelihood of spread may include:

* suitability of the natural and/or managed environment for natural spread of the pest
* presence of natural barriers
* potential for movement with commodities, conveyances or by vectors
* intended use of the commodity
* potential vectors of the pest in the PRA area
* potential natural enemies of the pest in the PRA area.

##### Assigning likelihoods for entry, establishment and spread

Likelihoods are assigned to each step of entry, establishment and spread. Six descriptors are used: high; moderate; low; very low; extremely low; and negligible (Table 1). Definitions for these descriptors and their indicative probability ranges are given in Table 1. The indicative probability ranges are only provided to illustrate the boundaries of the descriptors and are not used beyond this purpose in qualitative PRAs. These indicative probability ranges provide guidance to the risk analyst and promote consistency between different pest risk assessments.

Table 1 Nomenclature of likelihoods

|  |  |  |
| --- | --- | --- |
| Likelihood | Descriptive definition | Indicative range |
| High | The event would be very likely to occur | 0.7 < to ≤ 1 |
| Moderate | The event would occur with an even likelihood | 0.3 < to ≤ 0.7 |
| Low | The event would be unlikely to occur | 0.05 < to ≤ 0.3 |
| Very low | The event would be very unlikely to occur | 0.001 < to ≤ 0.05 |
| Extremely low | The event would be extremely unlikely to occur | 0.000001 < to ≤ 0.001 |
| Negligible | The event would almost certainly not occur | 0 < to ≤ 0.000001 |

##### Combining likelihoods

The likelihood of entry is determined by combining the likelihood that the pest will be imported into the PRA area and the likelihood that the pest will be distributed within the PRA area, using a matrix of rules (Table 2). This matrix is then used to combine the likelihood of entry and the likelihood of establishment, and the likelihood of entry and establishment is then combined with the likelihood of spread to determine the overall likelihood of entry, establishment and spread.

For example, if the likelihood of importation is assigned a descriptor of ‘low’ and the likelihood of distribution is assigned a descriptor of ‘moderate’, then they are combined to give a likelihood of ‘low’ for entry. The likelihood for entry is then combined with the likelihood assigned for establishment of ‘high’ to give a likelihood for entry and establishment of ‘low’. The likelihood for entry and establishment is then combined with the likelihood assigned for spread of ‘very low’ to give the overall likelihood for entry, establishment and spread of ‘very low’. This can be summarised as:

importation x distribution = entry [E] **low x moderate = low**

entry x establishment = [EE] **low x high = low**

[EE] x spread = [EES] **low x very low = very low**

Table 2 Matrix of rules for combining likelihoods

|  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- |
|  | High | Moderate | Low | Very low | Extremely low | Negligible |
| High | High | Moderate | Low | Very low | Extremely low | Negligible |
| Moderate | Low | Low | Very low | Extremely low | Negligible |
| Low | Very low | Very low | Extremely low | Negligible |
| Very low | Extremely low | Extremely low | Negligible |
| Extremely low | Negligible | Negligible |
| Negligible | Negligible |

##### Time and volume of trade

One factor affecting the likelihood of entry is the volume and duration of trade. If all other conditions remain the same, the overall likelihood of entry will increase as time passes and the overall volume of trade increases.

The department normally considers the likelihood of entry on the basis of the estimated volume of one year’s trade. This is a convenient value for the analysis that is relatively easy to estimate and allows for expert consideration of seasonal variations in pest presence, incidence and behaviour to be taken into account. The consideration of the likelihood of entry, establishment and spread and subsequent consequences takes into account events that might happen over a number of years even though only one year’s volume of trade is being considered. This difference reflects biological and ecological facts, for example where a pest or disease may establish in the year of import but spread may take many years.

The use of a one year volume of trade has been taken into account when setting up the matrix that is used to estimate the risk and therefore any policy based on this analysis does not simply apply to one year of trade. Policy decisions that are based on the department’s method that uses the estimated volume of one year’s trade are consistent with Australia’s policy on appropriate level of protection and meet the Australian Government’s requirement for ongoing quarantine protection. If there are substantial changes in the volume and nature of the trade in specific commodities then the department will review the risk analysis and, if necessary, provide updated policy advice.

#### Assessment of potential consequences

The objective of the consequence assessment is to provide a structured and transparent analysis of the potential consequences if the pests or disease agents were to enter, establish and spread in Australia. The assessment considers direct and indirect pest effects and their economic and environmental consequences. The requirements for assessing potential consequences are given in Article 5.3 of the SPS Agreement (WTO, 1995), ISPM 5 (FAO, 2019b) and ISPM 11 (FAO, 2019c).

Direct pest effects are considered in the context of the effects on:

* plant life or health
* other aspects of the environment.

Indirect pest effects are considered in the context of the effects on:

* eradication, control
* domestic trade
* international trade
* non-commercial and environmental.

For each of these six criteria, the consequences were estimated over four geographic levels, defined as:

**Local**—an aggregate of households or enterprises (a rural community, a town or a local government area).

**District**—a geographically or geopolitically associated collection of aggregates (generally a recognised section of a state or territory, such as ‘Far North Queensland’).

**Regional**—a geographically or geopolitically associated collection of districts in a geographic area (generally a state or territory, although there may be exceptions with larger states such as Western Australia).

**National**—Australia wide (Australian mainland states and territories and Tasmania).

For each criterion, the magnitude of the potential consequence at each of these levels was described using four categories, defined as:

**Indiscernible**—pest impact unlikely to be noticeable.

**Minor significance**—expected to lead to a minor increase in mortality/morbidity of hosts or a minor decrease in production but not expected to threaten the economic viability of production. Expected to decrease the value of non-commercial criteria but not threaten the criterion’s intrinsic value. Effects would generally be reversible.

**Significant**—expected to threaten the economic viability of production through a moderate increase in mortality/morbidity of hosts, or a moderate decrease in production. Expected to significantly diminish or threaten the intrinsic value of non-commercial criteria. Effects may not be reversible.

**Major significance**—expected to threaten the economic viability through a large increase in mortality/morbidity of hosts, or a large decrease in production. Expected to severely or irreversibly damage the intrinsic ‘value’ of non-commercial criteria.

The estimates of the magnitude of the potential consequences over the four geographic levels were translated into a qualitative impact score (A‑G) using Table 3. For example, a consequence with a magnitude of ‘significant’ at the ‘district’ level will have a consequence impact score of D.

Table 3 Decision rules for determining the consequence impact score based on the magnitude of consequences at four geographic scales

|  |  |
| --- | --- |
| Magnitude | Geographic scale |
| Local | District | Region | Nation |
| Indiscernible | A | A | A | A |
| Minor significance | B | C | D | E |
| Significant | C | D | E | F |
| Major significance | D | E | F | G |

Note: In earlier qualitative PRAs, the scale for the impact scores went from A to F and did not explicitly allow for the rating ‘indiscernible’ at all four levels. This combination might be applicable for some criteria. In this report, the impact scale of A to F has been changed to become B‑G and a new lowest category A (‘indiscernible’ at all four levels) was added. The rules for combining impacts in Table 4 were adjusted accordingly.

The overall consequence for each pest is achieved by combining the qualitative impact scores (A–G) for each direct and indirect consequence using a series of decision rules (Table 4). These rules are mutually exclusive, and are assessed in numerical order until one applies.

Table 4 Decision rules for determining the overall consequence rating for each pest

|  |  |  |
| --- | --- | --- |
| Rule | The impact scores for consequences of direct and indirect criteria | Overall consequence rating |
| 1 | Any criterion has an impact of ‘G’; ormore than one criterion has an impact of ‘F’; ora single criterion has an impact of ‘F’ and each remaining criterion an ‘E’. | Extreme |
| 2 | A single criterion has an impact of ‘F’; orall criteria have an impact of ‘E’. | High |
| 3 | One or more criteria have an impact of ‘E’; orall criteria have an impact of ‘D’. | Moderate |
| 4 | One or more criteria have an impact of ‘D’; orall criteria have an impact of ‘C’. | Low |
| 5 | One or more criteria have an impact of ‘C’; orall criteria have an impact of ‘B’. | Very Low |
| 6 | One or more but not all criteria have an impact of ‘B’, andall remaining criteria have an impact of ‘A’. | Negligible |

#### Estimation of the unrestricted risk

Once the assessment of the likelihood of entry, establishment and spread and for potential consequences are completed, the unrestricted risk can be determined for each pest or groups of pests. This is determined by using a risk estimation matrix (Table 5) to combine the estimates of the likelihood of entry, establishment and spread and the overall consequences of pest establishment and spread. Therefore, risk is the combination of likelihood and consequence.

When interpreting the risk estimation matrix, note the descriptors for each axis are similar (for example, low, moderate, high) but the vertical axis refers to likelihood and the horizontal axis refers to consequences. Accordingly, a ‘low’ likelihood combined with ‘high’ consequences, is not the same as a ‘high’ likelihood combined with ‘low’ consequences—the matrix is not symmetrical. For example, the former combination would give an unrestricted risk rating of ‘moderate’, whereas, the latter would be rated as a ‘low’ unrestricted risk.

Table 5 Risk estimation matrix

|  |  |
| --- | --- |
| Likelihood of pest entry, establishment and spread | Consequences of pest entry, establishment and spread |
| Negligible  | Very low | Low  | Moderate | High | Extreme  |
| High  | Negligible risk | Very low risk | Low risk | Moderate risk | High risk | Extreme risk |
| Moderate | Negligible risk | Very low risk | Low risk | Moderate risk | High risk | Extreme risk |
| Low | Negligible risk | Negligible risk | Very low risk | Low risk | Moderate risk | High risk |
| Very low | Negligible risk | Negligible risk | Negligible risk | Very low risk | Low risk | Moderate risk |
| Extremely low | Negligible risk | Negligible risk | Negligible risk | Negligible risk | Very low risk | Low risk |
| Negligible  | Negligible risk | Negligible risk | Negligible risk | Negligible risk | Negligible risk | Very low risk |

#### The appropriate level of protection (ALOP) for Australia

The SPS Agreement defines the concept of an ‘appropriate level of sanitary or phytosanitary protection (ALOP)’ as the level of protection deemed appropriate by the WTO Member establishing a sanitary or phytosanitary measure to protect human, animal or plant life or health within its territory.

Like many other countries, Australia expresses its ALOP in qualitative terms. The ALOP for Australia, which reflects community expectations through government policy, is currently expressed as providing a high level of sanitary or phytosanitary protection aimed at reducing risk to a very low level, but not to zero. The band of cells in Table 5 marked ‘very low risk’ represents the upper boundary of the ALOP for Australia.

Stage 3 Pest risk management

Pest risk management describes the process of identifying and implementing phytosanitary measures to manage risks to achieve the ALOP for Australia, while ensuring that any negative effects on trade are minimised.

The conclusions from pest risk assessment are used to decide whether risk management is required and if so, the appropriate measures to be used. Where the unrestricted risk estimate does not achieve the ALOP for Australia, risk management measures are required to reduce this risk to a very low level. The guiding principle for risk management is to manage risk to achieve the ALOP for Australia. The effectiveness of any proposed phytosanitary measures (or combination of measures) is evaluated, using the same approach as used to evaluate the unrestricted risk, to ensure the restricted risk for the relevant pest or pests achieves the ALOP for Australia.

ISPM 11 (FAO, 2019c) provides details on the identification and selection of appropriate risk management options and notes that the choice of measures should be based on their effectiveness in reducing the likelihood of entry of the pest.

Examples given of measures commonly applied to traded commodities include:

* options for consignments—for example, inspection or testing for freedom from pests, prohibition of parts of the host, a pre-entry or post-entry quarantine system, specified conditions on preparation of the consignment, specified treatment of the consignment, restrictions on end-use, distribution and periods of entry of the commodity
* options preventing or reducing infestation in the crop—for example, treatment of the crop, restriction on the composition of a consignment so it is composed of plants belonging to resistant or less susceptible species, harvesting of plants at a certain age or specified time of the year, production in a certification scheme
* options ensuring that the area, place or site of production or crop is free from the pest—for example, pest-free area, pest-free place of production or pest-free production site
* options for other types of pathways—for example, consider natural spread, measures for human travellers and their baggage, cleaning or disinfestations of contaminated machinery
* options within the importing country—for example, surveillance and eradication programs
* prohibition of commodities—if no satisfactory measure can be found.

Risk management measures are identified for each quarantine pest where the level of biosecurity risk does not achieve the ALOP for Australia.

Glossary

| Term or abbreviation | Definition |
| --- | --- |
| Appropriate level of protection (ALOP) | The level of protection deemed appropriate by the Member establishing a sanitary or phytosanitary measure to protect human, animal or plant life or health within its territory (WTO 1995). |
| Appropriate level of protection (ALOP) for Australia | The *Biosecurity Act 2015* defines the appropriate level of protection (or ALOP) for Australia as a high level of sanitary and phytosanitary protection aimed at reducing biosecurity risks to very low, but not to zero. |
| Australian territory | Australian territory as referenced in the *Biosecurity Act 2015* refers to Australia, Christmas Island and Cocos (Keeling) Islands. |
| Biological control agent | A natural enemy, antagonist or competitor, or other organism, used for pest control (FAO 2019b). |
| Biosecurity | The prevention of the entry, establishment or spread of unwanted pests and infectious disease agents to protect human, animal or plant health or life, and the environment. |
| Biosecurity measures | The *Biosecurity Act 2015* defines biosecurity measures as measures to manage any of the following: biosecurity risk, the risk of contagion of a listed human disease, the risk of listed human diseases entering, emerging, establishing themselves or spreading in Australian territory, and biosecurity emergencies and human biosecurity emergencies.  |
| Biosecurity import risk analysis (BIRA) | The *Biosecurity Act 2015* defines a BIRA as an evaluation of the level of biosecurity risk associated with particular goods, or a particular class of goods, that may be imported, or proposed to be imported, into Australian territory, including, if necessary, the identification of conditions that must be met to manage the level of biosecurity risk associated with the goods, or the class of goods, to a level that achieves the ALOP for Australia. The risk analysis process is regulated under legislation. |
| Biosecurity risk | The *Biosecurity Act 2015* refers to biosecurity risk as the likelihood of a disease or pest entering, establishing or spreading in Australian territory, and the potential for the disease or pest causing harm to human, animal or plant health, the environment, economic or community activities.  |
| Control (of a pest) | Suppression, containment or eradication of a pest population (FAO 2019b). |
| The department | The Australian Government Department of Agriculture, Water and the Environment. |
| Endangered area | An area where ecological factors favour the establishment of a pest whose presence in the area will result in economically important loss (FAO 2019b). |
| Endemic | Belonging to, native to, or prevalent in a particular geography, area or environment. |
| Entry (of a pest) | Movement of a pest into an area where it is not yet present, or present but not widely distributed and being officially controlled (FAO 2019b). |
| Establishment (of a pest) | Perpetuation, for the foreseeable future, of a pest within an area after entry (FAO 2019b). |
| Fumigation | A method of pest control that completely fills an area with gaseous pesticides to suffocate or poison the pests within. |
| Genus | A taxonomic category ranking below a family and above a species and generally consisting of a group of species exhibiting similar characteristics. In taxonomic nomenclature the genus name is used, either alone or followed by a Latin adjective or epithet, to form the name of a species. |
| Host | An organism that harbours a parasite, mutual partner, or commensal partner, typically providing nourishment and shelter. |
| Host range | Species capable, under natural conditions, of sustaining a specific pest or other organism (FAO, 2019b). |
| Infection | The internal ‘endophytic’ colonisation of a plant, or plant organ, and is generally associated with the development of disease symptoms as the integrity of cells and/or biological processes are disrupted. |
| Infestation (of a commodity) | Presence in a commodity of a living pest of the plant or plant product concerned. Infestation includes infection (FAO 2019b). |
| Inspection | Official visual examination of plants, plant products or other regulated articles to determine if pests are present or to determine compliance with phytosanitary regulations (FAO 2019b). |
| Interception (of a pest) | The detection of a pest during inspection or testing of an imported consignment (FAO 2019b). |
| International Plant Protection Convention (IPPC) | The IPPC is an international plant health agreement, established in 1952, that aims to protect cultivated and wild plants by preventing the introduction and spread of pests. The IPPC provides an international framework for plant protection that includes developing International Standards for Phytosanitary Measures (ISPMs) for safeguarding plant resources. |
| International Standard for Phytosanitary Measures (ISPM) | An international standard adopted by the Conference of the Food and Agriculture Organization, the Interim Commission on Phytosanitary Measures or the Commission on Phytosanitary Measures, established under the IPPC (FAO 2019b). |
| Introduction (of a pest) | The entry of a pest resulting in its establishment (FAO 2019b). |
| Larva | A juvenile form of animal with indirect development, undergoing metamorphosis (for example, insects or amphibians). |
| National Plant Protection Organization (NPPO) | Official service established by a government to discharge the functions specified by the IPPC (FAO 2019b). |
| Non-regulated risk analysis | Refers to the process for conducting a risk analysis that is not regulated under legislation (Biosecurity import risk analysis guidelines 2016). |
| Nymph | The immature form of some insect species that undergoes incomplete metamorphosis. It is not to be confused with larva, as its overall form is already that of the adult. |
| Pathogen | A biological agent that can cause disease to its host. |
| Pathway | Any means that allows the entry or spread of a pest (FAO 2019b). |
| Pest | Any species, strain or biotype of plant, animal, or pathogenic agent injurious to plants or plant products (FAO 2019b). |
| Pest free area (PFA) | An area in which a specific pest does not occur as demonstrated by scientific evidence and in which, where appropriate, this condition is being officially maintained (FAO 2019b). |
| Pest risk analysis (PRA) | The process of evaluating biological or other scientific and economic evidence to determine whether an organism is a pest, whether it should be regulated, and the strength of any phytosanitary measures to be taken against it (FAO 2019b). |
| Pest risk assessment (for quarantine pests) | Evaluation of the probability of the introduction and spread of a pest and of the magnitude of the associated potential economic consequences (FAO 2019b). |
| Pest risk assessment (for regulated non-quarantine pests) | Evaluation of the probability that a pest in plants for planting affects the intended use of those plants with an economically unacceptable impact (FAO 2019b). |
| Pest risk management (for quarantine pests) | Evaluation and selection of options to reduce the risk of introduction and spread of a pest (FAO 2019b). |
| Pest risk management (for regulated non-quarantine pests) | Evaluation and selection of options to reduce the risk that a pest in plants for planting causes an economically unacceptable impact on the intended use of those plants (FAO 2019b). |
| Pest status (in an area) | Presence or absence, at the present time, of a pest in an area, including where appropriate its distribution, as officially determined using expert judgement on the basis of current and historical pest records and other information (FAO 2019b). |
| Phytosanitary certificate | An official paper document or its official electronic equivalent, consistent with the model of certificates of the IPPC, attesting that a consignment meets phytosanitary import requirements (FAO 2019b). |
| Phytosanitary certification | Use of phytosanitary procedures leading to the issue of a phytosanitary certificate (FAO 2019b). |
| Phytosanitary measure | Phytosanitary relates to the health of plants. Any legislation, regulation or official procedure having the purpose to prevent the introduction and/or spread of quarantine pests, or to limit the economic impact of regulated non-quarantine pests (FAO 2019b). In this risk analysis the term ‘phytosanitary measure’ and ‘risk management measure’ may be used interchangeably.  |
| Phytosanitary procedure | Any official method for implementing phytosanitary measures including the performance of inspections, tests, surveillance or treatments in connection with regulated pests (FAO 2019b). |
| Phytosanitary regulation | Official rule to prevent the introduction and/or spread of quarantine pests, or to limit the economic impact of regulated non-quarantine pests, including establishment of procedures for phytosanitary certification (FAO 2019b). |
| Polyphagous | Feeding on a relatively large number of hosts from different plant family and/or genera. |
| Practically free | Of a consignment, field or place of production, without pests (or a specific pests) in numbers or quantities in excess of those that can be expected to result from, and be consistent with good cultural and handling practices employed in the production and marketing of the commodity (FAO 2019b). |
| Pupa | An inactive life stage that only occurs in insects that undergo complete metamorphosis, for example butterflies and moths (Lepidoptera), beetles (Coleoptera) and bees, wasps and ants (Hymenoptera). |
| Quarantine | Official confinement of regulated articles for observation and research or for further inspection, testing or treatment (FAO 2019b). |
| Quarantine pest | A pest of potential economic importance to the area endangered thereby and not yet present there, or present but not widely distributed and being officially controlled (FAO 2019b). |
| Regulated article | Any plant, plant product, storage place, packaging, conveyance, container, soil and any other organism, object or material capable of harbouring or spreading pests, deemed to require phytosanitary measures, particularly where international transportation is involved (FAO 2019b). |
| Regulated non-quarantine pest | A non-quarantine pest whose presence in plants for planting affects the intended use of those plants with an economically unacceptable impact and which is therefore regulated within the territory of the importing contracting party (FAO 2019b). |
| Regulated pest | A quarantine pest or a regulated non-quarantine pest (FAO 2019b). |
| Restricted risk | Restricted risk is the risk estimate when risk management measures are applied. |
| Risk analysis | Refers to the technical or scientific process for assessing the level of biosecurity risk associated with the goods, or the class of goods, and if necessary, the identification of conditions that must be met to manage the level of biosecurity risk associated with the goods, or class of goods to a level that achieves the ALOP for Australia.  |
| Risk management measure | Are conditions that must be met to manage the level of biosecurity risk associated with the goods or the class of goods, to a level that achieves the ALOP for Australia. In this risk analysis, the term ‘risk management measure’ and ‘phytosanitary measure’ may be used interchangeably. |
| Saprophyte | An organism deriving its nourishment from dead organic matter. |
| Spread (of a pest) | Expansion of the geographical distribution of a pest within an area (FAO 2019b). |
| SPS Agreement | WTO Agreement on the Application of Sanitary and Phytosanitary Measures. |
| Stakeholders | Government agencies, individuals, community or industry groups or organizations, whether in Australia or overseas, including the proponent/applicant for a specific proposal, who have an interest in the policy issues. |
| Surveillance | An official process which collects and records data on pest occurrence or absence by surveying, monitoring or other procedures (FAO 2019b). |
| Systems approach(es) | The integration of different risk management measures, at least two of which act independently, and which cumulatively achieve the appropriate level of protection against regulated pests. |
| Treatment | Official procedure for the killing, inactivation or removal of pests, or for rendering pests infertile or for devitalisation (FAO 2019b). |
| Unrestricted risk | Unrestricted risk estimates apply in the absence of risk management measures. |
| Vector | An organism that does not cause disease itself, but which causes infection by conveying pathogens from one host to another. |
| Viable | Alive, able to germinate or capable of growth. |

References

Briese, D 2005, ‘Translating host-specificity test results into the real world: the need to harmonize the yin and yang of current testing procedures’, *Biological Control*, vol. 35, pp. 208-214.

FAO 2017, *International Standards for Phytosanitary Measures (ISPM) no. 3: Guidelines for the export, shipment, import and release of biological control agents and other beneficial organisms*, Food and Agriculture Organization of the United Nations, Rome, available at [ippc.int/en/core-activities/standards-setting/ispms/](https://www.ippc.int/en/core-activities/standards-setting/ispms/).

FAO 2019a*, International Standards for Phytosanitary Measures (ISPM) no. 2: Framework for pest risk analysis*, Food and Agriculture Organization of the United Nations, Rome, available at [ippc.int/en/core-activities/standards-setting/ispms/](https://www.ippc.int/en/core-activities/standards-setting/ispms/).

FAO 2019b, *International Standards for Phytosanitary Measures (ISPM) no. 5: Glossary of phytosanitary terms*, Food and Agriculture Organization of the United Nations, Rome, available at [ippc.int/en/core-activities/standards-setting/ispms/](https://www.ippc.int/en/core-activities/standards-setting/ispms/).

FAO 2019c, *International Standards for Phytosanitary Measures (ISPM) no. 11: Pest risk analysis for quarantine pests*, Food and Agriculture Organization of the United Nations, Rome, available at [ippc.int/en/core-activities/standards-setting/ispms/](https://www.ippc.int/en/core-activities/standards-setting/ispms/).

WTO 1995, ‘The WTO agreement on the application of sanitary and phytosanitary measures (SPS Agreement)’, World Trade Organisation, Switzerland, available at [wto.org/english/tratop\_e/sps\_e/spsagr\_e.htm](https://www.wto.org/english/tratop_e/sps_e/spsagr_e.htm).