

Australian Government

Department of Agriculture, Fisheries and Forestry

Microbiology of food and animal feeding stuffs – Horizontal method for the detection of *Escherichia coli* O157 – AS 5013.26:2020

SCOPE

This method is applicable to products intended for human consumption or for animal feeding stuffs.

PRINCIPLES

The detection of *E. coli* 0157 necessitates four successive stages:

Enrichment

A test portion¹ is enriched in nine times the weight of pre-warmed modified tryptone soya broth plus novobiocin (mTSB+N) at 41.5 °C \pm 1°C for 6 h and subsequently for a further 12 to 18 h.

Separation and concentration

E. coli 0157 are separated and concentrated using immunomagnetic beads coated with antibodies to *E. coli* 0157 after 6 h² and again, if necessary, after a further 12 to 18 h incubation.

Isolation

Immunomagnetic particles with adhering bacteria are subcultured onto cefixime tellurite sorbitol MacConkey agar (CT-SMAC) and a second selective isolation agar (complementary to CT-SMAC agar) of the laboratories choosing. CT-SMAC is incubated at 37°C for 18 to 24 h. The second agar of choice should be incubated following the manufacturer's recommended procedures.

Confirmation³

Five typical sorbitol negative colonies from CT-SMAC and five typical *E. coli* O157 colonies on the second isolation agar are streaked onto nutrient agar and incubated at 37°C for 18 to 24 h. *E. coli* O157 is confirmed by indole production and agglutination with *E. coli* O157 antiserum.

¹ A larger test portion than that initially validated may be used, if a validation/verification study has shown that there are no adverse effects on the detection of *E. coli* 0157. This method has not been validated at lower dilution ratios, i.e. 375g in 1000 mL.

² In some cases positive results after 6 h incubation can become negative after a further 18 h incubation.

³ Commercially available miniaturized biochemical identification kits that permit the identification of sorbitol-negative and indole-positive *E. coli* and latex agglutination kits for *E. coli* 0157 may be used, provided appropriate tests with known positive and negative strains are carried out to confirm performance

CHECKLIST

Enrichment	Is the sample enriched in mTSB+N?	
	Is mTSB+N pre-warmed to 41.5°C prior to use?	
	Is the correct amount of broth used for the weight of sample analysed i.e. $3,375 \text{ mL}$ for a 375 g sample? Is enrichment at $41.5 \pm 1^{\circ}$ C?	
	Is enrichment carried out at 41.5 ± 1°C incubated for 6 h and then, if necessary, for a further 12 to 18 h?	
	Is a positive control run with each batch of samples analysed?	
	Are reference cultures inoculated into primary enrichment broth at a level of 10 to 100 cells?	
Separation	Is <i>E. coli</i> 0157 separated and concentrated from the enrichment broth using IMS?	
	Is IMS carried out after 6 h and again, if necessary, after a further 12 to 18 h?	
Isolation	Are immunomagnetic beads subcultured onto CT- SMAC and a second selective isolation agar?	
	Are CT-SMAC plates incubated at 37 °C for 18 to 24 h?	
	Is the second isolation agar incubated at the recommended time and temperature?	
Confirmation	Are suspect colonies streaked onto nutrient agar and incubated at 37°C for 18 to 24 h.	
	Is <i>E. coli</i> 0157 confirmed by:	
	Indole production?	
	Agglutination of <i>E. coli</i> 0157 antiserum?	