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Status of the vegetation plots for the spectral library project

K Pfitzner & A Bollhöfer

December 2008

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Supervising Scientist Division GPO Box 461, Darwin NT 0801

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Status of the vegetation plots for the spectral library project

K Pfitzner & A Bollhöfer

1 Introduction

This paper summarises the status of the vegetati on plots that are used to measu re the spectral reflectance of vegetative ground covers for the spectral library project (File SG2005/0075). The plots are contained within three sites located at research facilities in the greater Darwin region: Berrimah Farm, CSIRO and Crocody lus Park. A brief introduction that outlines the project aims, limitations and achievements is provided below. The main purpose of the report is to summarise the 2006 and 2007 activities for each site. The species measured and frequency of data collection for each site is listed. The status of the plots as of January 2008 is given.

Since this manuscript was prepared, the spect ral sam pling of vegetative plots has been discontinued due to resource constraints.

1.1 Project overview

The aim of the spectral library project is to develop a spectral reflectance database of land covers pertinent to rem ote sensing applications in the mine environment. The focus was on vegetation components, with priority species being weeds and native ground covers that are of concern to the revegetation success at minesites. Plots of home ogenous covers wer e established at Berrimah Farm , CSIRO and Croc odylus Park. A field spe ctrometer th at measures reflectance continuously across the 0.35 -2.50 µm wavelength range is used to obtain the spectral data. Meteorological data, measurement metadata and cover descriptions are obtained with the s pectral measurements. W here possible, sa mpling at fortnightly intervals is undertaken. The data are expected to provide insight into the variations between and within plant species during diff ering environmental conditions and over tem poral variations in spectral signatures. Subtle phe nological spectral c hanges between and within species can be used to make re commendations on the use of rem ote sensing for m inesite assessment studies with applicability to all environments. The hypothesis is that with a well designed approach to collecting field spect ral measurements and metadata, potenti extraneous factors can be spectrally accounted for and accurate post-processing of spectra can be performed. Thus the first validated database of spectra relevant to the mine environment in the Top End can be developed. Further project details can be found in Pfitzner et al (2006) and Pfitzner et al (2009).

1.2 Project Innovation

Potential exists for remote monitoring of ground cover type and condition over minesites and their surrounds. Quantitative inform ation on ground cover, such as percent species cover (including weed cover) and derived information such as fuel load, require that species c an be discriminated and identified. Plant spect ral data can potentially be used for this purpose but data on the spectral response of ground covers is limited.

This project addresses the following research questions:

- What are the temporal spectral responses of ground cover vegetative species?
- Can ground- cover vegetative species be di stinguished using ground based reflectance spectra, and if so, what spectral resolution is required?
- At what phenological stage is maximum separability detected and is there a growth or seasonal stage when plant species cannot be distinguished?
- What are the implications for remote sensing throughout the year?

Many remote sensing applications will remain in the research realm without a ground-based knowledge base to define species separability over time.

1.3 Known project limitations and project achievements

To achieve the required f ortnightly fre quency of spectral measurements, long travel times were not feasible and site selection was therefore restricted to the greater Darwin region, and limited to a few sites. The sites were chosen because they were located and supported within research facilities and unlikely to become disturbed or inaccessible. The species available for sampling were dependant on these constraints, and limitations in the project design are acknowledged, particularly with respect to the number of species sampled and replication of the sampled species.

Suitable sites for additional speci es were continually being sourced to discount potential confounding factors. Species replication was considered as important as species variety although most species were not represented more than once. Ideally, replicates of species were sampled at different locations.

Considering that a vegetat ive species spectral response is a function of a variety of factors ranging from soil type and soil condition to local meteorological conditions, a further known limitation was the vary ing growth media and environmental conditions prevailing during sample collection. The spectral response can also be expected to vary over scale s in time (diurnal to se asonal) and space. It is acknowledged that the spectral response measured is a function only of a point in time for a particular vegetation sample. To minimise and account for external variation, species were sampled from maintained plots that maximise a homogenous response (ie non-target species were continually weeded out). The soil interspaces was spectrally measured wherever a less than 100% cover was real ised. Detailed metadata was used to define any change in localised conditions both within the target plant and for external condition.

Much focus to date has been on the developm—ent of protocols including accurate—metadata recording so that data are comparable over time and so the data has long term—value and is suitable for new applications. Detail of these protocols can be found in Pfitzner et al (2006) and Pfitzner et al (2009). Despite known project limitations, the project is expected to provide a knowledge base far greater than that ever obtaained for vegetation spectra—with respect to species numbers, frequency of sam pling, duration of sampling and method and metadata documentation. The envisaged outcome of the project was—a spectral library of vegetation reflectances pectra, which can be used to—determine the timing and frequency—for spectral measurements of vegetation endmembers. Despite vegetative sampling being discontinued, the data collected will go some way in addressing the project's aim.

2 Berrimah Farm

2.1 Summary of 2006 activities – Berrimah Farm

There were five plots est ablished at Berrimah Farm in 2006. Four species of agricult ural grasses are represented:

BF01 Digitaria milanjiana (Jarra Grass)
 BF02 Brachiaria humidicola (Tully Grass)
 BF03 Brachiaria humidicola (Tully Grass)
 BF04 Digitaria swynnertonii (Arnhem Grass)
 BF05 Digitaria eriantha (Pangola Grass)

While these grasses are not a high priority in terms of threat to mine rehabilitation success, these plots were established opportunistically as they represent dense and homogenous stands of introduced pastoral grasses that are also apparent in the Alligator Rivers Region (ARR). The five plots are easily accessible and can be sampled in one morning on a cloud-free day. Plots BF01 and BF02 are at a distance of less than 10 m, and plots BF03 and BF04 are within 20 m distance of each other. The distance between BF1/BF2 and BF3/BF4 is over 300 m. BF5 is approximately 200 m from BF1, BF2, BF3 and BF4.

Five complete replicates of spectral samples and metadata were taken at Berrimah Farm between September and December 2006 (Table 1). Spectral sampling was undertaken on September 26, October 9, October 30, November 15, November 30 and December 01. Where environmental conditions changed during sampling (eg winds picked up), the site or number of sites that were not same pled on that day were sampled at the next possibele opportunity (eg see Table 1, where sites BF03–05 were not sampled on the 30/11/06, but were sampled on the 01/12/06). In November 2006 it was noted that the plots would need securing as cowswere in the paddock.

Table 1 Spectral and metadata sample collection 2006 - Berrimah Farm

			Berrin	nah Farr	n samp	le
	Date	1	2 3		4 5	i
S	26/09/06	~	~	~	~	~
2006 sampling dates	09/10/06	•	•	•	•	•
ing	30/10/06	•	•	~	•	•
ldme	15/11/06	•	•	•	•	•
)6 S	30/11/06	•	•			
20	01/12/06			•	•	•

2.2 Summary of 2007 activities – Berrimah Farm

There were no changes to the number of plots or species sampled at Berrimah Farm during 2007. Sampling of the five plots established at Berrimah Farm in 200 6 was continued into 2007.

The Berrimah Farm plots were sam pled 12 tim es in 20 07 o ver 15 fieldtri ps (Table 2). Sampling did not occur in 2007 until April (as the spectrometer was overseas for repair).

Sampling was undertaken on: April 11 and 13, April 23, Ma y 10, May 22 and 23, Ju ne 12, July 1 8, Au gust 1, Sept ember 6, Septem ber 21, October 4, October 15 and 17 and November 26.

Table 2 Spectral and metadata sample collection 2006 – Berrimah Farm

			Berrim	ah Fari	m samp	le
	Date	1	2 3		4 5	
	11/04/07	~	~	~	~	
	13/04/07					•
	23/04/07	•	•	•	•	•
	10/05/07	•	•	•	•	•
	22/05/07	~	~			
ates	23/05/07			•	•	•
2007 sampling dates	12/06/07	~	•	•	•	•
ıldı	18/07/07	~	•	•	•	•
7 sar	01/08/07	~	•	•	•	•
2007	06/09/07	~	•	•	•	•
	21/09/07	•	•	•	•	•
	04/10/07	~	~	~	•	
	15/10/07	~	•			
	17/10/07			•	•	
	26/11/07	~	~	~	~	•

Appendix 1 highlights the frequency of spectral samples taken at Berrimah Farm. The data is displayed by weeks and shows that since the establishment of these plots in 2 006, the five plots have been sampled regularly, although not always fortnightly. There was a five week gap where no data were recorded during June–July 2007. No data were recorded in August 2007 and the re was a six week gap in sampling during October and November. In addition, sampling was undertaken every three weeks rather than every two weeks on a number of occasions.

2.3 Status of plots – Berrimah Farm

In May 2 007, warning was given that site s containing BF01, BF02 and BF03 would be grazed. By June 2007, BF01 and BF02 were completely grazed by cows. Plate 1 shows the status of plots as of Nov ember 2007. BF01 and BF02 had been fenced securely by late November 2007. The effect of grazing can be seen on BF02 in Plate 1. BF03 and BF04 were still only pegged with flagging tape and BF05 was fenced with chicken wire only. There was some mixing of grass species within these plots. Rains in late November had initiated a green flush (compare BF01–BF04 from late November with BF05 from early November in Plate 1)





BF1_2007_11-26 BF2_2007_







BF3_2007_11-26 BF4_2007_

11-26



BF_5_2007_11_01 (latest November image available)

Plate 1 Status of plots at Berrimah Farm (November 2007)

Plate 2 (below) illustrates the status of the Be rrimah Farm plots as of December 2007. BF01, BF02, BF04 and BF05 had been fenced securely, however, BF03 still required secure fencing to be erected. BF05 shows some loss of homogeneity around the fenced perimeter.





12-17





BF3_2007_12-17 BF4_2007_

12-17



BF_5_2007_12-17

Plate 2 Status of plots at Berrimah Farm (December 2007)

2.4 Outlook for 2008 - Berrimah Farm

Once BF03 is securely fenced, there is the need for regular maintenance of the plots. This includes checking that fencing has not been damaged by cows and ensuring that any alien species are weeded out. Because it has been realised that Berrimah Farm is affected by winds in the afternoon, spectral sampling should be undertaken in the early morning to maximise the ability to capture the five plots at the same frequency. Spectral sampling should be undertaken at a fortnightly interval for 2008 to o btain a complete seasonal cycle of information (as data was lost in 2007 due to inadequate fencing and grazing cows).

The five species that are ready for sampling in 2008 are:

BF01 Digitaria milanjiana (Jarra Grass)

BF02 Brachiaria humidicola (Tully Grass)

BF03 Brachiaria humidicola (Tully Grass)

BF04 Digitaria swynnertonii (Arnhem Grass)

BF05 Digitaria eriantha (Pangola Grass)

One new site needs to be securely fenced a nd maintained as soon as possible to enable a complete seasonal cycle of information:

BF06 Hyparrhenia rufa (Thatch Grass)

Note that since this manuscript was pr epared, the spectral sa mpling of vegetative plots has been discontinued due to resource constraints.

3 CSIRO

3.1 Summary of 2006 activities - CSIRO

Nine plots were established at CSIRO in 2005 and six plots were maintained and sampled at CSIRO in 2006:

- CS01 Stylosanthes humilis (Townsville Stylo)
- CS02 Sorghum stipodeum (Sorghum)
- CS05 Pennisetum polystachion (Perennial Mission Grass)
- CS06 Passiflora foetida (Wild Passionfruit)
- CS07 Pennisetum pedicellatum (Annual Mission Grass)
- CS09 Urochola maxima (Guinea Grass or Green Panic)

Replicate pl ots of *S. humilis* and *S. stipodeum* were estab lished (CS03 and CS04 , respectively) in 2005 but found to not be feasible for spectral sampling in 2006 due to canopy cover casting shade over the plots in the dry season. The six species all represent important covers for minesite rehabilitation assessment in the ARR. The plot of CS05 is represented by a few individual grass clumps, and CS06 covers only a small part of the plot. The other covers are quite dense and hom ogenous when maintained. The sites are all located within walking distance, although CS01 and CS02 are over 100 m from the other sites.

Nine replicat es of spectra 1 and m etadata sa mples were taken f or six species at CSIR O between July and Decemb er 2006. A li mited number of data were acquired between January and June 2006 while the sampling protocols were being developed. Once the protocols were established, spectral sampling was undertaken o n Jul y 3, August 30, September 12, September 27, October 10, October 25, November 6, November 7, November 20 and November 29. Spectral sampling was also unde rtaken on Augu st 4 but no metadata were recorded.

Table 3 Spectral and metadata sample collection 2006 - CSIRO

				CSIR) sampl	е	
	Date	1	2 5		6 7		9
	03/07/06	~	~	~		~	~
	30/08/06	~	•	•	•	~	•
ites	12/09/06	~	•	•	~	~	~
2006 sampling dates	27/09/06	•	~	•	•	~	•
nplir	10/10/06	•	~	•	•	~	•
san	25/10/06	~	•	•	•	~	•
2006	06/11/06	•		•		~	•
	07/11/06		•		•		
	20/11/06	~	•	•	•	~	•
	29/11/06	~	~	~	~	~	~

3.2 Summary of 2007 activities - CSIRO

The six plots established at CSIRO in 2006 continued to be sampled in 2007 (CS01 S. humilis or Townsville Sty lo, CS02 S. stipodeum or sorg hum, CS05 P. polystachion or perennial

mission grass, CS06 *P. foetida* or wild passionfruit, CS07 *P. pedicellatum* or annual mission grass and CS09 *U. maxima* or Guinea grass).

Two new plots were established at CSIRO during 2007:

CS10 Brachiaria humidicola (Tully Grass)

CS12 Melinis repens (Red Natal Grass)

CS11, *Sorghum stipodeum*, was established, but the cover dis appeared and the site was discontinued. CS11 was sampled once in 2007 on April 26.

The CSIRO plots were sampled around 8 times in 2007 over 11 field trips, although not all species were sampled on every trip. Sampling was undertaken on April 12 and 26, May 11, May 22, June 13, July 11, August 7 and 8, September 7 and 10 and October 16 (Table 4). Spectra were also recorded on July 18 and November 5, but no metadata details were recorded.

Table 4 Spectral and metadata sample collection 2007 - CSIRO

					CSIRO	sample	!		
	Date	1	25		67		9	10	12
	12/04/07	~		~	~		~	~	
	26/04/07	•	•	•	•		•	~	~
	11/05/07	•		•		•	•	~	
2007 sampling dates	22/05/07	•		•		•	•	~	•
p gu	13/06/07	•	~	•		•	•	~	•
mpli	11/07/07	•	~	•	•	•	•	•	•
7 saı	07/08/07			•	•	•	•	•	
200	08/08/07	•	~						•
	07/09/07	•	•	•	•	•	•	•	
	10/09/07								•
	16/10/07	~						~	

Appendix 1 highlights the frequency of spectra 1 sam ples taken at CSIRO. The data is displayed by weeks and shows that since the establishment of these plots in 2006, the six plots have been sampled regularly, although not always fortnightly. In 2006, the sites were sampled fortnightly. From April 2007 – November 2007, the sites were often sampled monthly.

3.3 Status of plots - CSIRO

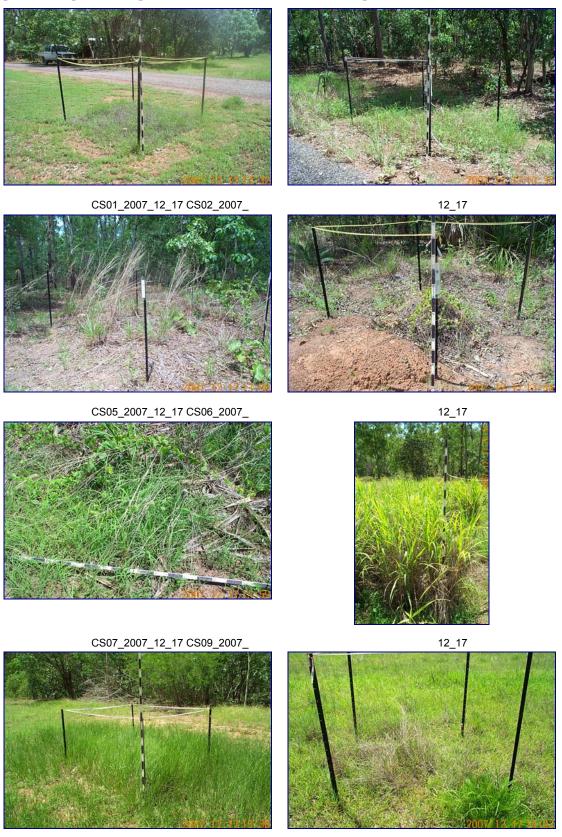
Plates 3 and 4 show the status of the CSIRO plots for early November and December (2007), respectively. CS01, 02, 05, 06, 10 and 12 have been pegged with flagging tape. CS07 and CS09 have not been plotted out (but ur gently need to be defined). There has been an obvious flush of vegetation with rains between early November and December (compare Plates 3 and 4). All plots but CS01 (*S. humilis*) and CS10 (*B. humidicola*) show a patchy cover (Plate 3). In particular, CS05 (*P. Polystachion*) and CS06 (*P. foetida*) are quite sparse and a decision was made to transplant extra individuals into these plots.



Plate 3 Status of plots at CSIRO (November 2007)

Plate 4 shows the CSIRO plots in December 2007 and illustrate a flush in cover compared to Plate 3. CS0 5 and CS06 still require n ew seedlings to be transplanted into these plots in an attempt to achieve a denser cover and CS07 and CS 09 still need to be plotted out. Some sites

need to be maintained with flaggi ng tape and alien species weeded out. The sites require site identification (SSD sa mple tags are att ached to inform new comers to the purpose of these plots). No spectral samples were taken after 19/10/2007 to capture the flush evident in Plate 4.



CS10_2007_12_17 CS12_11_02

Plate 4 Status of plots at CSIRO (December 2007)

3.4 Outlook for 2008 - CSIRO

Maintenance for the CSIRO plots needs to be c ontinued. In particular, alien species to the plots need to be regularly weeded out. The plots require aesthetic maintenance, including neat flagging tape and waterproof site labels to ensure the purpose of the plots is clear. The eight species that will continue into 2008 are:

CS01 Stylosanthes humilis (Townsville Stylo)
CS02 Sorghum stipodeum (Sorghum)
CS05 Pennisetum polystachion (Perennial Mission Grass)
CS06 Passiflora foetida (Wild Passionfruit)
CS07 Pennisetum pedicellatum (Annual Mission Grass)
CS09 Urochola maxima (Guinea Grass or Green Panic)
CS10 Brachiaria humidicola (Tully Grass)

Melinis repens (Red Natal Grass)

Because CS05 and CS06 are sparse in cover, replicate plots, preferably at Crocody lus Park, need to be identified and plotted. Wild passi onfruit is invading CS09, so this is a potential source to increase the cover of CS06. Alternatively, new dense covered sites could be established, but sampling of CS05 and CS06 should continue so that a complete annual data record is obtained.

There is pote ntial for CS07 to be expanded into two plots, and while these plots would be adjacent to one another, extra spectra on this important species would be obtained and with little extra sampling effort required.

During flowering, confirmation of the species composition of CS09 with the NT Herbarium is still required. CS09 needs to be plotted out and any alien species weeded out.

Note that since this manuscript was pr epared, the spectral sa mpling of vegetative plots has been discontinued due to resource constraints.

4 Crocodylus Park

CS12

4.1 Summary of 2006 activities – Crocodylus Park

There were 21 plots established at Crocodylus Park in 2006:

CP01 *Hyptis suaveolens* (Hyptis) CP02 Stylosanthes humilis (Townsville Stylo) CP03 *Hyptis suaveolens* (Hyptis) CP04 Pennisetum pedicellatum (Annual Mission Grass) CP05 Calopogonium mucunoides (Calopo vine) CP06 *Urochloa mutica* (Para grass) CP07 Stachytarpheta cayennensis (Snake weed – white flowers) CP08 Sida cordifolia (Flannel weed) CP09 Stachytarpheta australis (Snake weed – purple flowers) CP10 Panicum mindanese

- CP11 Panicum mindanese
- CP12 Chloris inflata (Purple top chloris)
- CP13 Cynodon dactylon (Couch)
- CP14 Heteropogon contortus
- CP15 Schizachyrium sp
- CP16 Crotalaria goreensis (Gambia Pea)
- CP17 Stylosanthes hamata (Caribbean stylo)
- CP18 Senna sp (Sicklepod)
- CP19 *Lemna* sp (Duckweed)
- CP20 Stachytarpheta cayennensis (Snake weed white flowers)
- CP21 Andropogon gayanus (Gamba grass)

Combined with the Berrimah Far m and CSIRO plots, the species mix established at Crocodylus Park incorporated many of the species important for m inesite rehabilitation studies. All species except CP05, CP06, CP07, CP09, CP19 and CP20 were located in the upper western paddock of Crocodylus Park. Species in plots CP05, CP06, CP07, CP09, CP19 and CP20 were often collected on one sam pling trip (Table 5), as the equip ment had to be packed and unpacked to sample these sites. The majority of plots were not sampled until late August/early September. The data collection appears patchy in Table 5, but this is a function of the two different trips required to capture the spectral data. Appendix 3 displays the data by weeks (data collected during the same week is combined into the one row), illustrating that fortnightly data was mostly collected.

Limited data was collect ed April 5, 18 and 24 and May 18 and 24. More intens ive sampling was undertaken on the following dates: June 08, June 21, June 23, July 04, July 21, August 02, August 31, September 01, September 13, September 14, September 28, September 29, October 12, October 13, October 26, October 31, November 01, November 03, November 13, November 14, November 15 November 27, November 28, November 30 and December 01 (Table 5).

Species in plots CP05, CP06, CP07, CP09, CP19 and CP20 were located away from the remaining plots and often sampled in isolati on to the remaining plots in the upper western paddock. The data sampling appears patchier because those species in the upper western paddock were captured at different dates to those plots external from the paddock.

Table 5 Spectral and metadata sample collection 2006 – Crocodylus Park

									Cro	cody	lus P	ark s	ample)							
Date	1 2		3 4	5		6 7	8		9 10)	11	12	13	14	15	16	17	18	19	20	21
05/04/06	~																				
18/04/06	~																				
24/04/06					•																
18/05/06		~																			
24/05/06						•			•												
08/06/06	~	~	•	•																	
21/06/06	~	~	•	•																	
23/06/06						•															
04/07/06	•	~	•	•																	
21/07/06	•																				
02/08/06																•					
31/08/06		~			~	•	•		•			•							•	•	
01/09/06	•		•	~						•	•		•	•		•	•	•			
13/09/06		~	•	•				•				•	•	•							
14/09/06	•				~	•			•	•	•					•	•		•	•	
28/09/06		~	•	•				•		•		•	•				•	•			
29/09/06	•				•	•			•		•								•	•	
12/10/06			•	•				•		•		•	•	•		•	•	•			
13/10/06					~	•			•		•								•	•	
26/10/06		~	•	•				•													
31/10/06					•				•											•	
01/11/06		~	•					•				•									
03/11/06	~									•			•								
13/11/06					~	•			•										•	•	
14/11/06		•	•	~				•				•	•	•				•			
15/11/06	•						•			~	•					•	•				
27/11/06		•		•				•				•		•				•			
28/11/06	•					•	•			•	•						•		•		
30/11/06					•		•														
01/12/06																					,

4.2 Summary of 2007 activities – Crocodylus Park

Six plots of the 21 plots established at Crocodylus Park in 2006 continued to be sampled from April 2007 (CP02 S. humilis, CP03 H. suaveolens, CP04 P. pedicellatum, CP12 C. inflata, CP13 C. dactylon and CP14 H. contortus).

Some plots were not fenced securely and either donkeys destroyed these plots or the samples were collected until the s pecimen sen esced and the location of the plot became unknown: CP22 (*H. suaveolens* or Hypt is), CP 23 (Morning Glory), CP 24 (*C. mucunoides* or Calopo vin e), CP25 (*Digitara bicornis*), CP 26 (*Aeschynomene americana* or Joint vetch), CP27 (*Spermacoe leptoloba*

or purple sed ge), CP 28 (*Setaria apiculata*), C 29 (*Cyperus Rotundus*), CP 30 (*P. foetia* or wild passionfruit vine) CP 31 (*S. australis*), CP32 (*S. cayennensis*) and CP33 (*Sida cordifolia*).

To rationalise time spent in the field, a d ecision was made to abandon those sites not located within the western top paddock because an extra day of sam pling had been spent at times during 2006 to capture those sites. Those sites were: CP05 (*C. mucunoides* or Calopo vine), CP06 (*U. mutica* or Para grass), CP07 and CP20 (*S. cayennensis* or Snake weed - white flowers), CP09 (*S. australis* or snake weed – purple flowers) and CP19 (*Lemna* sp. Duckweed). *C. mucunoides*, *S. cayennensis* and *S. australis* could be sampled from the top paddock and new sites were identified (CP 24, CP32 and CP33 respectively). CP24 was discontinued, but two new sites were identified which were characterised as a single shrub of:

CP31 Stachytarpheta cayennensis

CP32 Stachytarpheta australis

CP06 had not been an ideal patch of *U. mutica*, as the site was difficult to access and had been grazed by donkeys. Because *U. mutica* is considered a priority species, an easily accessible site was identified along a drain near Palmerston. The advantages of the Palmerston site were that the para grass patch was large and would allow a number of samples to be collected. In addition, there was a gradient in soil moisture away from the drain, which would allow spatiotemporal samples to be collected. Unfortunately this site was not sampled at all in 2007 and no spectra of Para grass were obtained.

In early 2007 a number of potential new plots we re identified in the western top paddock at Crocodylus Park. These included native grasses (*Bothriochloa bladhii*, *Digitaria bicornis*, *Setaria apiculata*), introduced grass es, sedge and cypress (*A. gayanus*, *Cyperus rotundus*, *Dactyloctenium aegyptium*, *Melinus repens*, *Spermacoe leptoloba* or purple sedg e, *Sporobolus jacquemontii*, *Urochloa pubigera*), native shrub (*Euphorbia heterophylla*), introduced shrubs (*Aeschynomene americana* or Joint vetch, *Desmodium tortuosum*, *Hibiscus sabdariffa*, *H. suaveolens*, *Indigofera tinctoria*, *S. australis*, *S. cayennensis*, *Scroparia dulcis*, *Sida Cordifolia*, *Spermacoce leptoloba*), native vine (*Ipomoea pes-tigridis*) and introduced vines (*C. mucunoides* or Calopo vine, *P. foetia* or wild passionfruit vi ne). Opportunistic readings (frequency between 1–3) were taken for a further 6 samples which were of isolated grass clumps or shrubs. These included:

CP21 A. gayanus (Gamba grass clump)

CP23 potted example of Morning Glory

CP25 Digitara bicornis

CP26 Aeschynomene americana or American Joint Vetch

CP33 Sida cordifolia

CP35 Hibiscus sabdariffa (or Rosella).

No complete sets of data were recorded for 2007 and there are large gaps (u p to 6 weeks) between data collection events (see Appendix 3).

A number of Crocody lus Park plots were sa mpled in 2007 over 18 field trips (Table 6). Sampling was undertaken on April 18,19 and 20, May 15, 17, 18, 30 and 31, June 14, July 23 and 25, August 9, 10 and 18, September 5 and 19, October 15 and 31.

Table 6 Spectral and metadata sample collection 2007 - Crocodylus Park

							Crocody	lus P	ark s	ample					
	Date	2	3	4	12	13 1	4 21 23			25 26	3	31 3	2 33 35	5	
	18/04/07	~			~										
	19/04/07			~		~	•							~	
	20/04/07		~						•	•	•		~		
	15/05/07	•	~	•	~										~
	17/05/07					~	•					~	~		
	18/05/07					~					•			~	
	30/05/07	•		•	~	~	•								
	31/05/07		~									~	~		~
Trip 2007	14/06/07	•			~	~	•					~	~		
Trip	23/07/07	•			~										
•	25/07/07			~		•	•					~			
	09/08/07	•		•	~							~			
	10/08/07					•	•								
	18/08/07	•			~	•	~								
	05/09/07				~	~	•						•		
	19/09/07	•		•	•	•	~	•				•			
	15/10/07	•		•	•	~	•						~		
	31/10/07					~	•								

CP23 – a pot of morning glory, CP25 Digitara, CP26 Joint Vetch, CP33 Sida Cordifolia, CP35 – Rosella

CPO2, 03, 04, 12, 13, 14 and 31 were the only samples collected regularly in 2007 (Stylosanthes humilis, Hyptis suaveolens, Pennisetum pedicellatum, Chloris inflate, Cynodon dactylon, Heteropogon contortus and Stachytarpheta sp, respectively).

4.3 Status of plots - Crocodylus Park

Plate 5 shows the status of the plots at Crocodylus Park as of early Novem ber 2007. Only eight plots remained, with six original plots (CP02, CP03, CP04, CP12, CP13, CP14) and two new plots with individual shrubs of snakeweed (CP31 and 32).

CP02, a dense and homogenous patch of *S. humilis*, with a good perimeter, makes for a very suitable plot for tem poral sampling. The plot required a more sturdy fence, as chicken wire has proven inadequate for keeping out grazing donkeys.

CP03, CP04, CP12, CP13, CP14, CP31 and CP32 all required secure fencing to be erected so that the donkeys do not graze the remaining plots. The snakeweed specimens were being measured but had not been fenced out (Plate 5). CP21 (Gam ba grass) had been burnt, but a secure plot was required and a replicate plot of this priority species was to be established nearby.



Plate 5 Status of plots at Crocodylus Park (November 2007)

The rains had caused a flush and CP0 3 appeared dense and ho mogenous, although it was noted that some alien species needed to be removed as soon as possible to avoid an y competition. In December 2007, five species had been contained in one large fenced plot (Plate 6).



Plate 6 Image of the larger plot containing CP03 (closest green dense patch) and CP04 (background dense green patch) as well as new identified species (Rattlepod, American Joint Vetch and Gambia Pea)

Plate 7 shows the status of the plots at Crocodylus Park as of December 2007. CP02 has been securely fenced (note the perimeter to the Sty lo within the plot has flushed and this will require frequent maintenance). Referring to Plate 7, the image r epresenting CP03 shows a dense patch of hyptis that is part of this larger plot. The larger fence was convenient to erect and will make access into these plots for weed ing easier. However, concern is that these 'plots' within the larger fenced plot will easily become merged and mixed and thus frequent maintenance and weeding of this plot is indispensable.

CP04 is also contained within the larger plot. Concern is that until flowering, the grass is unidentifiable and keeping this patch hom ogenous will be difficault (note the other sterile grasses next to the 'plot' of mission grass). The chicken wire fence marking the plot of CP12 was taken do wn in Novem ber and the location of the chloris grass lost. CP1 3 (couch) still requires that alien species be eliminated so that the couch is not out competed before the rains. CP 14 has been securely fenced and makes for a good plot for temporal sampling. CP31 and CP32 have been fenced within the one secure plot.

Note that the fencing of CP12 was taken down and the sample 'lost' between November and December.

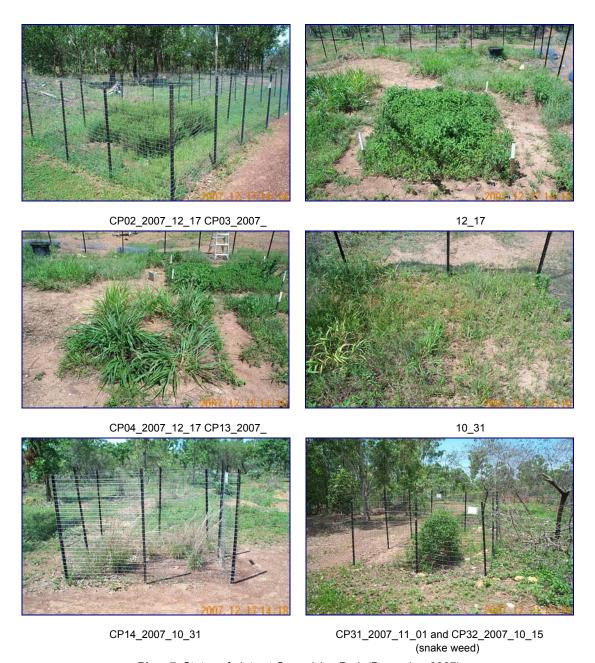


Plate 7 Status of plots at Crocodylus Park (December 2007)

4.4 Outlook for 2008 - Crocodylus Park

Regular maintenance is required for the remaining Crocodylus Park plots. In particular, alien species to the plots need to be regularly weeded out. The plots require regular inspection for damage to plot fences, and prompt fence maintenance.

The seven species of plots that will continue into 2008 are:

CP02 S. humilis

CP03 H. suaveolens

CP04 P. pedicellatum

CP14 H. contortus

CP21 A. gayanus (Gamba grass)

- CP31 S. cayennensis
- CP32 S. australis

New species in the larger plot containing CP3 and CP4 include the following three species:

- CP23 *Ipomoea* sp (Morning Glory vine) formerly CP01
- CP26 Aeschynomene americana (American Joint Vetch)
- CP34 Crotalaria pallida (Rattlepod) (established January 2008)
- CP36 Crotalaria goreensis (Gambia pea) (established January 2008)

Any new species identified as potential plots require that immediate fencing is erected. Effort to identify and erect secure fences for the priority species that have previously been identified at Crocody lus Park should be undertaken as—soon as f easible (ie when flowering of grasses begins or as soon as an nuals appear). Sites should not be abandoned when plants die off, but any obvious aliens of non-target species should be weeded out and the site reassessed when a new ground cover emerges.

Note that since this manuscript was pr epared, the spectral sa mpling of vegetative plots has been discontinued due to resource constraints.

5 Summary

5.1 Berrimah Farm

Five plots of introduced agricultural grasses were established at Berri mah Far m in 2006. Spectral sampling at Berrimah Far m began in late S eptember 2006. While these speci es are not of the highest priority for minesite rehabilitation studies, the pastoral grasses are common in the landscape and found around the ARR minesites. The plots at Berrimah Farm are easily accessible and all plots can be sampled in one morning on a cloud-free day. Sampling has not always been undertaken at fortnightly intervals. There was a one month gap in data collection between Jun e–July 2007. The ve getation covers in the plots are dense, but need frequent monitoring for homogeneity and weeding out of non-target species. Since 2006, so me cover has been lost due to grazing. As of De cember 2007, all but one of the plots was securely fenced to m inimise disturbance through grazing. A new site of *Hyparrhenia rufa* (Thatch Grass) has been identified and will be fenced and ready for sampling during 2008. All sites require regular maintenance. It is i mportant that the 2008–09 season of data collection is made now that the plots have been securely fenced and contain dense homogenous covers.

5.2 CSIRO

Nine plots were established at CSIRO i n 2005 with six of these p lots being maintained and sampled though 2006 and 2007. Two additional plots were established and sampled in 2007. The specie's at CSIR O represent so me of the vegetation covers pertinent for minesite rehabilitation assessment studies. Li mited spectral samples were acquired bet ween January and July 2006 as the measurement pro tocols were being developed. Fortnightly data was captured from April 2006 – November 2006 and April 2007 – May 2007. From June through October 2007, much of the CSIRO d ata was captured monthly. Two of the plots are very sparse in cover and require specimens to be tran splanted in to increase e cover. No sites at CSIRO have been fenced as there are no grazing animals. Most sites are marked by star pickets and flagging tape. Two sites are still unmarked and require de lineation and all site's require regular maintenance.

5.3 Crocodylus Park

There were 21 plots established at Crocod ylus Park in mid 2006. The mix and number of species were very pertinent to minesite rehabilitation studies. Most plots were established in the upper paddock, where species diversity was ap parent. Six sites were located away from the top paddock. All sites within the upper paddock were easi ly accessible and in close proximity. The six sites external from the upper paddock were r eached by vehicle and the equipment needed to be packed and unpacked to sample these plots. Spectral sampling was undertaken fortnight ly be tween August 2006 a nd November 2 006. Som e sites were not sampled regularly in 2006. Six of the 2 1 plots established at Crocod ylus Park in 2006 were sampled in 2007. The main reason for loss of plots was a loss of cover due to grazing by donkeys due to inadequate and/or a lack of fencing. Some annual species were out competed. The six exter nal plots (from the top paddock) were discontinued in 2007 because of their location, as up to a whole day was being spen t sampling these six sites, and four of thes e species were available in the top paddock. A further 24 sites were identified in the upper paddock at the start of 2007. None of these sites were plotted or fenced and little data was obtained. No complete sets of data were r ecorded for 2007 and there were la rge (up to 6 weeks) gaps in data collection. As of late 2007, there were seven plots continuing for 2008 and four new plots being developed.

5.4 Plots ready for sampling – 2008

The twenty-five sites (Plate 8) ready for sampling in 2008 are:

- BF01 Digitaria milanjiana (Jarra Grass)
- BF02 Brachiaria humidicola (Tully Grass)
- BF03 Brachiaria humidicola (Tully Grass)
- BF04 Digitaria swynnertonii (Arnhem Grass)
- BF05 Digitaria eriantha (Pangola Grass)
- CP02 S. humilis
- CP03 H. suaveolens
- CP04 P. pedicellatum
- CP13 C. dactylon
- CP14 H. contortus
- CP21 A. gayanus
- CP22 H. Suaveolens
- CP23 *Ipomoea* sp (Morning Glory vine) formerly CP01
- CP26 Aeschynomene americana (American Joint vetch) (established January 2008)
- CP31 S. cayennensis
- CP32 S. Australis
- CP34 Crotalaria pallida (Rattlepod) (established January 2008)
- CP36 Crotalaria goreensis (Gambia pea) (established January 2008)
- CS01 Stylosanthes humilis (Townsville Stylo)
- CS02 Sorghum stipodeum (Sorghum)
- CS05 Pennisetum polystachion (Perennial Mission Grass)

Pennisetum pedicellatum (Annual Mission Grass) CS07

Urochola maxima (Guinea Grass or Green Panic) CS09

Brachiaria humidicola (Tully Grass) CS10

CS12 Melinis repens (Red Natal Grass)

As well as the yet to be established:

BF06 Hyparrhenia rufa (Thatch Grass)





BF1_2008_01-23 BF2_2008_







BF3_2008_01-23 BF4_2008_

01-23





BF5_2008_01-23 CP02_2008

-03_05









CP32_2008_03_05 CP34_2008_

03_04



5.5 Missing priority species – expanding activities at Crocodylus Park

The new sites identified for *Urochloa mutica* (Para grass) we re not established or sa mpled with the cessation of sam pling in the lower grounds of Crocodylus Park. The documentation of these sites, including herbarium identification must be done with priority so that this species can be sampled. Investigation into the sample at the lower dam of Crocodylus Park should be undertaken.

Species sampled in 2008 (see Section 5 .4) require replicates where possible. Of priorit y for Crocodylus Park are *P. pedicellatum*, *P. ploystachion* and *P. foetida*. Also of importance are: *C. inflate*, *C. dactylon* and *A. gayanus* (Gamba grass).

Because *Heteropogon contortus* is so morphologically variable, a number of plots within the three sampled sites would be very useful. Important species pertinent to minesite assessment and that have been identified at the top paddock at Crocodylus Park should be established as soon as possible and this includes: *C. mucunoides* (Calopo vine), *S. cordifolia* (Flannel weed), *P. mindanese*, *C. inflata* (Purple top chloris), *Schizachyrium* sp, *S. hamata* (Caribbean stylo) and P. foetida (wild passionfruit vine).

Others species of high int erest that have been identified at the top paddock of Crocod ylus Park, and their presence, if found again, noted for possible plot examples (even if for a short phenological cycle) include: native grasses (Bothriochloa bladhii, Digitaria bicornis, Setaria apiculata), introduced grasse s, s edge and cy press (A. gayanus, Cyperus rotundus, Dactyloctenium aegyptium, Melinus repens, Spermacoe leptoloba or purple sedge, Sporobolus jacquemontii, Urochloa pubigera), native shrub (Euphorbia heterophylla), introduced shrubs (Aeschynomene americana or Joint Vetch, Desmodium tortuosum, Hibiscus sabdariffa, H. suaveolens, Indigofera tinctoria, S. australis, S. cayennensis, Scroparia dulcis, Sida cordifolia, Spermacoce leptoloba), native vi ne (Ipomoea pes-tigridis) and in troduced vines (C. mucunoides or Calopo vine, P. foetia or wild passionfruit vine).

Species also on the priority list that have yet to be sited at the existing sites include: *Cenchrus cilaris* (Buffel grass), *Cenchrus echinatus* (Mossman River grass), *Chloris virgata* (Feather Top Rhodes Grass), *Themeda quadrivalvis* (Grader grass), *Centrosema pubescens* (Centro vine), and *Macroptilum atropureum* and *M. lathyroides* (Siratro and Phasey bean, respectively).

5.6 Important lessons learnt

- Securely fence a site as soon as a potential plot is realised;
- Regularly inspect and maintain fences of plots;
- Regularly inspect plots for alien species and weed these from the plot to maximise the chance of maintaining a homogenous plot;
- Maintain plo ts where the target species di sappears. Weed out any obvious non-target species to minimise out competition of the target species;
- As soon as species are identifiable, search for those species of prio rity, ensuring any new species are confirmed with the NT Herbarium: and,
- Ensure that species are being sampled fortnightly

References

Pfitzner K, Bollhöfer A & Carr G 2006. A standard design for collecting vegetation reference spectra: Implementation and implications for data sharing. *Journal of Spatial Sciences* 52 (2), 79–92.

Pfitzner K, Carr G & Bollhöfer A 2009. Standar ds for collecting field reflectance spectra. Supervising Scientist Report 195. Supervising Scientist, Darwin, NT (submitted).

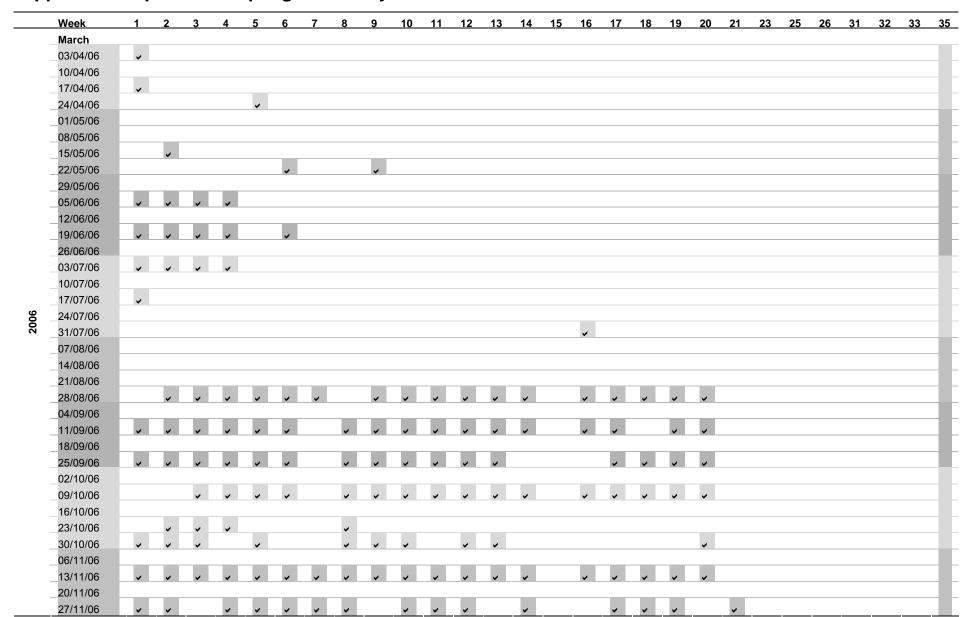
Appendix 1 Spectral sampling – Berrimah Farm

	Week starting	01	02	03	04	05
	April					
	May					
	June					
	July					
	Aug					
	25/09/06	•	✓	✓	•	•
2006	09/10/06	✓	✓	✓	✓	✓
20						
		_				
	30/10/06	✓	✓	✓	✓	✓
		_				
	13/11/06	✓	✓	✓	✓	✓
		_				
	27/11/06	✓	✓	✓	✓	✓
	Dec					
	Jan					
	Feb					
	March					
	09/04/07	•	✓	•	•	•
	23/04/07	•	✓	✓	•	✓
	30/04/07					
	07/05/07	✓	✓	✓	✓	✓
			_			
	21/05/07	✓	✓	✓	✓	✓
	28/05/07					
			_		_	
	11/06/07	✓	✓	✓	✓	✓
	25/06/07					
2007						
Ñ						
	16/07/07	✓	•	•	✓	✓
	30/07/07	•	•	✓	•	✓
	07/00/07					
	27/08/07					
	03/09/07	✓	✓	✓	✓	✓
	47/00/07					
	17/09/07	~	✓	✓	✓	•
	24/09/07					
	45/40/07					
	15/10/07	✓	✓	✓	✓	
	20/40/07					
	29/10/07					
	05/11/07					
	12/11/07					
	19/11/07					
	26/11/07	✓	✓	✓	✓	✓
	Dec					

Appendix 2 Spectral sampling – CSIRO

	Week starting	01	02	05	06	07	09	10	12
	April								
	May								
	June								
	03/07/06	~	~	~		~	~		
	10/07/06								
	17/07/06								
	24/07/06								
	31/07/06								
	07/08/06								
	14/08/06								
	21/08/06								
	28/08/06	~	✓	~	~	~	~		
2006	04/09/06								
7	11/09/06	~	✓	~	~	~	~		
	18/09/06								
	25/09/06	~	•	~	✓	~	•		
	02/10/06								
	09/10/06	~	✓	~	~	~	~		
	16/10/06								
	23/10/06	~	✓	~	~	~	~		
	30/10/06								
	06/11/06	✓	•	•	✓	•	•		
	13/11/06								
	20/11/06	✓	•	•	•	•	•		
	27/11/06	✓	✓	~	•	~	~		
	Dec								
	Jan								
	Feb								
	March								
	02/04/07								
	09/04/07	✓		V	•		•	✓	
	16/04/07								
	23/04/07	✓	✓	•	•		V	✓	~
	30/04/07								
	07/05/07	✓		V		•	~	✓	
	14/05/07								
	21/05/07	✓		V		✓	~	✓	~
	28/05/07								
	04/06/07								
	11/06/07	✓	✓	~		V	~	✓	~
	18/06/07								
	25/06/07								
	02/07/07								
20	09/07/07	✓	✓	~	~	~	~	✓	
2007	16/07/07								
	23/07/07								
	30/07/07								
	06/08/07	~		J	~	J	J	✓	
	13/08/07								
	20/08/07								
	27/08/07								
	03/09/07	~	-	,	~	V	~	✓	
	10/09/07				•	_		•	
	17/09/07								
	24/09/07								
	_01/10/07 08/10/07								
	UO/TU/U/								
								✓	
	15/10/07	•						•	
	15/10/07 22/10/07	✓						•	
	15/10/07	•						•	

Appendix 3 Spectral sampling – Crocodylus Park



CP23 – a pot of morning glory, CP25 Digitara, CP26 Joint Vetch, CP33 Sida cordifolia, CP35 – Rosella