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### **THE ASSESSMENT OF SEVEN NORTHERN TERRITORY GASTROPOD SPECIES FOR USE AS BIOLOGICAL MONITORS OF RANGER URANIUM MINE RETENTION POND WATERS**

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Issued November 1992

The Supervising Scientist for the Alligator Rivers Region has research, supervisory and co-ordination responsibilities related to effects on the environment of uranium mining in the Alligator Rivers Region and research and supervisory responsibilities related to effects on the environment of non-uranium mining in a conservation zone declared within the Region.

Views expressed do not necessarily reflect the current views and policies of the Commonwealth, the Supervising Scientist, or any collaborating organisation.

**Supervising Scientist for  
the Alligator Rivers Region**

**THE ASSESSMENT OF SEVEN  
NORTHERN TERRITORY GASTROPOD SPECIES  
FOR USE AS BIOLOGICAL MONITORS OF  
RANGER URANIUM MINE RETENTION POND WATERS**

Submitted as a requirement for the degree of  
Bachelor of Applied Science (Honours) - Environmental Biology

This project was funded for a period of 5 months by  
The Office of the Supervising Scientist  
in collaboration with  
The University of Technology, Sydney

## ACKNOWLEDGMENTS

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I would like to thank my supervisors in Jabiru, Dr Chris Humphrey and Dr Ross Hyne, for an instructive and enjoyable period of practical work, which formed the basis of this thesis. The work was partly funded by the Office of the Supervising Scientist. Thanks to the OSS staff in Jabiru for the fine hospitality and generosity in allowing five students such an opportunity. Also, after 4 years at UTS, I would like to thank my teachers, who have done a marvellous job in opening my eyes to the complex world of ecology. The electron microscope work done by Sieglinde Jobson of the Department of Dentistry, University of Melbourne has given this thesis another dimension. I should also like to thank Dr. John Walker, Dr Winston Ponder, Dr Ross Jeffree, Mr Scott Marchick, Dr David Morrison, Dr Carina Clark, Dr Bonnie McConnell and Dr Tamara Stzynda for their help and advice and Dr Richard Lim, who would've been of immeasurable help. Many thanks also to Ms Narelle Clark and Mr Nigel Lovell, whose help and patience is deeply appreciated. Thanks also to Ian, Jamo, Hollywood and Tim. Much appreciation also to Rosie Elliott who has been my comrade in pens for four rewarding years (well, three anyway).

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## ABSTRACT

Seven species of freshwater snails were examined for potential as use for the biological monitoring of Ranger Uranium Mine retention pond waters. Uranium is the main source of toxicity in this water, and because molluscs in general display a sensitivity to, or otherwise react to metal contamination, they were considered likely to respond. To test this, several stages of the life cycle of the species were examined. It was found that some species were unsuitable for use as biological monitors because of handling, rearing or other reasons not related to toxic response. One of the species tested was relatively insensitive to the toxicant. Two species of *Amerianna* were sensitive to low concentrations of Ranger RP2 waters, and the most sensitive and consistent endpoints were related to fecundity. To pursue this effect, electron microscopy was carried out on a selection of tissues possibly affected by toxicants in the waters; it was found that uranium had accumulated in the ovotestis. Other endpoints in the literature commonly seen to be sensitive indicators of metal contaminated conditions, such as juvenile and embryonic mortality, were not observed to be as sensitive as the fecundity endpoints in this series of trials.

## INTRODUCTION

A large temporal gap exists between the first bioassay performed by Aristotle testing seawater effects on freshwater organisms and the single species fish bioassay formulated by Hart in 1945 (Cairns and Pratt, 1989). Aristotle's curiosity probably did not encompass the environmental effects of mining, but by the sixteenth century concern had already been expressed. The world's first mining textbook, *De Re Metallica*, by Georgius Agricola in 1556 (Down & Stocks, 1978), contains reference to damage and dissent: "...The strongest argument of the detractors is that the fields are devastated by mining operations...the woods and groves are cut down, for there is need of an endless amount of wood for timbers, machines and the smelting of metals. And when the woods and groves are felled, then are exterminated the beasts and birds, very many of which furnish a pleasant and agreeable food for man. Further, when the ores are washed, the water which has been used poisons the brooks and streams, and either destroys the fish or drives them away... Thus it is said, it is clear to all that there is greater detriment from mining than the value of the metals which the mining produces". *De Re Metallica* does not specify ownership arrangements or ultimate responsibility in those early days.

Mankind's use of metals has increased to the extent at which mineral extraction and associated processes have significantly affected on our natural heritage (Down & Stocks, 1978; Moore & Winner, 1989).

Initially, (partially because classical biologists did not regard industry as a relevant study area,) monitoring of aquatic pollution was physico-chemical, dealing with such parameters as dissolved oxygen, biological oxygen demand, temperature and dissolved solids. In 1945, Hart published the first single species toxicity test protocol, subsequently accepted by the American Society for Testing and Materials (Cairns & Pratt, 1989). In 1948, Ruth Patrick (under Hart) was to convince regulatory authorities in the USA that biological methods should be used alongside physico-chemical ones, to adequately protect aquatic ecosystems (Cairns & Pratt, 1989). The rationale was (and is) as follows: physico-chemical methods did not account for periods between sampling; the toxicity of complex mixtures could not be instrumentally quantified; biological effects often occurred below the level of chemical detection; and, the results were in themselves meaningful only in terms of human use of the system (Cairns, 1982). Biologists reply thus: the species inhabiting the water body provide a static, temporal record of events therein; organisms will be exposed to and therefore react to any contaminants present; consistently sensitive endpoints exist; and meaningful extrapolations to any ecological level, including man's needs, are possible. This argument was accepted by industry, regulatory authorities, and the scientific community. After the 1951 establishment of

Doudoroff's protocol (using the test formulated by Hart), many single species toxicity tests were developed, including tests upon invertebrates and algae (Cairns & Pratt, 1989), and were used to some extent in the establishment of emission limits (ie. effluent-based standards).

In the 1970's, a growing awareness on the part of industry and legislators of contamination problems produced pollution-limiting technology. A technological philosophy was particularly attractive to these groups, and the terms Best Available Technology (BAT) and Best Practicable Technology (BPT) were soon encountered in legislative documents (Cairns & Pratt, 1989). BPT (= affordable technology) was often implemented without consideration of an industries local situation. Criticisms of BAT and BPT practices included: the size and assimilative capacity of an ecosystem was not considered; each discharge was regarded uniquely, without factoring other discharges into the same receiving system; there was a chance of over-treatment of no benefit to the ecosystem; there was no incentive to develop better technology, once a BPT was in place (Cairns & Pratt, 1989). The shortfalls inherent in effluent-based and technology-based standards have belatedly resulted in the incorporation of biological evidence to hazard evaluation (Cairns & Pratt, 1989). As such, biological monitoring is in its infancy.

In all, biological monitoring assesses the impact of pollutants and, in association with appropriate regulation, derives standards of release limitation. As well as its regulatory role, biological monitoring should be predictive (Stephan, 1986). By this is meant that conclusions drawn under one regime should be applicable to another, perhaps after some form of transformation.

## MINING AND HEAVY METALS

Although mining accounts for only a small percentage of total societal water use (in the US 1960's, 2% of industrial water use, or around 1.8 billion litres/day), and returns around 78% of this to the environment, the water used in the various stages of processing is often highly contaminated. Mining effluent can contain 2 to 10 times the amount of contaminant metal than untreated ore (Down & Stocks, 1978).

Contamination due to metal extraction *per se* of aquatic systems is of several forms:

- dissolved solids - soluble salts
- organic reagents - frothers, collectors, flocculants etc
- suspended solids
- acidic drainage
- heavy metals (Down & Stocks, 1978).

Other impacts of the mining process can arise from site runoff water, mine drainage, pumped mine water, contamination of ground water flows and deposition into the aquatic system of airborne contaminants from smelting (Down & Stocks; 1978, Humphrey *et al.*, 1990).

Estimations of anthropogenic mobilisation of heavy metals into the biosphere is presented below (Nriagu & Pacyna, 1988).

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### Estimated annual heavy metal input into biosphere

	Total input	Mining and processing
Arsenic	120kt	6kt
Cadmium	20kt	2kt
Chromium		11kt
Copper	2150kt	14kt
Manganese		43kt
Mercury	11kt	.01kt
Molybdenum	110kt	.5kt
Nickel	470kt	13kt
Lead	1160kt	6kt
Antimony	72kt	4kt
Selenium	79kt	12kt
Vanadium	71kt	1.2kt
Zinc	2340kt	28kt

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The total toxicity of these metals (in terms of water needed to dilute to safe drinking standard) is more than radioactive and organic waste toxicity combined (Nriagu & Pacyna, 1988).

## THE FATE OF METALS IN THE AQUATIC ECOSYSTEM

Heavy metals are generally less toxic in hard waters (high  $\text{Ca}^{2+}$ , high  $\text{CO}_3^{2-}$ ) due, in some part, to competition between the heavy metal and  $\text{Ca}^{2+}$  for membrane binding sites (Persoone *et al.*, 1989; Rozsa & Salanki, 1989).

Many of the toxic mechanisms of heavy metals remain uncertain although some work has been carried out.

Inhibition of metabolic coupling via gap junctions was demonstrated using cell culture (Loch-Caruso *et al.*, 1991). The metals investigated were arsenic, cadmium, lead and zinc. Wild-type Chinese hamster V79 cells proficient in the phosphorylation of 6-thioguanine (6-TG) by hypoxanthine phosphoribosyltransferase were cultured with mutant V79 cells lacking the enzyme. The phosphorylation of 6-TG produces a toxic compound, and inhibition of the gap junction transfer of 6-TG was shown by an increased survival of the mutant cells compared to the proficient cells. This implies that heavy metals inhibit the intercellular transfer of metabolites and possibly other types of molecules.

The junction membrane permeability of the salivary glands of *Chironomus* sp. (Diptera) was investigated in terms of the divalent cations of the alkaline earth metals, barium, calcium, magnesium and strontium (Oliviera-Castro & Lowenstein, 1971). It was found that the permeability of the membrane was depressed in the order  $\text{Ca}^{2+} > \text{Mg}^{2+} > \text{Sr}^{2+} > \text{Ba}^{2+}$ . This explains the mitigating role  $\text{Ca}^{2+}$  (and to a lesser extent  $\text{Mg}^{2+}$ ) displays in the toxicity of heavy metals, in that it has a higher membrane binding potential than the heavier elements in the series. Spehar *et al.*, (1978) also attribute the effect of cadmium on gastropods and trichopterans to a disruption of membrane permeability.

Heavy metals also produce teratogenic effects. Paulij *et al.*, (1990) investigated the effects of copper on the embryogenesis of the squid *Sepia officinalis*. They reported a shortening of the developmental period, but a lowered embryonic viability. Sublethal exposure of lead to Green Frog (*Rana clamitans*) tadpoles was seen to inhibit acquisition and retention of discriminant avoidance learning, (Strickler-Shaw & Taylor, 1990) implying a neurological pathology.

Hyne *et al.*, (1991) report on the deposition of uranium in the discharged nematocyst of *Hydra viridissima*. This was thought to inhibit the growth of new nematocysts resulting in a decreased ability to catch prey. Wrenn *et al.*, (1985) review the literature of uranium toxicity in mammals, including humans. They report no evidence of bone sarcoma having been induced by uranium ingestion, but do report bone sarcomas caused by decay chain products of uranium, and soft tissue sarcoma at sites of particulate uranium deposition. Most damage by uranium ingestion occurs in the proximal renal tubule cells. If the dose is not great enough to cause mortality but massive renal failure, the tubule cells are replaced, although displaying differences from original cells. Uranium in sublethal quantities is rapidly excreted (95% removal over 24hrs), but a small amount may be deposited throughout the skeleton, where it appears to be immobilised. The metabolism of uranium was said to resemble that of the alkaline earth metals.

Because many of the heavy metals are essential trace elements (Chromium, cobalt, copper, iron, manganese, molybdenum, nickel, vanadium) cellular mechanisms operate to facilitate their uptake. Therefore, in higher concentrations the uptake of excess metal is guaranteed.

## **FACTORS AFFECTING THE SPECIATION OF METALS IN THE AQUATIC SYSTEM**

In general, it is the speciation of the metal ions which determines the toxicity in a given situation, as well as total metal concentration. Complex organic and inorganic reactions, under the influence of edaphic factors, determine the speciation and toxicity of metals in the aquatic system.

Acidity determines the carbonate activity, and at low pHs the usually stable carbonate/heavy metal complex is largely absent. Low oxygen levels can favour the formation of insoluble sulphides (Kelly, 1988). Naturally occurring organic substances (fulvic and humic acids of plant origin, and various microbial and algal products) are capable of complexing metals, and to varying degrees determine bio availability (Moore & Winner, 1989; McKnight, 1981; McKnight & Morrel, 1979). Clays and silts have a metal binding capacity indirectly proportional to particle size. Norris *et al.*, (1981) observed high metal concentrations in stream water, where the stream bed was sand. A corollary is that the biota of oligotrophic acidic (pH 5 to 6.5) streams are much more sensitive to metal contamination than mesotrophic neutral stream biota.

What follows is a short discourse on the speciation of selected heavy metals, namely copper, lead, zinc and nickel. Uranium is discussed in the last section of this introduction.

## Copper

The aqueous chemistry of copper is complex because it can exist in three stable oxidation states. Copper as the divalent cation, tends to disappear quickly, due to complexation, precipitation and adsorption with inorganic, and complexation with organics (Effler *et al.*, 1980). Taub *et al.*, (1986) demonstrated that 25% of the copper recoverable from a laboratory microcosm was adsorbed to quartz sand. In some instances, the chemistry of copper added to natural ecosystems is controlled largely by humic substances. Modelling the speciation  $\text{Cu}^{2+} + \text{HA} \rightarrow \text{CuA}^+ + \text{H}^+$ , McKnight and Morrel (1979) derived an equilibrium constant  $K_{\text{eq}} = 10^{1.5}$ . Certain chlorophytic algae are capable of releasing humic material which is a weak copper complexing agent, and the blue-green algae can produce both weak and strong complexing agents (Moore & Winner, 1989). These reactions render the copper biologically unavailable, and reduce toxicity.

The general consensus is that most copper is adsorbed under normal conditions and the concentration of the free ion is very low. The concentration in the filterable fraction ( $<0.4\mu\text{m}$ ) may range from 3 to 80% of total copper, and is largely bound to organics (Kelly, 1988). The tendency for copper to bind to inorganics is more pronounced when the particles are modified by manganese or iron oxide coatings. Concentrations typical of unpolluted rivers range widely (0.6 to 400ppb), with most median concentrations from 5 to 10ppb (Kelly, 1988). Historically, the copper content of available ore has decreased from 40% to around 0.2% processed today (Down & Stocks, 1978).

## Lead

Lead, a group IV element along with carbon, germanium, silicon and tin, is capable of forming organoderivatives. These derivatives are considerably more toxic than the free stable ion (Kelly, 1988). Under anaerobic conditions *in vitro*, lead methylation has been observed. This has not been observed in the field (Kelly, 1988). Background levels of lead have been measured as low as 0.006 to 0.05 ppb, although in pristine mineralised areas, concentrations two or three orders of magnitude greater have been measured (Kelly, 1988). The divalent cation is most stable under oxidising conditions, forming carbonate above pH 7.2 and hydroxide above pH 10. The carbonate is assumed to act as the major factor controlling the free ion concentration, although at acidic pH the sulphate may assume this role. Lead and zinc often occur together.

## Zinc

Zinc exists in only one stable oxidation state, and its chemistry is more easily understood. Wide use of zinc makes analysis of very dilute solutions difficult because of micro-



contamination, but measures of filterable fractions in pristine waterbodies range from 1.8 to 20ppb. Adsorption onto particulates is the major factor in determining free ion concentration. Hydroxides and carbonates formed above pH 8 remain readily soluble. Adsorption is strongly pH dependant, almost nil at pH 6, and increasing with pH (Kelly, 1988).

### **Nickel**

In most natural waters, pH 5 to 9, the most prevalent nickel ion is  $\text{Ni}^{2+}$  or the carbonate, with hydroxide, chloride, sulphate and ammonium also present. A large proportion (5 to 80%) of the total nickel concentration is found in the non-filterable fraction, probably adsorbed onto particulate material. Particulate adsorption, rather than precipitation, is thought to control free ion concentrations (Kelly, 1988).

Particulate phases of the metal, as organic or inorganic precipitant complexes, can enter the sediment (n3). Disturbance, both physical and biological (benthic macroinvertebrates) can at any time remobilise these sediment-bound metals (m8). For both rivers and lakes the nature of the sediment is an important factor in controlling sedimental adsorption. Adsorption and precipitation are functions of surface area, and the capacities of the sediments increase as the particle size decreases (Kelly, 1988). Where there is little retention of fine clay and organics, the concentrations of heavy metals in the sediment is likely to be low (Norris *et al.*, 1981).

## BIOLOGICAL MONITORING - METHODOLOGY

A current source of contention in the biological monitoring field centres around the aptness or otherwise of traditional single species toxicity tests to accurately reflect effects in the field. Proponents hold that natural variability in the field precludes use of wider ranging studies for regulatory purposes, that information gained from multispecies testing is difficult to interpret and that the results of the two methods are in any case mostly comparable (Tebo, 1985). Critics say that it cannot predict effects at the population and community levels, that an all-encompassing standard of toxicant limitation is not effective in protecting diverse systems (Tebo, 1985), and that unanticipated effects can be observed in multispecies testing (Cooper & Stout, 1985). All acknowledge that effects of contamination on populations and communities is but poorly understood, and emphasise the need for an escalation of effort in this area (Tebo, 1985; Cairns & Pratt, 1989; Loewengart & Maki, 1985).

Current methodologies in biological monitoring include (Humphrey *et al.* 1990):

- Bioassays - the use of defined species to estimate toxicity *in vitro*;
- Early detection systems - physiological, behavioural biochemical and histopathological techniques (bioassays) used *in situ*;
- Indicator organisms - the presence or absence of sensitive or resistant organisms thought to characterise particular conditions *in vivo*
- Autecological (population) studies - the natural history of populations of species that provides essential baseline data;
- Community structure and function - how species and populations interact in terms of niche structure and energy movement *in vivo* and *in vitro*; and
- Bioaccumulation - the accumulation of contaminants in exposed organisms, serving as a record of ambient contaminant concentrations over time.

There is no ideal biological monitoring method. An ecological impact study or monitoring program should include those methods considered appropriate to the task.

There is a large literature on single species toxicity tests that cover all levels of community structure of the aquatic ecosystem. Extensive reference lists can be found in Phillips, (1980)' Kelly, (1988) and Krenkel, (1975). The literature of vertebrate biological monitoring will not be discussed.

### **Microorganisms**

A commercially available microbial (Microtox) assay uses luminescent bacteria. The bacteria are freeze dried and resuspended for use. Light production is measured for a range of dilutions of the toxicant, and toxicological parameters can be then estimated. Chang *et al.*, (1981) describe the use of this system for pesticides, and Loewengart and Maki (1985) provide results using Microtox in comparison to multispecies methods. In the latter instance, Microtox took 5% of the time, was cheaper, yielded fairly reproducible results, required no culturing and was standardised and easy to handle.

### **Algae**

Algae, the primary producer in many aquatic systems, have been well investigated in terms of metal susceptibility. Also, due to copper sulphate treatment of algal blooms in eutrophied water bodies, copper relations with a variety of alga are well understood. Although a micro-nutrient requirement, copper sensitivity becomes important at concentrations higher than 10mg/L, beyond which more resistant species may become dominant (McKnight & Morrel, 1979; McKnight, 1981). Accidental inoculation of a copper resistant strain of alga into a multispecies trial using ten algae, resulted in a more complex co-dominance pattern compared with those not so treated, with several species able to maintain populations under the dominance of the resistant strain (Taub *et al.*, 1986).

Some authors recommend against the use of algae in single species toxicity tests because they may increase production under stress *et al.*, 1980), and in stationary growth phase are known to release metal binding substances (McKnight & Morrel, 1979). It is also difficult to maintain mono-species cultures in many situations, and there are problems associated with agglutination (Effler *et al.*, 1980). Other authors have reported excellent results and recommend their use as biological monitors (Kallqvist, 1984) Algae tend to be included in most multispecies tests, particularly in microcosm and mesocosm trials, to determine how effects at the bottom of the food chain are transmitted upward, and also how events at higher levels affect the algae (Carpenter *et al.*, 1987).

Copper inhibition of nitrogen fixation by the blue-green algae was investigated by Horne and Goldman (1974), and was more sensitive than either photosynthesis or chlorophyll *a* for the whole lake. Inhibition was seen to occur at 10ug/L.

### Macroinvertebrates

Macroinvertebrates have become important in single species toxicity tests, *in situ* sampling and in multispecies tests. They have a wide range of sensitivities and are involved in complex interactions, and hold promise as the best group for biological monitoring purposes (Hellawell, 1986).

Various members of the cladoceran *Daphnia* genus have well established protocols (Mount & Norburg, 1984). The cladocerans as a group are excellent subjects for short-term testing, because they have a rapid life cycle of 4 to 7 days, during which they may produce up to three broods. A 4-day *Ceriodaphnia* test recently published (Masters *et al.*, 1991), shows comparable sensitivity to the 7-day test of Mount and Norburg.

*Hydra* species have been used in various situations. OSS uses *Hydra viridissima* in its seasonal testing of retention pond waters from Ranger Uranium Mine (OSS, 1988). Goss *et al.*, (1986) provide a protocol for *Hydra* sp. in human health hazard evaluation.

### Aquatic Insects

In the past, aquatic insects have been extensively used to assess the status of water bodies. Indices such as the Trent Biotic Index (Woodiwiss, 1964) and the Chandler Score (Chandler, 1970) were devised to assess organic pollution and are generally disfavoured today, although the concept of indicator species maintains its importance. Multivariate analysis and methods applied to presence/absence or species abundance is the most common contemporary analysis. Several Australian publications report success using this approach (Marchant, 1990; Norris *et al.*, 1981; Weatherly *et al.*, 1967). However, the aptness of accepted methodology and mathematical technique has been questioned, with some authors preferring distance measures such as the Brau-Curtis over the Chi-squared measures used in detrended correspondence analysis, among others (D. Faith *pers comm.*, 1991). Also, natural background fluctuations can often swamp trends if sufficient data through time are not present in the analysis (C. Humphrey *pers comm.*, 1991).

Apart from extensive use as *in situ* biological monitors, aquatic insects have received scant attention. Their size and complex life history (which usually includes several moults) make them unacceptable in single species testing (Smock, 1983). Several authors have included aquatic insects in multispecies microcosm testing, with variable results, and in some cases, tenuous endpoints, e.g. avoidance behaviour (Pontasch & Cairns, 1991).

### Gastropods as biological monitors

Much pollution work has been carried out on the marine molluscs, particularly bivalves in the genera *Mytilus*, *Perma* and *Crassostrea*. These species have a broad marine distribution, permitting their use for wide-ranging metal pollution studies. Marine gastropods have been used to a lesser extent, and freshwater gastropods still less.

Ravera (1977) examined the effects of heavy metals on *Biomphalaria glabrata*, a freshwater pulmonate, in terms of mortality and fecundity of the adults, and the viability of the embryos. It was seen that the species was sensitive both as adult and embryo. Larval survival after exposure to chrome and copper at higher concentration (>1.0ppm) was of the same order as adult survival, but was greater than adult survival at concentrations from 0.1 to 0.5ppm. He concluded that the species should be used in toxicity testing because it was easily reared and was sensitive to the heavy metals.

Other early testing includes that of Borgman *et al.*, 1978 (lead effects on *Lymnaea palustris*), Rehwoldt *et al.*, 1973 (copper, zinc, nickel, cadmium, mercury and chromium effects on *Amnicola* sp.) while reference lists can be found in Mance (1987), Phillips (1980) and Simkiss and Mason (1983).

Holcombe *et al.* (1984) provide methods for conducting embryo through to adult exposures. They tested the effects of cadmium and reduced pH on the pulmonate *Aplexa hypnorum* and concluded that the species was suitable for use as a biological monitor in all its life stages. Brown (1980) showed that *Hydrobia jenkinsi* responded (mortality) to *in situ* metal levels, but not to dieldrin levels, which reached 30ppb. Marigomez *et al.*, (1986) demonstrated feeding suppression and weight loss in the terrestrial pulmonate (slug) *Arion ater*. Percentage weight loss was linear at high concentrations and exponential at the lower concentrations, suggesting the failure of a compensatory mechanism after 15 days. Watton and Hawkes (1984) report on a series of copper and ammonia trials conducted on the prosobranch *Potamopyrgus jenkinsi*. Juveniles were seen to be less tolerant than adults, and were generally less tolerant than most other invertebrates reported in the literature. They concluded that the species would be useful as a biological monitor of copper and ammonia in the various ionic forms. Munzinger and Guarducci (1988) report on the exposure of *Biomphalaria glabrata* to low levels of zinc. The embryo was seen to be the most sensitive stage. Juveniles were roughly 3 times more sensitive (comparing percentage mortalities after 33 days exposure) than the adult. They concluded that the species was a successful laboratory indicator of low level zinc pollution. Russell *et al.*, (1981) investigated the effect of cadmium on *Helix aspersa* and its accumulation and entry into the food chain.

Among other observations, they noted a disruption of gametogenesis. Ramesh Babu and Venkateswara Rao (1985) investigated the biochemical basis of copper toxicity in *Lymnaea luteola* and concluded that copper altered the redox potential of the snails metabolism by reducing the activity of cytochrome oxidase and NADH to NAD ratio. Skidmore *et al.*, (1988) demonstrated a reduction in fecundity of *Physastra gibbosa* following static exposure to copper concentrations above 16ppb. They recommend the fecundity endpoint as a means of measuring sublethal toxicity of substances to Australian aquatic species.

Although the work performed to date on the pulmonates does not comprise a large part of the molluscan metal-related literature, what does exist indicates their sensitivity to heavy metals and authors frequently recommend them as biological monitors of heavy metals.

## URANIUM AND URANIUM MINING

The extraction and on-site processing of uranium ore poses risks to the aquatic environment through the release of uranium and its decay chain products into local systems. On-site processing is invariably carried out because of the saving of transport costs of a semi refined product over raw ore. The Alligator Rivers Region, area of operation for the Ranger Uranium Mine (RUM), receives a highly seasonal rainfall. Dams have been constructed around the mine and mill to receive runoff water during the wet season. Because the rainfall can be intense, RUM is licensed under recommendations of the Fox Report (1977), to release water from its Retention Pond #4 once the flow rate in the nearby Magela Ck exceeds  $20\text{m}^3/\text{sec}$ . This retention pond receives runoff from the waste rock dump, and is low in uranium (40ppb) (Hyne, 1989). Retention Pond #2 receives runoff from the milling area and has highly variable uranium concentrations (660-3000ppb). In the 1991 wet season, values of between 1300 and 1700ppm were recorded. RUM is licensed to release from RP2 once every ten years. Release from this dam would only occur if there was a possibility of overflow during the wet season. There has been no release from RP2 to date (Diagrams 1 and 2 show respectively, a map of the Alligator Rivers Region and layout of RUM).

The environmental chemistry of uranium is complicated by the variety of forms the uranyl ion may take. In Magela Ck, slightly acid conditions (pH 6) and low alkalinity (as  $\text{CaCO}_3$  1.6mg/L) (OSS, 1988) favour  $\text{UO}_2\text{OH}^+$  (50%) and  $\text{UO}_2^{2+}$  (10%) over neutral or amnionic forms, the remainder comprising complexed forms of  $\text{UO}_2$  (S. Machett *pers. comm.*, 1991). The toxicity of the cations is much higher than the anions because of membrane permeability (R. Hyne *pers. comm.* 1991).

Giesy *et al.*, (1986) report stability constants for various species of the uranyl ion complexed with Aldrich<sup>R</sup> humic acid. The stability constant ( $K=[\text{ML}]/([\text{M}][\text{L}])$ ) of the neutral and amnionic carbonate forms were 2 to 3 times larger than those of the free uranyl anions. They determined the percentage presence of the various species in a soft, acid, humate rich water (pH=5.5, Ca=5.2mg/l, TDOC=14mg/l). 77.1% of the uranyl ion was present as the free anion (31.1) or the mono-hydroxide (46.0). The remainder (22%) was bound to humic acid, with 0.6 bound to sulphate. These figures compare reasonably well with Machett's figures above, and apart from having a lower dissolved organic carbon level, Magela Ck water is fairly similar to the water used in the Giesy *et al.*, report.

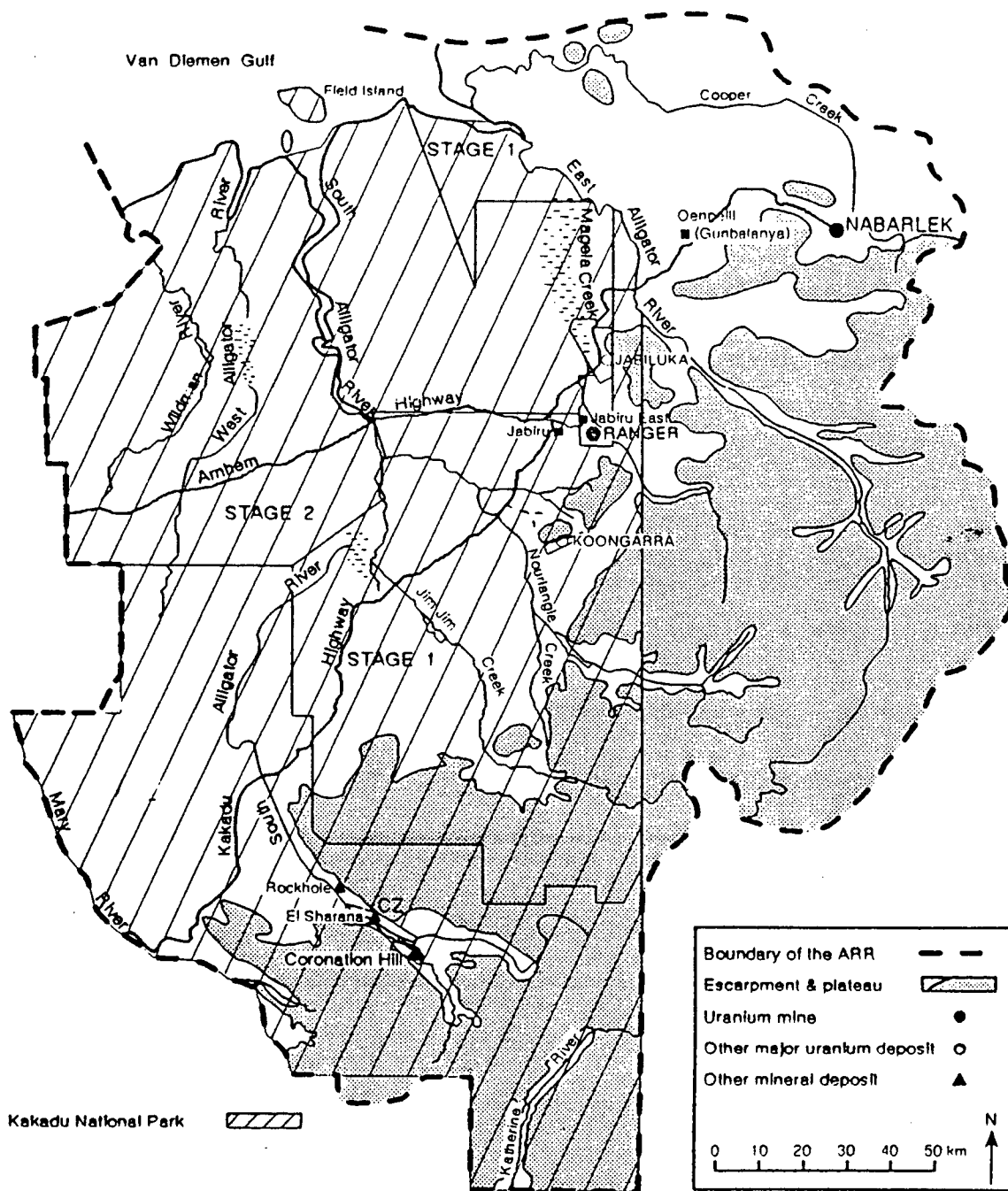


Diagram 1. The Alligator Rivers Region



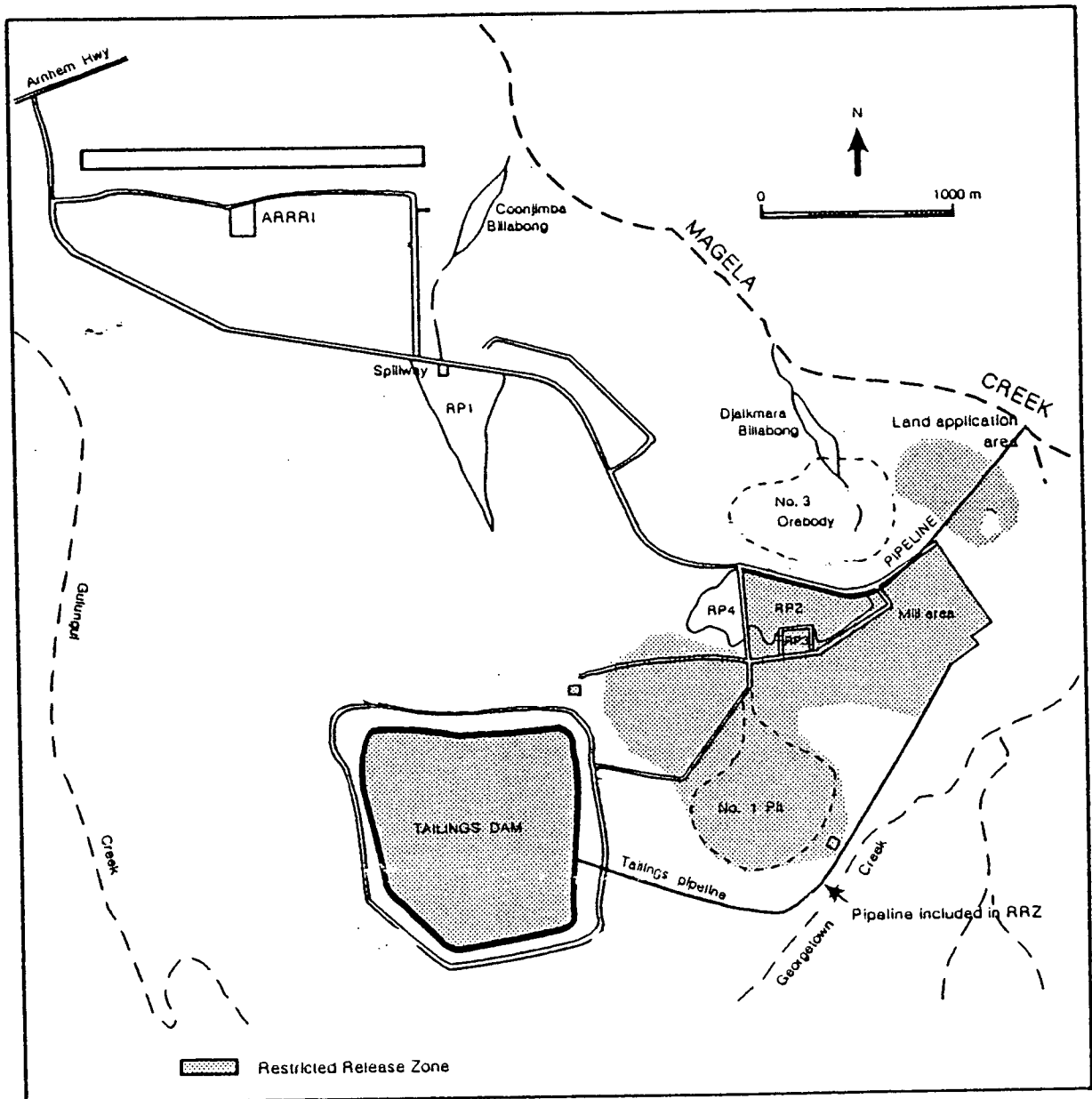


Diagram 2. Layout of operations and water management system at Ranger

Hydrous oxides of iron and manganese may also remove the uranyl ion from solution. Soluble manganese will co-precipitate with heavy metals under oxidising conditions. Under slightly acidic and reducing conditions, the manganese will redissolve and release the bound heavy metal back into the water column (OSS, 1988).

Uranium present in retention pond waters released into Magela Ck will be present to a significant extent in toxic cationic forms. Precipitation and remobilisation will presumably occur along the length of the stream. In the cationic form the uranyl ion poses a potential threat to fauna of the creek and floodplain system.

## MATERIALS AND METHODS

### Species descriptions

The seven local gastropod species selected for assessment are distributed amongst the freshwater bodies of the region. They were:

<i>Amerianna carinata</i>	Pulmonata:Planorbidae
<i>Amerianna cumingii</i>	Pulmonata:Planorbidae
<i>Gabbia</i> sp.	Prosobrancha:Bithyniidae
<i>Glyptophysa</i> sp.	Pulmonata:Planorbidae
<i>Gyraulus</i> sp.	Pulmonata:Planorbidae
<i>Helicorbis</i> sp.	Pulmonata:Planorbidae
<i>Lymnaea</i> sp.	Pulmonata:Lymnaeidae

*A. carinata* is sinistral with a flattened carinate spire. It measures around 10 mm in length by 8 mm in width at full size. The shell is moderately robust. It is monoecious.

*A. cumingii* is sinistral with a rounded depressed spire. Mature individuals measure from 8 to 12 mm in length and 8 mm in width. Growth continues into senescence where individuals may be up to 15 mm in length. The shells are moderately robust, slightly stronger than those of *A. carinata*. It is monoecious.

*Gabbia* sp. is dextral and turbate. It measures up to 5 mm in length and 4 mm in width. It is dioecious and not sexable by gross morphology. The shell is robust.

*Glyptophysa* sp. is sinistral with a tall pointed spire. It measures 20 mm in length and 13 in width, although larger individuals are commonly encountered. It is monoecious. The shell is robust but without distinctive sculpturing typical of the genus.

*Gyraulus* sp. is dextral and discoidal. It measures up to 6 mm in diameter. It is monoecious. The shell is fragile.

*Helicorbis* sp. is dextral and discoidal. It is up to 7 mm in diameter, and is flattened on the underside. It is monoecious. In habit it is very similar to *Gyraulus* sp., and both species were observed in the wild to be epiphytic grazers on the underside of water lily leaves. The shell is fragile.

*Lymnaea* sp. was the largest species assessed. It is dextral and turbinate. It measures 30 mm in length by 25 mm in width at full size. It is monoecious. The shell is fragile, and care was needed in handling this species. It produced copious slime and excreta. Individuals tended to cluster together and necrophagy apparently occurred.

## REARING CONDITIONS

All species were fed washed outer lettuce leaves as required. An extra calcium source was not supplied. Fine to medium sand (<2mm) was added to the rearing tanks. All rearing tanks were aerated with oil-free compressed air using aquarium stones.

The larger species were reared in wide, slightly tapered, circular tanks 1.5m in diameter and up to 0.6m deep (Plate 1b). The conductivity varied according to the degree of evaporation and the period between cleaning and half replacement which occurred on an irregular basis. No attempt was made to hold conductivity at any particular level. Measurements made indicated 80µS/cm as an average value. Species reared successfully in these tanks were *A. carinata*, *A. cumingii*, *Glyptophysa* sp. and *Lymnaea* sp..

Tanks 1.5m deep by 0.6m square and tanks 0.4m deep by 0.8m by 0.4 m were also used for rearing. The 1.5m deep tanks were quite suitable for *A. carinata*, *Gabbia* sp. and *Glyptophysa* sp., but best conditions appeared to prevail in the wider shallower tanks. *A. carinata* and *A. cumingii* were successfully reared in the 0.8m X 0.8m X 0.4m tanks.

*Gabbia* sp., *Gyraulus* sp. and *Helicorbis* sp. were reared in Magela Ck water in 20L aquaria. For the large tanks, the rearing water was from the town main supply, which was much higher in conductivity and carbonate than Magela Ck water. Populations of *A. carinata*, *A. cumingii* and *Glyptophysa* sp. were maintained in 40L aquaria of Magela Ck water to provide acclimated adults for sub-lethal testing. This procedure was not wholly successful; cleaning by siphonation sometimes injured individuals and recruitment was low, possibly because (of loss of) juveniles during siphoning.

*A. carinata*, *A. cumingii*, *Glyptophysa* sp. and *Lymnaea* sp. were in adequate supply throughout the period of testing. The smaller species were not successfully maintained in sufficient numbers. *Gabbia* sp. and *Helicorbis* sp. stocks dwindled to zero.

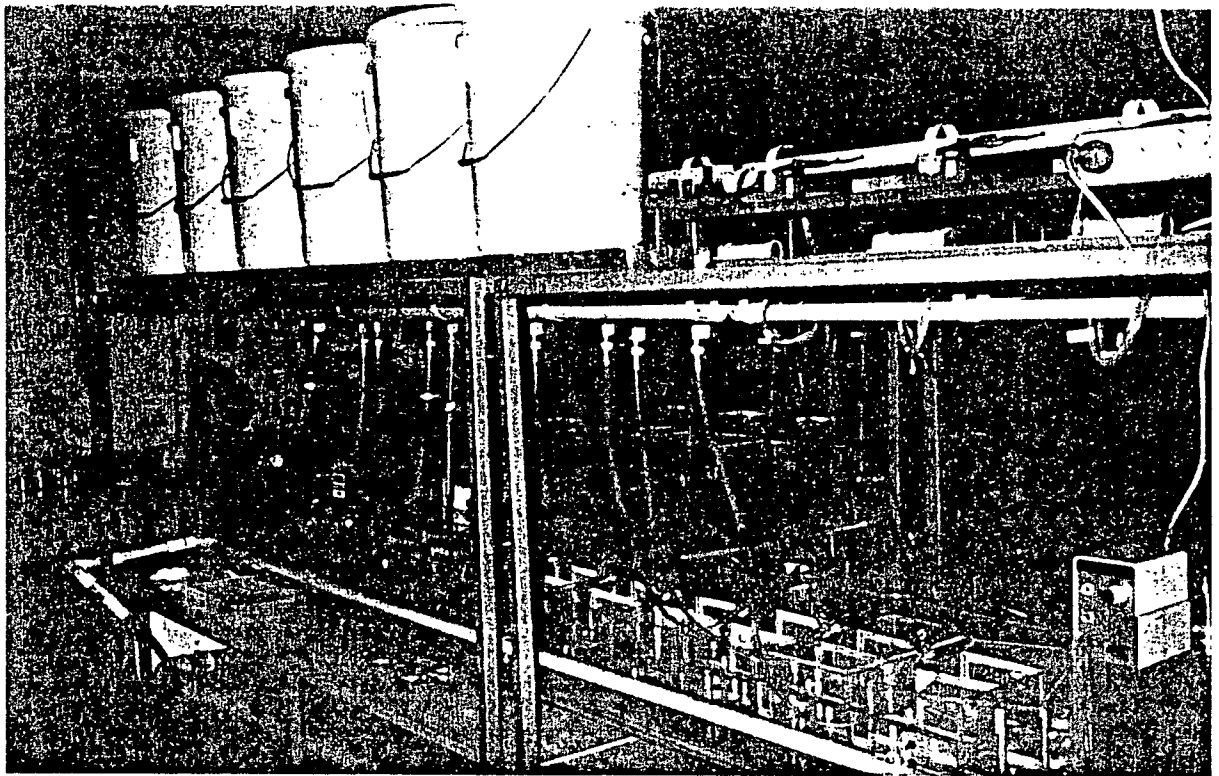


Plate 1a. Dosing apparatus

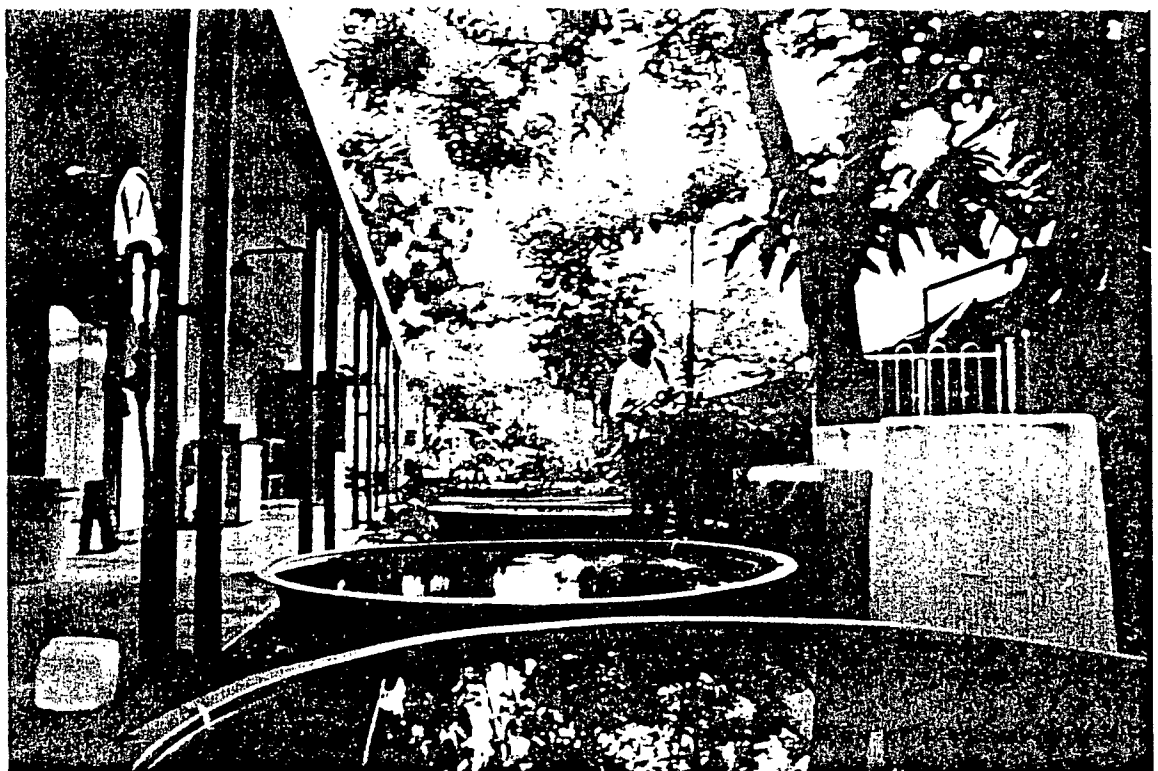


Plate 1b. Rearing tanks



Plate 1c. Magela Ck. High flow



Plate 1d. Magela Ck. Low flow

Confinement of snails, for exposure to toxicant or to obtain eggs for observation or exposure, was in clear open-ended 50 mL plastic sample jars. It had previously been established at OSS that the snail species readily oviposited on the inner surface of the vials. This allowed unobstructed observation of the developing embryos. In the case of adult confinement, open ends were covered with nylon mosquito netting (around 2mm gauge), held in place with rubber bands. For juvenile exposure, nylon netting of between 0.4 and 0.5mm gauge was used.

When confined, pairs of snails were fed daily a standard disc (20mm diameter) of washed outer lettuce leaf. Observations of egg masses were made under a dissecting microscope. To prevent desiccation or overheating of the egg masses during observation, vials were transferred into a glass histology box containing rearing water.

## PRELIMINARY OBSERVATION #1

Prior to the commencement of toxicity testing, preliminary observations were undertaken to become familiar with the various species in terms of reproductive capacities and developmental characteristics in the laboratory.

The first observation involved *A. carinata*, *A. cumingii*, *Glyptophysa* sp., *Gyraulus* sp. and *Helicorbis* sp. Four replicates per species were used except for *A. cumingii*, which had seven replicates. Each replicate was a vial containing a pair of snails. These were placed into the breeding tanks. Observations of egg mass and egg numbers were made daily for 5 days, commencing 19/1/91. Plates 2a to 2h show the developmental stages of *A. carinata*, which are typical for the other species examined. Plates 3a to 3d show some of the irregularities seen during embryonic development.

## PRELIMINARY OBSERVATION #2

This was conducted in a similar manner to the previous observations (#1) and commenced 25/1/91. At the end of the 5 day laying period, the development of each egg mass was followed through to hatching by taking daily counts of the developmental stages present. This meant that on the first day of developmental observation (i.e. after 5 day laying period), early stages were present from only the previous day's laying, and later stages were from days prior to this. However, since the age of each egg case was known, it was possible to derive percentage data relating to the duration of each stage. The species observed were *A. carinata*, *A. cumingii*, *Gabbia* sp., *Glyptophysa* sp., *Gyraulus* sp. and *Lymnaea* sp..

Preliminary observations #1 and #2 provided information on the reproductive output and developmental characteristics (such as duration and %mortality) of each embryonic stage, and the variation associated with these parameters. In terms of the ensuing toxicity trials, the variation associated with each parameter was considered to be as important as the mean value. On the basis of these observations, it was possible to "score" each species and parameter as:

Good (+=low variability and sufficient quantity of eggs/egg cases)

Moderate (0=moderate variability and sufficient quantity)

Poor (-=high variability and/or low quantity).



Developmental stages of *A. carinata*

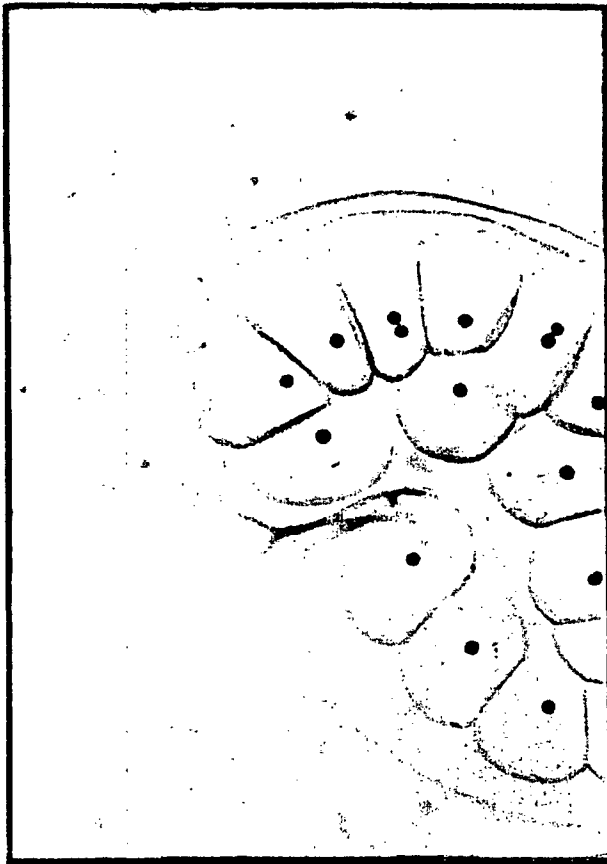


Plate 2a. Gastrula

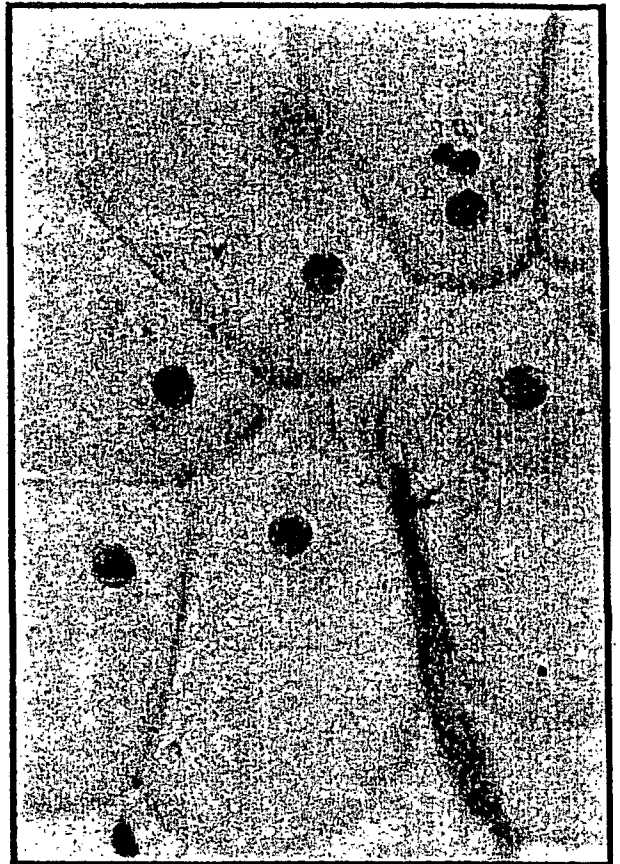


Plate 2b. Early trochophore

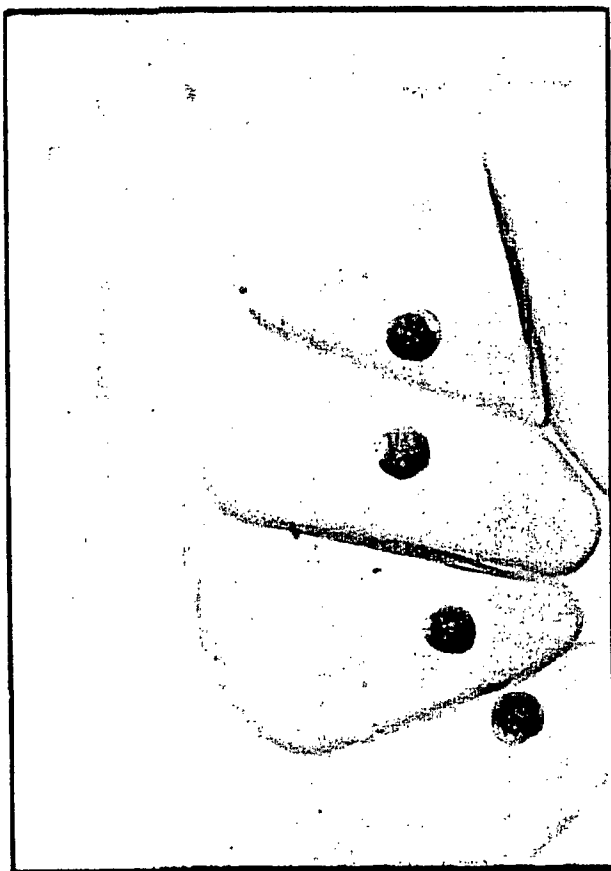


Plate 2c. Late trochophore

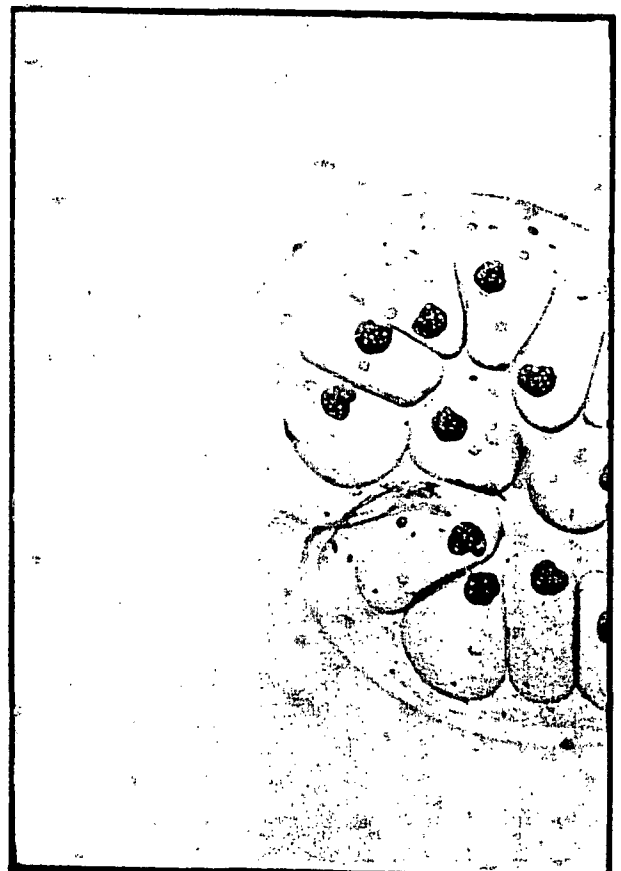


Plate 2d. Velliger

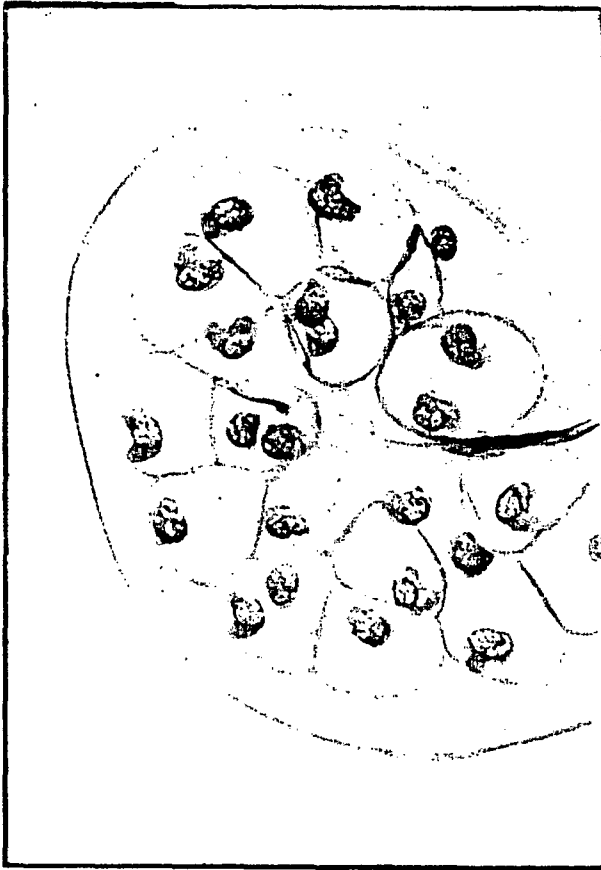


Plate 2e. Hippo



Plate 2f. Early hatchling



Plate 2g. Mid hatchling



Plate 2h. Late hatchling

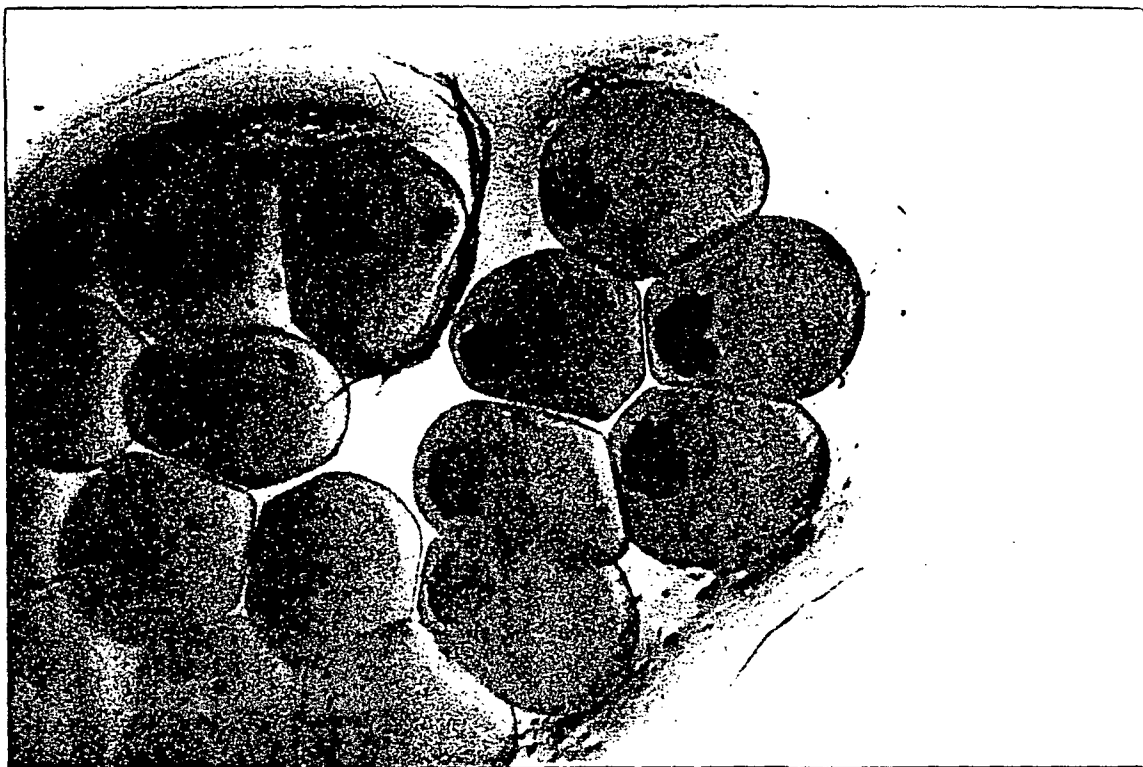


Plate 3c. Coagulation at hippo stage. *A. cummingii*

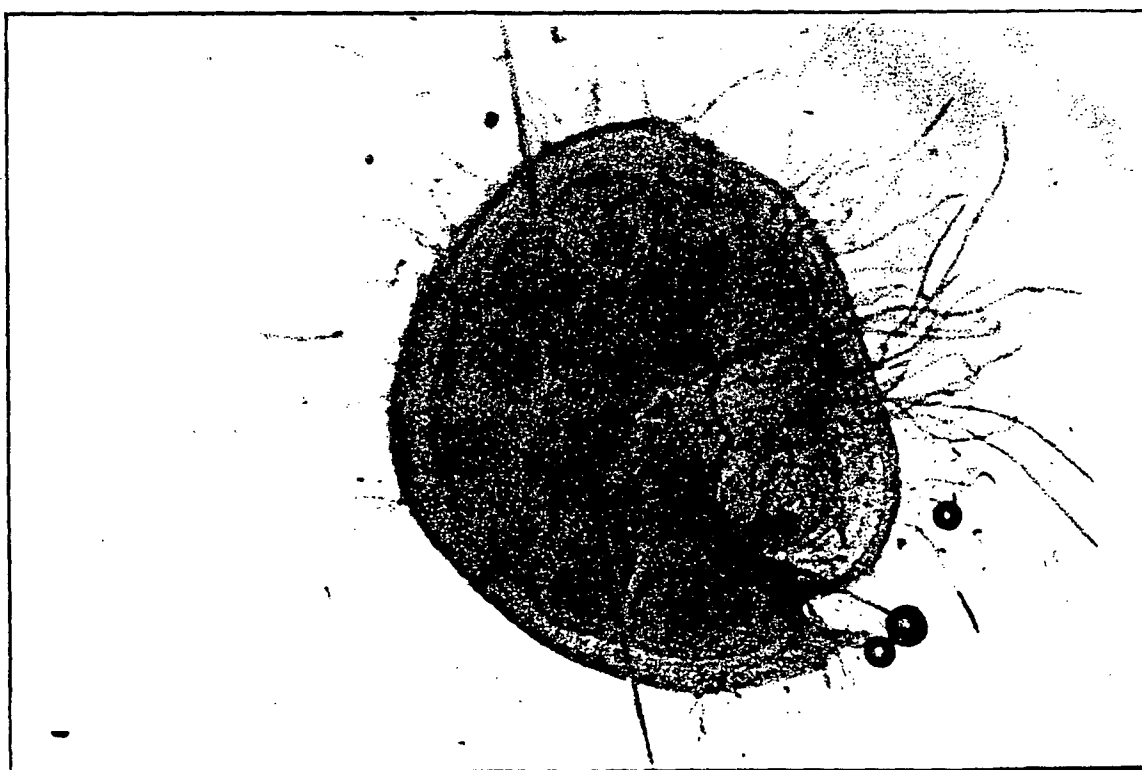
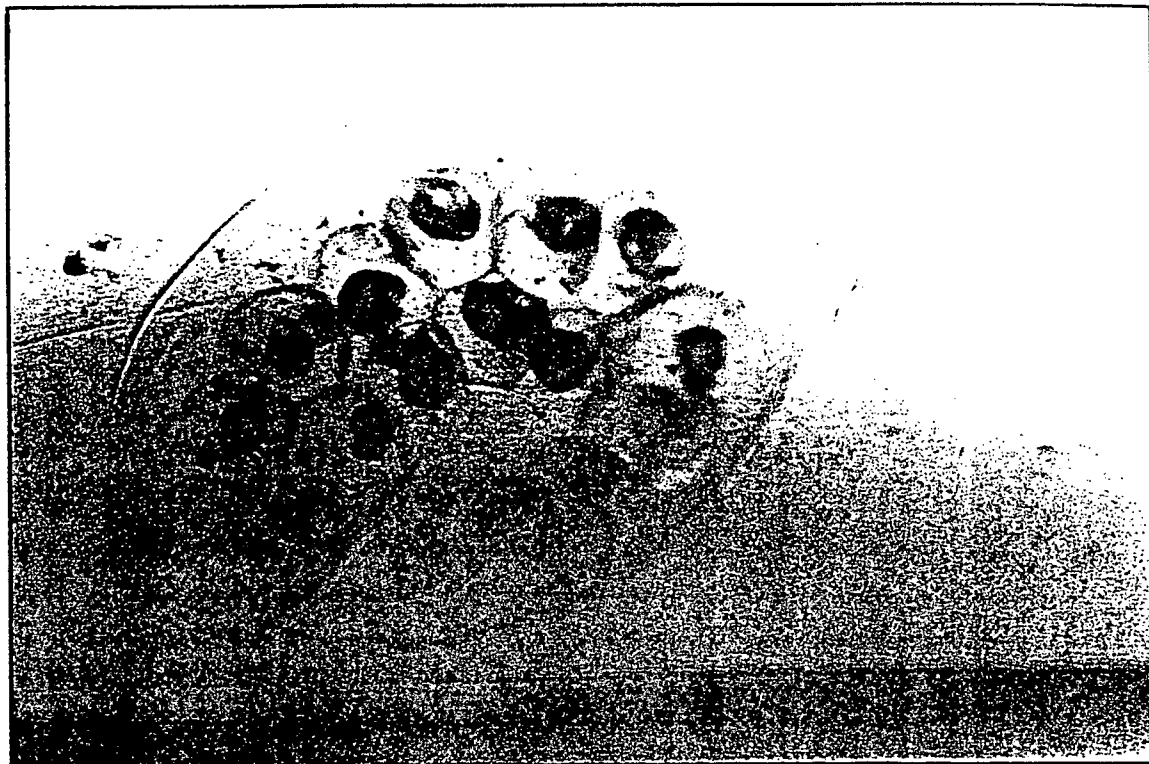
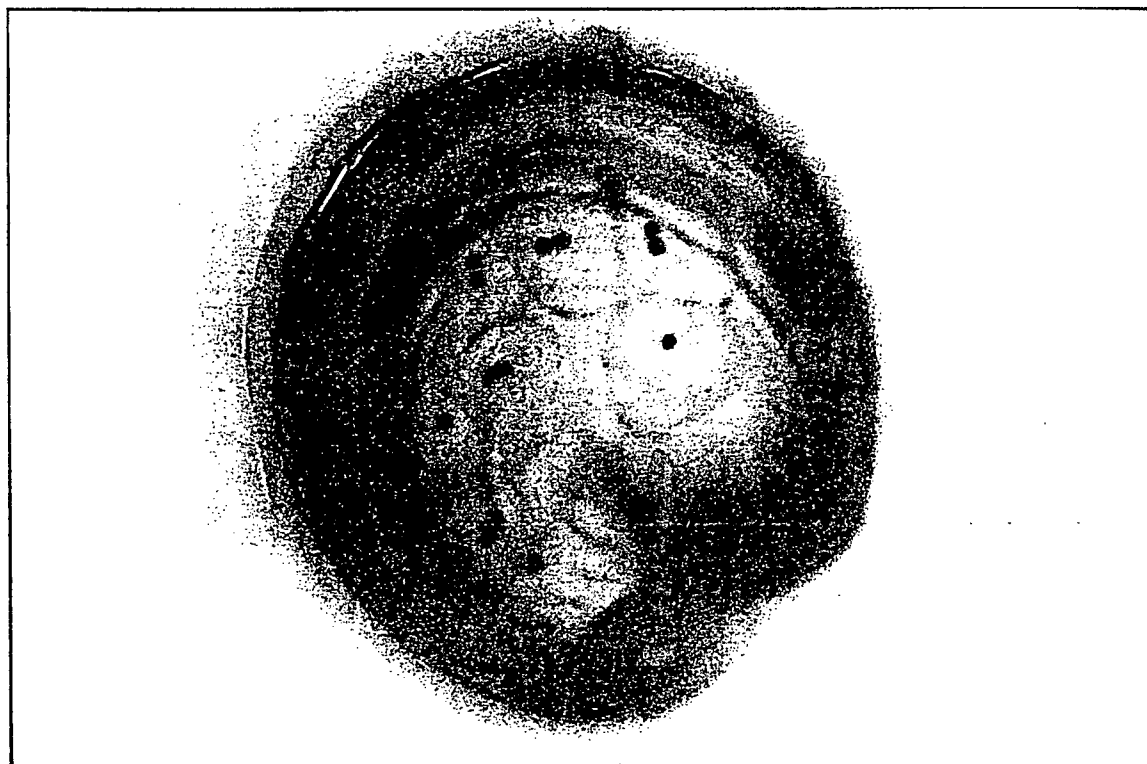


Plate 3d. Fungal Infection. *Glyptoplysa* sp.



**Plate 3a. Normal and ghost hatchlings. *A. carinata***



**Plate 3b. Polyvitelliny. *A. carinata***

## TOXICITY TRIALS

### SCHEDULE 1 CONDUCT OF TOXICITY TRIALS

Trial no.	Endpoints tested	Species tested	Toxicant regime
1	Developmental period	<i>A. carinata</i>	Control
	Juvenile mortality	<i>A. cumingii</i>	0.3% RP4
		<i>Glyptophysa</i> sp.	1.0% RP4
			3.2% RP4
			10% RP4
			32% RP4
Commenced 18/2/91		Terminated 27/2/91	
2	Developmental period	<i>A. cumingii</i>	Control
	Embryonic mortality	<i>Gyraulius</i> sp.	1.0% RP2
	Juvenile mortality	<i>Lymnaea</i> sp.	3.2% RP2
			10% RP2
			32% RP2
			100% RP2
Commenced 28/2/91		Terminated 12/3/91	
3	Developmental period	<i>A. carinata</i>	Control
	Embryonic mortality	<i>A. cumingii</i>	1.0% RP2
	Juvenile mortality	<i>Glyptophysa</i> sp.	3.2% RP2
			10% RP2
			32% RP2
			100% RP2
Commenced 13/3/91		Terminated 25/3/91	
4	Developmental period as:	<i>A. carinata</i>	Control
	pre-hatchling	<i>A. cumingii</i>	32% RP2
	hatchling	<i>Glyptophysa</i> sp.	42% RP2
	total		56% RP2
			75% RP2
			100% RP2
Commenced 17/3/91		Terminated 31/3/91	pto

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**SCHEDULE 1 contd CONDUCT OF TOXICITY TRIALS**

Trial no.	Endpoints tested	Species tested	Toxicant regime
5	Egg mass laying	<i>A. carinata</i>	Control
	Egg laying	<i>A. cumingii</i>	1.0% RP2
			3.2% RP2
			10% RP2
			32% RP2
			100% RP2
Commenced 31/3/91		Terminated 7/4/91	
6	Egg mass laying	<i>A. carinata</i>	Control
	Egg laying	<i>A. cumingii</i>	1.0% RP2
	Adult weight change	<i>Glyptophysa</i> sp.	3.2% RP2
	Juvenile mortality		10% RP2
	Control reared		32% RP2
	juvenile mortality		100% RP2
Commenced 12/4/91		Terminated 29/4/91	
Known parent			
	Fertility	<i>A. carinata</i>	32% RP2
	Juvenile mortality	<i>A. cumingii</i>	
Commenced 11/4/91		Terminated 13/4/91	
7	Egg mass laying	<i>A. carinata</i>	1.0% RP2
	Egg laying	<i>A. cumingii</i>	17ppb totU
	Juvenile mortality		10% RP2
	Control reared		170ppb totU
	juvenile mortality		32% RP2
			540ppb totU
Commenced 1/5/91		Terminated 15/5/91	
Algae/size	Juvenile mortality	<i>A. carinata</i>	32% RP2
		<i>A. cumingii</i>	
Commenced 2/5/91		Terminated 14/5/91	

### **Water collection**

Control water was collected from Magela Ck, upstream from the Ranger outlet pipe at the Georgetown creekside monitoring site. This was pumped into a 2,000L tanker and transported back to the lab. This tanker, pump and hoses was used exclusively for collecting control water. The retention pond waters were collected similarly, using a tanker, pump and hoses expressly for that purpose, or in an acid washed, 500L polythene tank, using the same pump and hoses collected, the water was transferred via the appropriate pump and hoses from the tankers into 3,000L polythene storage tanks. These had been thoroughly scrubbed and rinsed, and were flushed out between trials with town supply water. The stored water was aerated with a rapidly bubbling aquarium stone, using oil free compressed air. It was assumed that this would keep redox precipitation of manganese to a minimum, but this was not tested at any stage.

The toxicity testing was carried out using the apparatus depicted in Diagram 1 and Plate 1. During trials, the header tanks were refilled daily with freshly prepared solutions of designated dilution water. Water from each 20L header tank flowed by gravity into two 4L replicate testing chambers. In Trial #1, the header tanks were aerated with oil free compressed air, but in subsequent trials, the replicate chambers were aerated. Dissolved oxygen concentrations in vials covered with the smaller mesh were measured and observed to be near saturation. Flow rate to each chamber was 7mL per minute. This resulted in around 93% replacement in 24hrs.

During trials, pH, dissolved oxygen concentration and conductivity were measured daily using electronic instruments. The pH of test waters was difficult to determine precisely because of the poorly buffered nature of Magela Ck water. A standard time interval (15 seconds) passed between the time at which the electrode was placed in the solution sample and the time at which the pH valve was read.

Uranium and manganese concentrations were analysed for Trials 1,2, 3, 5, 6, and 7. Uranium was measured by Scintrex Time Delay Fluorimetric technique and manganese was measured by Graphite Flameless Atomic Absorption Spectroscopy.

Whole snails and dissected tissue which had been exposed to a range of retention pond water concentrations were preserved for electron microscopy and EDS (Energy Dispersive X-ray Spectroscopy) analysis. Dissection of organs and tissues was carried out using diagrams in Barnes The preservation solution was a double phosphate buffered glutaraldehyde solution.

## ELECTRON MICROSCOPY AND EDS

	Treatment	Time
TEM series	Phosphate buffer 0.1M/1%OsO <sub>4</sub>	1hr
	Phosphate buffer	3X5min. washes
Dehydration for EDS and TEM	50% acetone	2X5min.
	70% acetone	1X5min.
	0.5% Uranyl acetate/70% acetone	1X60min.
	(TEM only)	
	70% acetone (EDS only)	1X60min.
	90% acetone	2X10min.
	Abs acetone	2X10min.
	Propylene oxide	2X10min.
	Propylene oxide:Spurrs (film)	2hr
	Cured overnight at 70°C	

Sections were cut with a Reichert - Jung Ultracut E microtome to approximately 80nm. These were stained with Reynolds lead citrate for 10min and #5 uranyl acetate for 10 min. The instrument was a Phillips EM300 set at 60kV. This was fitted with a goniometer and minilens. X-rays generated from the unstained specimens were collected in an EDAX 9100 detector. Background radiation was minimised by a beryllium ring insert. The analogue signal was converted to digital for display and analysis.

Operating voltage	60kV.
Probe diameter	Approx. 1µm.
Count live time	200 sec.
Goniometer angle	35°



## DETAILS OF TRIALS

### Procurement of egg masses

Diagram 3 represents the continuous flow dosing apparatus used throughout most of the trials. For Trials #1 through to #4, egg masses were procured by confining adult pairs in the manner described for a period of 24hrs in control water. An initial suppression of oviposition following confinement was often observed. In order to obtain sufficient embryos (>20 per sample) it was necessary to confine at least twice the number of pairs than samples required. At the start of the trial, the embryos were mostly at the gastrula stage, although some at the blastula stage were also used. Before exposure, the position on the vial wall, the developmental stage and embryo number of each egg mass were determined. Each egg mass was then observed daily and the numbers of embryos at each developmental stage were noted.

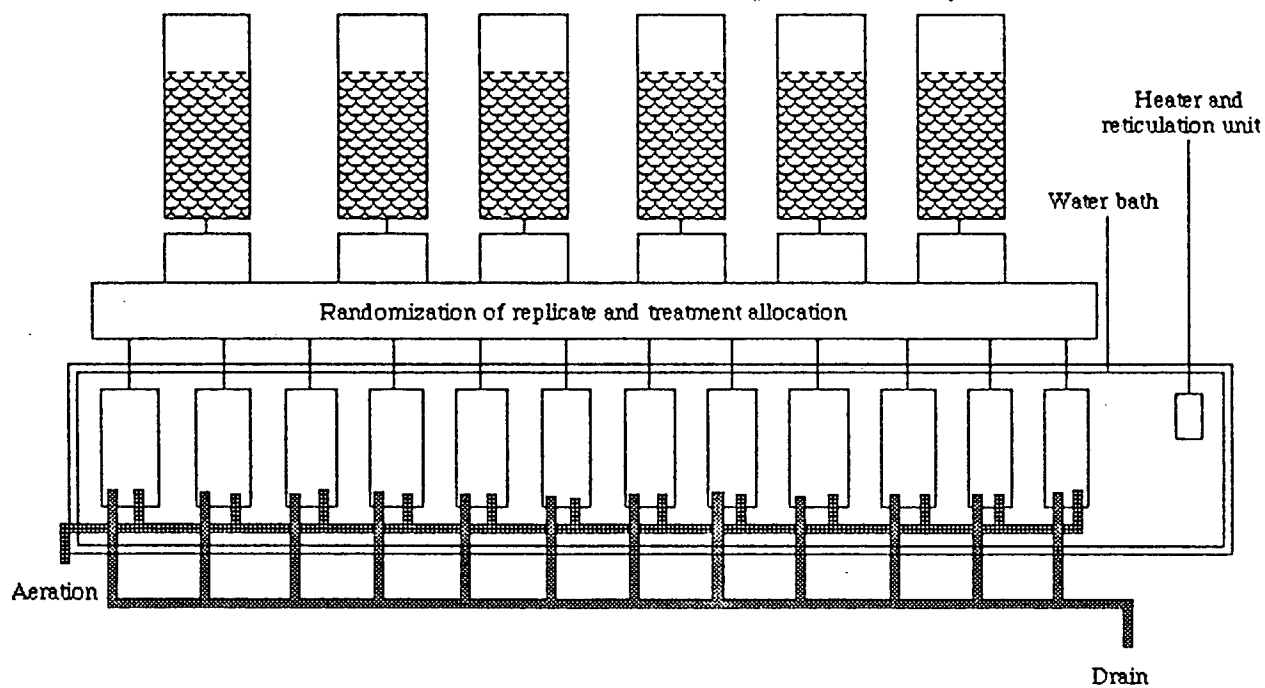
When it was necessary to use more than one egg mass per sample, egg masses of similar developmental stage were sought. The vials were distributed across replicates such that similar numbers of embryos were exposed to each treatment.

The developmental period was the interval from the first day of exposure (day 1=1 day of development) until observation of egg mass rupture. For developmental endpoints, termination of the trials occurred when either all egg masses had ruptured (hatching), or it was obvious that rupture would not occur in those still intact. In the appendices, the latter circumstance is denoted by "Termination". Exposure of adults was over an arbitrary period: 7 days for Trial #5; 5 days for Trials #6 and #7.

In Trials 1,2,3,6,7 and 'Known parent', juvenile exposure was performed on neonates hatched from exposed egg masses. Trials 6,7 and Algae/size used neonates hatched from egg masses that had been laid and reared in control water. At the observation of egg mass rupture, neonates were exposed for a 48hr period. Trial #7 included both 48 and 96hr exposure. Mortality was determined by subtracting the number of alive at the end of the exposure period from the numbers expected to hatch, as indicated by the last prerupture count. In the case of moribund individuals, the criterion of survival was the retraction of the foot in response to prodding by a dissection needle. Numbers of dead neonates did not appear to be a reliable measure of mortality, possibly because of rapid decay.

### DIAGRAM 3. THE DOSING APPARATUS.

20l header tanks - 5 toxicant concentrations and 1 control, allocated randomly.



For Trials #1 through to #3, 2 vials per replicate were exposed to each treatment. Because of difficulties in pooling data within and amongst replicates, this was increased to 3 vials per replicate in Trials #5 through to #7. Unless otherwise, all tests and control rearing were carried out at 30°C and aeration was by oil free compressed air. A representation of the continuous flow dosing apparatus is shown in Diagram 3.

#### **Trial #1**

Trial #1 was undertaken to assess the sensitivity of *A. carinata*, *A. cumingi* and *Glyptophysa* sp. The sensitivity of developmental retardation and juvenile mortality in *A. carinata* were observed using RP4 water, RP2 water was used in all subsequent trials, with a 100% treatment replacing the 0.3% treatment.

#### **Trial #2**

Trial #2 was undertaken partly to address the problems encountered in Trial #1. This trial also involved the exposure of control laid egg masses. Egg masses were derived from *A. cumingii*, *Gyraulus* sp. and *Lymnaea* sp.. The endpoints examined were the same as Trial #1, but more care was taken in following the embryos through their development. Embryonic mortality in egg masses was quantified at the various stages by daily counts.

#### **Trial #3**

Trial #3 was essentially a repeat of Trial #1 using the altered toxicant regime. The species tested were *A. carinata*, *A. cumingii* and *Lymnaea* sp.. The endpoints tested were Developmental period, Embryonic mortality and Juvenile mortality.

#### **Trial #4**

Trial #4 was undertaken to determine the toxicant concentration at which developmental retardation would first occur. Since a LOEC had previously been established at 100% RP2, a regime which bracketed 32% and 100% RP2 was used. This series descended by a factor of 0.75 between successive treatments. The trial used control laid egg masses and was conducted as a static test without treatment replication in 4L aquaria with daily half replacement.

#### **Trial #5**

In previous trials, a single vial each containing one *A. carinata* pair had been exposed to the treatments and an effect of egg laying depression had been observed. Trial #5 aimed to quantify this effect. The species tested were *A. carinata* and *A. cumingii*. Three pairs of snails of each species were used.

### **Known parent**

The 'Known parent' trial was conducted to investigate the influence that parentage might have on embryonic development and juvenile mortality during and after exposure of egg masses and juveniles to RP2 water. The species used were *A. carinata* and *A. cumingii*. Several pairs of snails were confined individually in vials in control water. Snails were observed daily and placed into new vials once egg masses had been deposited. The egg masses, from known parents, were then exposed to 32% RP2 and ensuing development and juvenile mortality recorded in the manner previously described.

### **Trial #6**

To determine the applicability of results from toxicity tests in the laboratory, trial #6 was conducted concurrently in creekside and laboratory trials using the same species and endpoints at each location. Flow rates in the creekside trial were 49mLs/min (cf 7mLs/min in the laboratory), and water temperature at this site was ambient Magela Ck water temperature, (28° to 32°C). Creek aeration was by oil-free compressed air.

### **Trial #7**

Trial #7 aimed to determine whether or not the observed effects of RP2 water were attributable solely to total uranium concentration. No control (creekwater) was run with this trial because of equipment limitations and, within the context of the experiment, a control was deemed unnecessary. Estimates of the total uranium levels of RP2 water were based on Scintrex determinations from previous trials. 100% RP2 was assumed to have a total U level of 1700ppb. Daily solutions using AR grade uranyl sulphate and Magela Ck water were made to correspond to 1% 10% and 100% RP2 total U levels. At the end of day 1, the 100% U treatment was replaced with a 32% treatment, following mortality of all *A. carinata* individuals in that treatment.

### **Algae/size**

The 'Algae/size' trial was carried out to determine whether the presence or absence of algae or the size of the vial, were contributing to variability observed in juvenile mortality. It was similar to the Known parent trial. Two sizes of vial were used: Small - 50mL clear plastic sample jars; large - 400mL sample jars. Vials were placed in the open in control water, and allowed to accumulate epiphytic algae for a few weeks. 20 neonates hatched in control water were used per sample and 4 samples used per treatment (large/small \* with/without algae). The species tested were *A. carinata* and *A. cumingii*. Snails were exposed to 32% RP2 in a 40L aquaria with daily half replacement and was for 48hrs.

## STATISTICAL ANALYSIS

Time limitations did not permit full statistical treatment of the data set. In particular Chi-squared testing for sample and replicate homogeneity was dispensed with.

Instead, data from replicates of each treatment were pooled and the data assumed to be normally disturbed for analysis of variance testing. The dangers in this approach involve an increased chance of committing a Type 2 error, where the null hypothesis of equality is not rejected when in fact differences do exist. However, in the context of the project, this conservative measure served to highlight significant differences.

Where analysis of variance testing indicated significant differences amongst the treatments of a trial, Dunnett's test was applied to the treatment means. Zar (1984) recommends that the number of replicates in the control treatment be the integer below the square root of the treatment degrees of freedom  $[(k-1)^{1/2}]$  times larger than the treatment replicate number. This was not done in these trials, and the discriminatory power of the statistical tests conducted on the data may not have been optimal.

Student-Newman-Keuls testing was employed when significant differences were found using a single treatment (eg fecundity parameters in the preliminary observations).

Statistical testing was performed at the 5% level. Although for regulatory purposes testing at the 10% significance level may be adopted (C. Humphrey pers comm, 1991), the assessment of species and endpoints at 5% makes the final distinctions more complete.

## RESULTS

### PRELIMINARY OBSERVATIONS #1 COMMENCED 19/1/91.

The first series of preliminary observations on snail reproduction and early development were carried out in the breeding tanks (conductivity around 80 $\mu$ S/cm). Tabulated data are shown in Appendix 1a. Table 1a shows statistics derived from the pooling of all replicate data. In the context of the project, the variation associated with these absolute values was as important as the means.

**Table 1a. Egg and egg mass means and coefficients of variation associated with the means.**

Data taken over five days. Commenced 19/1/91.

Species	No. of		Egg/egg mass		Eggmass/pair/day		
	Reps	n	Mean	C.V.%	n	Mean	C.V.%
<i>A. carinata</i>	4	41	13.4	53.0	20	1.85	89.1
<i>A. cumingii</i>	7	117	29.9	47.5	35	3.23	82.0
<i>Glyptophysa</i> sp.	4	55	9.36	33.7	16	3.50	101
<i>Gyraulus</i> sp.	4	150	6.55	39.1	20	3.78	55.3
<i>Helicorbis</i> sp.	4	83	4.66	36.1	20	2.1	83.3

Figures 1a to 1e display means across replicates for eggs per egg capsule. Means, the slopes of the cumulative daily means, and  $r^2$  values appear in Table 1b. These values were derived from daily means of pooled replicate data.

**Table 1b. Eggs per egg mass statistics.**

Using data across replicates over five days.

Species	Mean	Slope of line for cumulative data	( $r^2$ )
<i>A. carinata</i>	12.43 <sup>a</sup> 15.54 <sup>b</sup>	12.19 15.17	0.942 0.998
<i>A. cumingii</i>	29.62	28.32	0.998
<i>Glyptophysa</i> sp.	8.87	9.26	0.990
<i>Gyraulus</i> sp.	6.24	6.66	0.999
<i>Helicorbis</i> sp.	4.50	4.45	0.992

<sup>a</sup> Includes zero value of day 5. <sup>b</sup> Excludes zero value of day 5.

Correlation coefficients indicated a degree of predictability associated with egg numbers/capsule over the five days of observations. No species showed large variation except *A. carinata*, whose four replicate pairs did not oviposit on day 5. This species otherwise displayed similar variabilities to the others (Figures 1f to 1h). Discounting this observation showed *Helicorbis* sp. to be slightly more irregular in egg and egg mass production than the other species (Table 1b).

Figure 1f illustrates the variation associated with egg mass laying over four days across replicates. Analysis of variance performed on the coefficients of variation for each replicate over the days of observation revealed no significant difference between species ( $P>F=0.9305$ ). This was also the case for total daily egg production for each species/replicate (Figure 1g.), and for eggs per eggmass (Figure 1h.).

When replicates were pooled species differed in the mean values of eggs per egg mass and egg masses per day ( $P>F=0.016$  and  $0.002$  respectively), but no significant difference was found in eggs per day.

Table 1c presents results of Student-Newman-Keuls (SNK) testing on the significantly different parameters.

Table 1c. Results of comparisons from SNK testing.

Parameter	5 day replicate mean
<b>Egg masses/day</b>	
<i>Gyraulius</i> sp.	7.73
<i>Helicorbis</i> sp.	4.15
<i>A. cumingii</i>	3.19
<i>Glyptophysa</i> sp.	2.9
<i>A. carinata</i>	2.75
<b>Eggs/egg mass</b>	
<i>A. cumingii</i>	29.3
<i>A. carinata</i>	15.43
<i>Glyptophysa</i> sp.	8.95
<i>Gyraulius</i> sp.	6.36
<i>Helicorbis</i> sp.	5

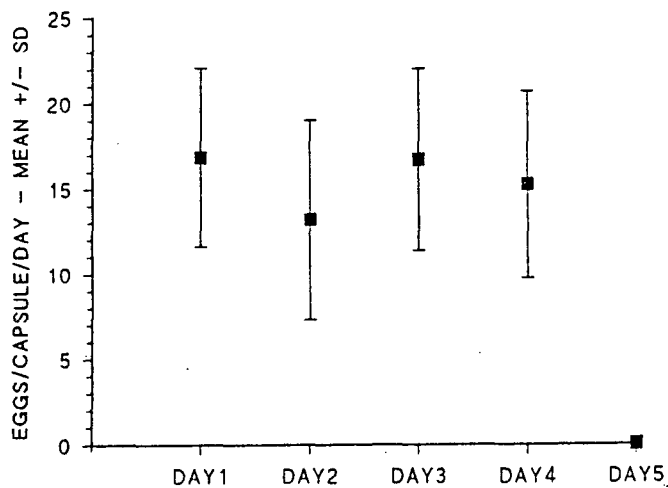


FIG. 1a SNAIL EGG LAYING CHARACTERISTICS  
AMERIANNA CARINATA COMM. 19/1/91

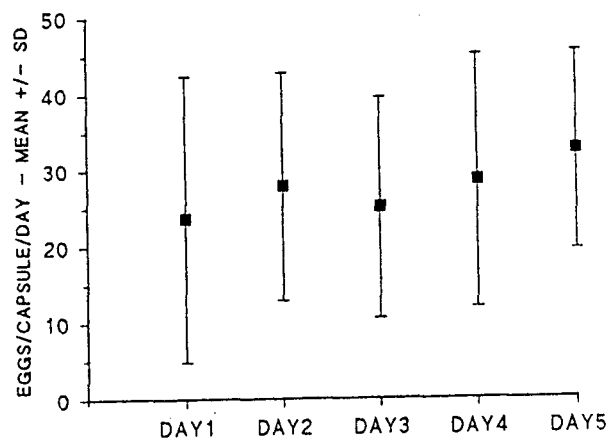


FIG. 1b SNAIL EGG LAYING CHARACTERISTICS  
AMERIANNA CUMMINGII COMM. 19/1/91

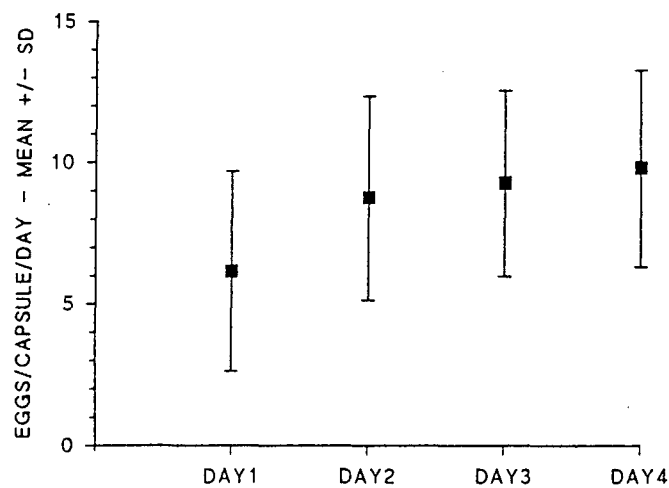


FIG. 1c SNAIL EGG LAYING CHARACTERISTICS  
GLYPTOPHYSA SP. COMMENCED 19/1/91

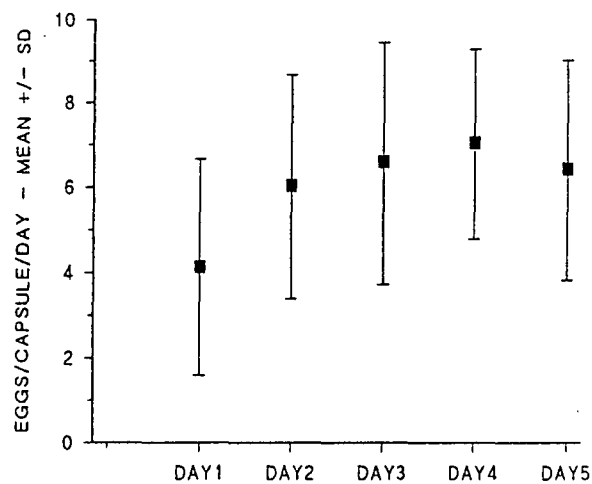


FIG. 1d SNAIL EGG LAYING CHARACTERISTICS  
GYRAULUS SP. COMMENCED 19/1/91

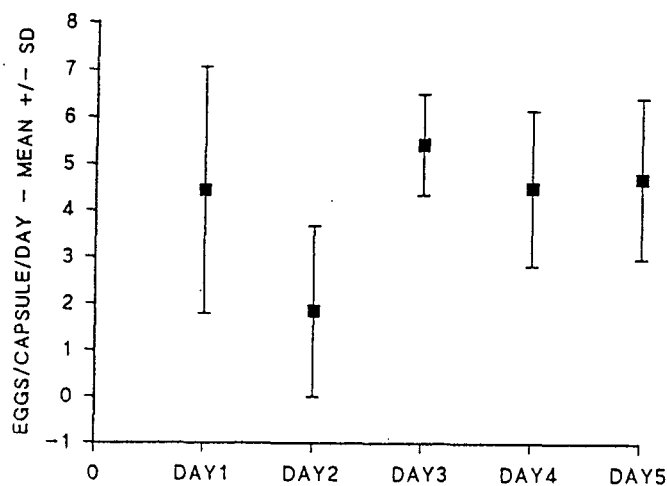


FIG. 1e SNAIL EGG LAYING CHARACTERISTICS  
HELICORBIS SP.



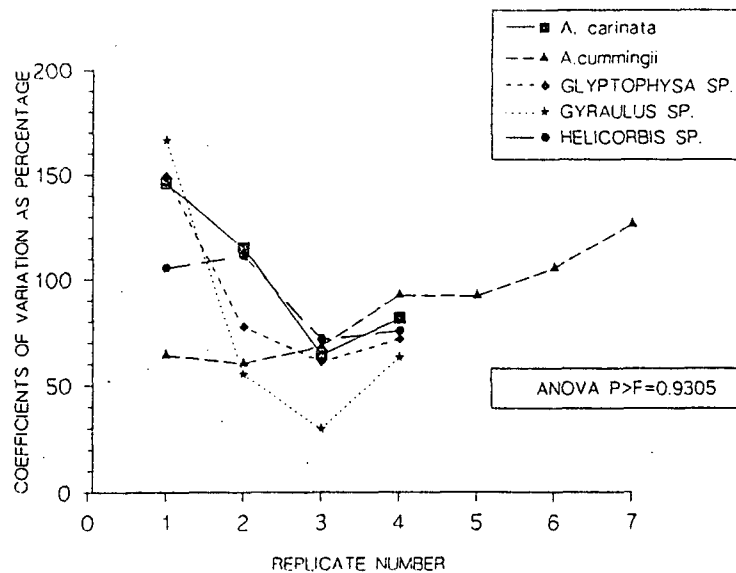


Fig. 1F EGG MASS LAYING COEFFICIENTS OF VARIATION COMM. 19/1/91

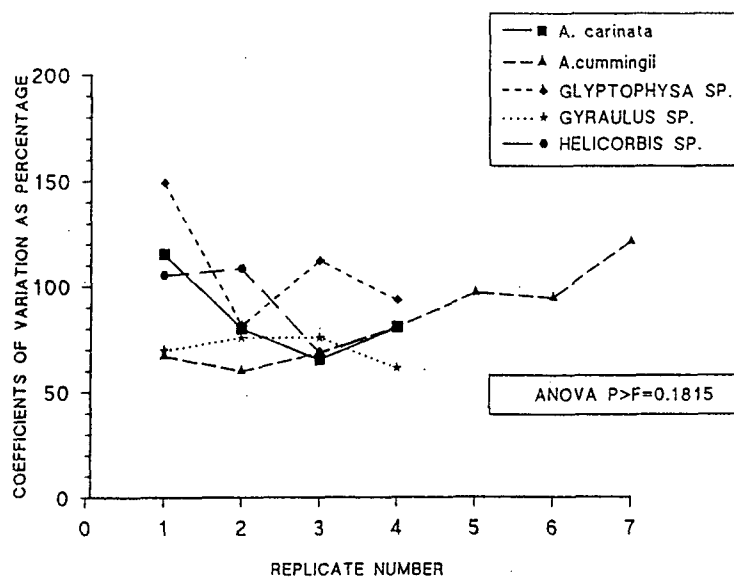


Fig. 1G EGG LAYING COEFFICIENTS OF VARIATION COMM. 19/1/91

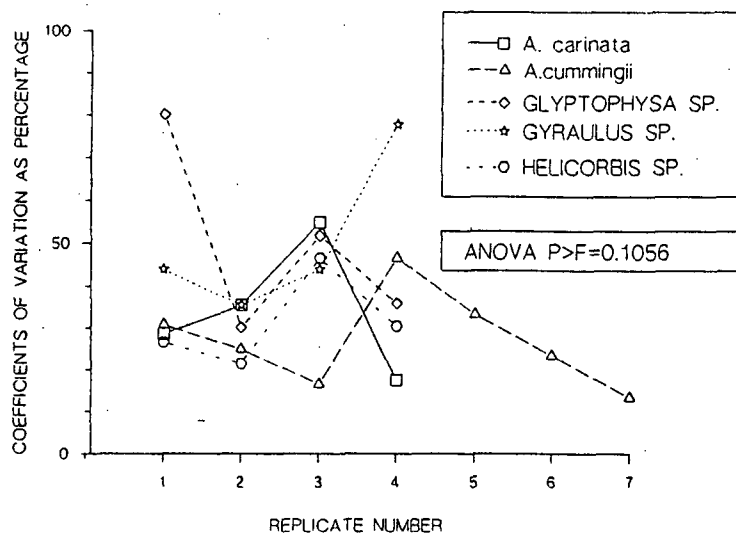


Fig. 1H EGG PER EGGMASS COEFFICIENTS OF VARIATION COMM. 19/1/91

## PRELIMINARY OBSERVATIONS #2 COMMENCED 25/1/91

These were performed using Magela Ck water (conductivity of around 20 usem/cm). Tabled data for these observations appear in Appendix 1b. Simple statistics appear in Table 2a. As in the case of Table 1a, these values were derived from pooled data of all replicates and all days.

**Table 2a. Egg and egg mass means and coefficients of variation.**

Data taken over five days. Commenced 25/1/91.

Species	Reps	No. of		Egg/egg mass		Eggmass/pair/day		
		n	Mean	C.V.%		n	Mean	C.V.%
<i>A. carinata</i>	4	46		3.51	22.8	20	2.3	49.1
<i>A. cumingii</i>	4	26		8.90	28.4	20	1.3	90.3
<i>Gabbia</i> sp.	4	17		0.24	22.9	20	0.85	203
<i>Glyptophysa</i> sp.	4	16		4.38	42.0	20	2.15	165
<i>Gyraulius</i> sp.	4	77		1.84	20.2	20	3.7	54.8
<i>Lymnaea</i> sp.	4	14		32.2	53.9	20	0.7	132

Means, the slopes of cumulative daily means, and  $r^2$  values of eggs per egg mass appear in Table 2b. These values were derived from daily means of pooled replicate data.

**Table 2b. Eggs per egg mass statistics.**

Using data across replicates over five days.

Species	Mean	Slope of line for cumulative data	( $r^2$ )
<i>A. carinata</i>	14.6	14.5	0.9997
<i>A. cumingii</i>	29.3	30.2	0.997
<i>Gabbia</i> sp.	1.02	1.03	0.9998
<i>Glyptophysa</i> sp.	8.65	6.89	0.970
<i>Gyraulius</i> sp.	9.16	9.20	0.9999
<i>Lymnaea</i> sp.	54.3	49.9	0.996

Analysis of variance was performed on replicate data summed over five days. The results of these analyses appear in Table 2c.

TABLE 2C. RESULTS OF ANALYSIS OF VARIANCE BETWEEN THE SIX SPECIES.

Probability values			
Parameter:	Egg masses/day	Eggs/day	Eggs/eggmass
Means	0.0001**	0.0017**	0.0006**
Coefficients of variation	0.0018**	0.0006**	0.0002**

Student Newman Kuels testing was performed on the significantly different data. The results of this appear in Table 2d.

Table 2d. Results of comparisons from SNK testing.

Parameter	5 day replicate means
EGG MASSES PER DAY.	

Means

<i>Gyraulus</i> sp.	3.75
<sup>3</sup> <i>A. carinata</i>	2.30
<sup>3</sup> <i>A. cumingii</i>	1.30
<sup>3</sup> <i>Glyptophysa</i> sp.	0.80
<sup>3</sup> <i>Gabbia</i> sp.	0.70
<sup>3</sup> <i>Lymnaea</i> sp.	0.70

Coefficients of variation

<i>Gabbia</i> sp.	165
<i>Glyptophysa</i> sp.	153
<i>Lymnaea</i> sp.	111
<i>A. cumingii</i>	96.3
<i>A. carinata</i>	50.4
<i>Gyraulus</i> sp.	45.2

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Table 2d. contd. Means comparison from SNK testing.

Parameter: 5 day replicate means

EGGS/DAY

Means

<i>Lymnaea</i> sp.	41.9
<i>A. cumingii</i>	40.8
<i>A. carinata</i>	35.5
<i>Gyraulus</i> sp.	34.0
<i>Glyptophysa</i> sp.	8.35
<i>Gabbia</i> sp.	0.90

Coefficients of variation

<i>Lymnaea</i> sp.	126
<i>Glyptophysa</i> sp.	40.2
<i>A. cumingii</i>	25.3
<i>A. carinata</i>	21.8
<i>Gyraulus</i> sp.	19.6
<i>Gabbia</i> sp.	11.0

EGGS/EGGMASS

Means

<i>Lymnaea</i> sp.	80.3
<i>A. cumingii</i>	31.4
<i>A. carinata</i>	15.3
<i>Glyptophysa</i> sp.	10.4
<i>Gyraulus</i> sp.	9.33
<i>Gabbia</i> sp.	1.05

Coefficients of variation

<i>Glyptophysa</i> sp.	178
<i>Gabbia</i> sp.	170
<i>Lymnaea</i> sp.	126
<i>A. carinata</i>	51.2

<i>A. cumingii</i>	98.5
<i>Gyraulus</i> sp.	42.1

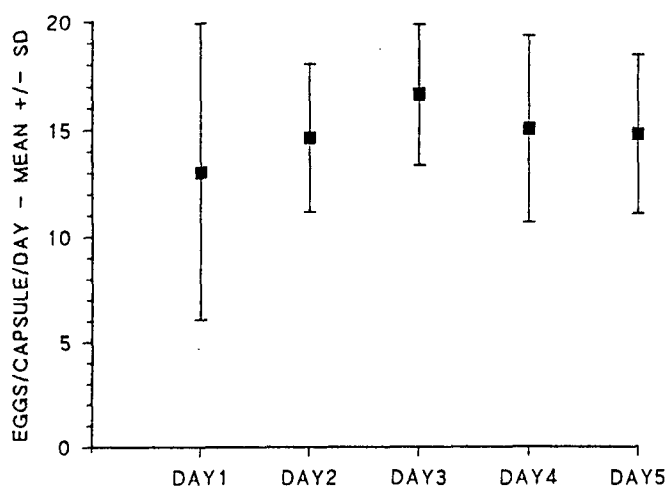


FIG. 2a SNAIL EGG LAYING CHARACTERISTICS  
AMERIANA CARINATA COMM. 26/1/91

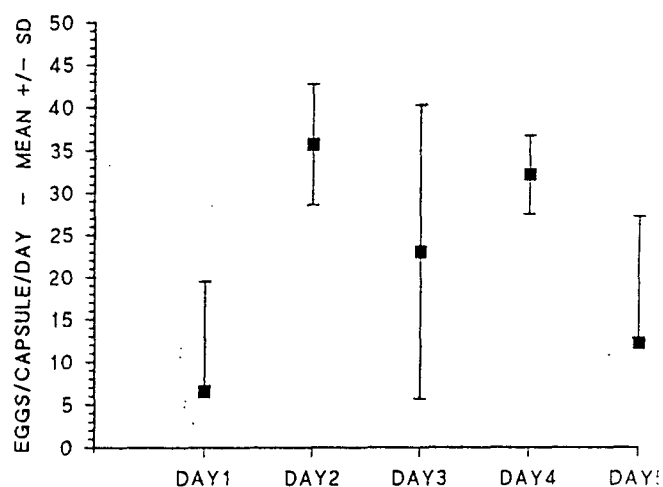


FIG. 2b SNAIL EGG LAYING CHARACTERISTICS  
AMERIANA CUMMINGII COMM. 26/1/91

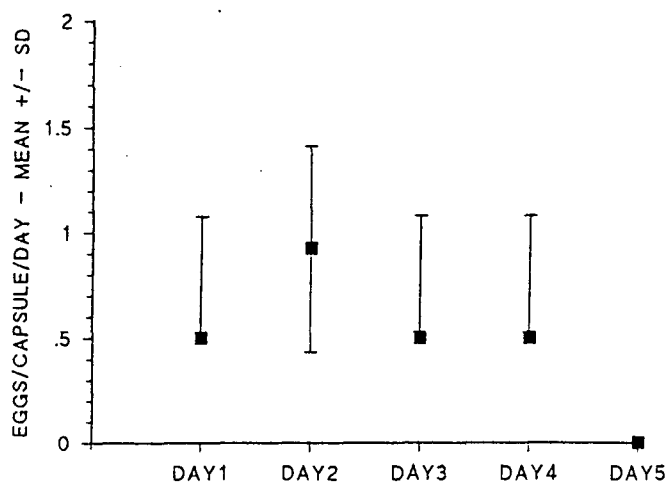


FIG. 2c SNAIL EGG LAYING CHARACTERISTICS  
GABBIA SP. COMMENCED 26/1/91

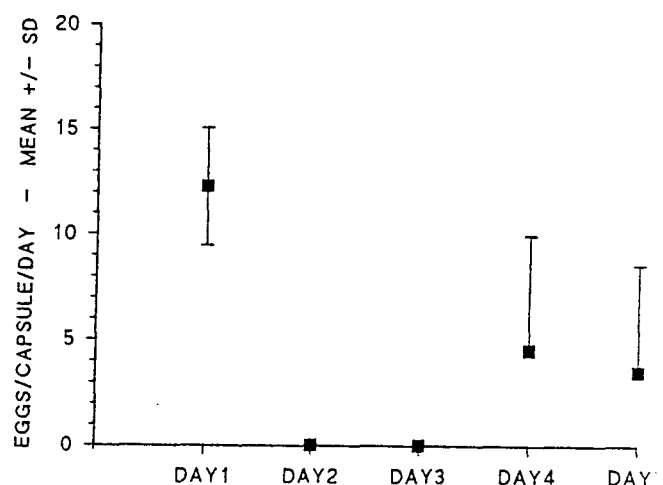


FIG. 2d SNAIL EGG LAYING CHARACTERISTICS  
GLYPTOPHYSA SP. COMMENCED 26/1/91

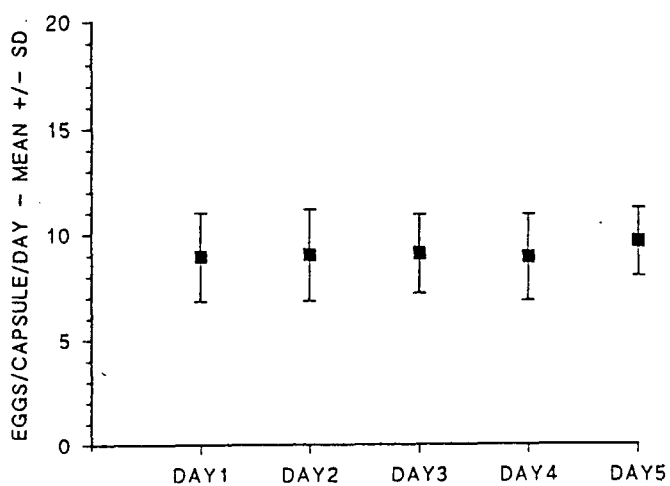


FIG. 2e SNAIL EGG LAYING CHARACTERISTICS  
GYRAULUS SP. COMMENCED 26/1/91

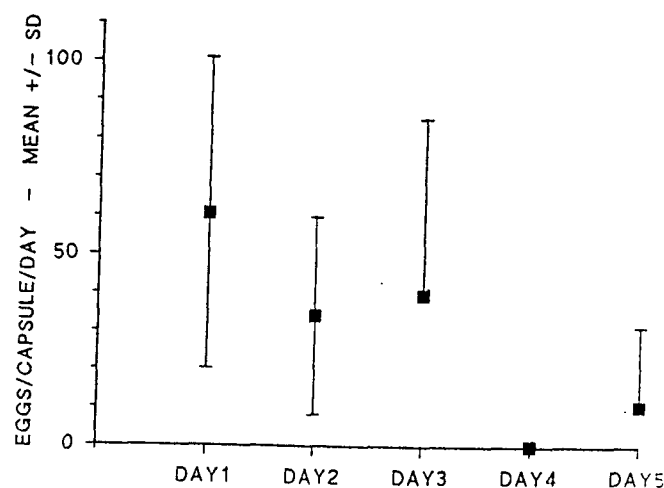


FIG. 2f SNAIL EGG LAYING CHARACTERISTICS  
LYMNAEA SP. COMMENCED 26/1/91

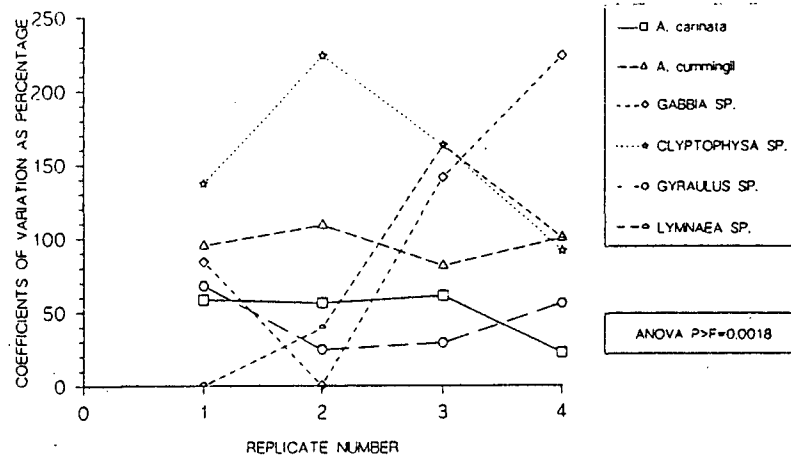


Fig. 2G EGG MASS COEFFICIENTS OF VARIATION COMM. 25/1/91

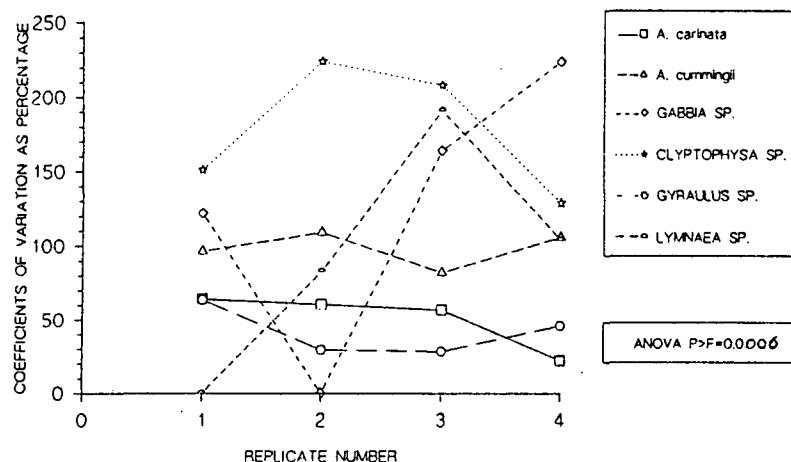


Fig. 2H EGG LAYING COEFFICIENTS OF VARIATION COMM. 25/1/91

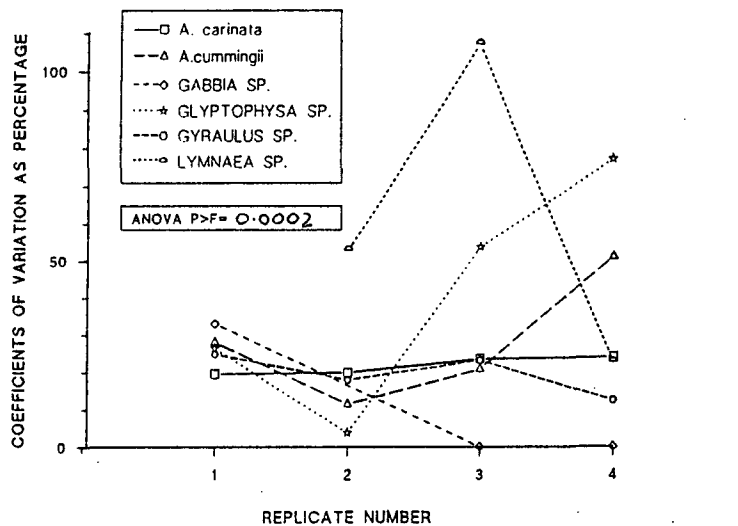


Fig. 2I EGG PER EGGMASS COEFFICIENTS OF VARIATION COMM. 25/1/91

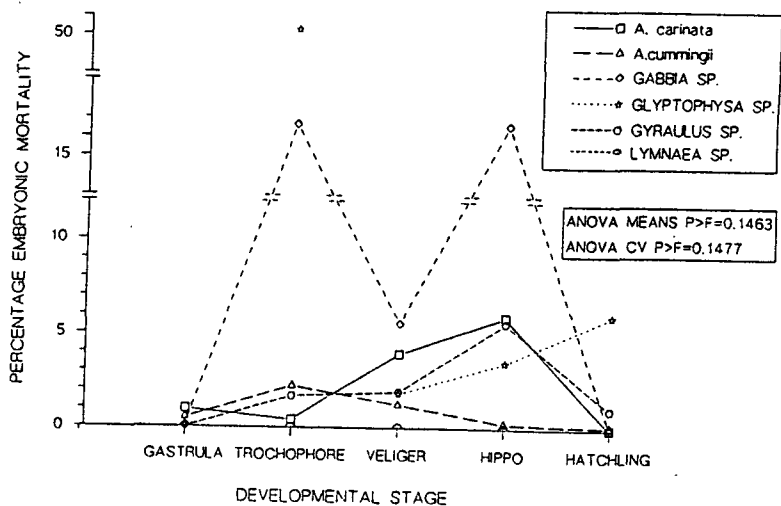


Fig. 2J PERCENTAGE EMBRYONIC MORTALITY COMM. 25/1/91

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One way analyses of variance performed on embryonic mortality across embryonic stages and species failed to reveal significant differences ( $P>F=0.1463$  and  $P>F=1477$  respectively).

The difference in results between the first and second series of observations could be due to water quality. In the first series, the vials were always at the bottom of the rearing tanks, and although these were aerated, oxygen levels may have been sub optimal. The second series was performed in 20L aquaria, where the aeration produced levels near saturation. Also, as previously stated, conductivities in the two trials were different. In many ways the first trial was an exercise in familiarisation, whilst the second was more relevant to later toxicity testing, which used Magela Ck water as the control.

From the results, it was concluded that *Gyraulus* sp., *A. carinata* and lesser extent *A. cumingii* showed least variation in the parameters measured. The least variable species was *Gyraulus* sp. However the difficulties in handling and rearing this species did count against it finally for use as a monitoring animal. It strongly maintained its hold on surfaces and was often damaged during routine handling. Also, the stock population underwent a decrease once regular handling of large numbers of individuals commenced. Prior to this decrease, it had been bred to numbers sufficient for toxicity testing. Similar problems occurred more severely with *Helicorbis* sp., and in the first trial it showed a larger, but non-significant variability in comparison to the other species. *A. carinata* and *A. cumingii* were quite robust, easily handled and maintained.

Particularly *Gabbia* sp., but also *Glyptophysa* sp., and *Lymnaea* sp. were seen to be variable in their fecundity parameters. *Gabbia* sp. along with *Helicorbis* sp. were not included in further trials. *A. carinata*, *A. cumingii*, *Glyptophysa* sp., *Gyraulus* sp. and *Lymnaea* sp. were tested further.

<b>TRIAL #1 COMMENCED 18/2/91</b>
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Trial #1 tested *A. carinata*, *A. cumingii* and *Glyptophysa* sp. The endpoints examined were developmental retardation and juvenile mortality. Full details of testing are presented in Schedule 1. Results of one way analysis of variance performed on pooled replicate data are given in Table 3a. Figures 3a to 3f present means and standard deviations for the species and endpoints assessed. Full results are in Appendix 11.1.a and 11.1.b.

**Table 3a. Anova results for Trial #1**

Species	Endpoint	P>F	Conclusion
<b>Days to hatching</b>			
<i>A. carinata</i>		0.1491	No significant difference
<i>A. cumingii</i>		0.6071	No significant difference
<i>Glyptophysa</i> sp.		0.8058	No significant difference
<b>Juvenile mortality</b>			
<i>A. carinata</i>		0.9122	No significant difference
<i>A. cumingii</i>		0.3855	No significant difference
<i>Glyptophysa</i> sp.		0.5757	No significant difference

The results from this trial indicated that neither endpoint was sensitive at the concentrations used. It was determined to remove the 0.3% RP2 treatment and use a 100% RP2 treatment as the most concentrated treatment.



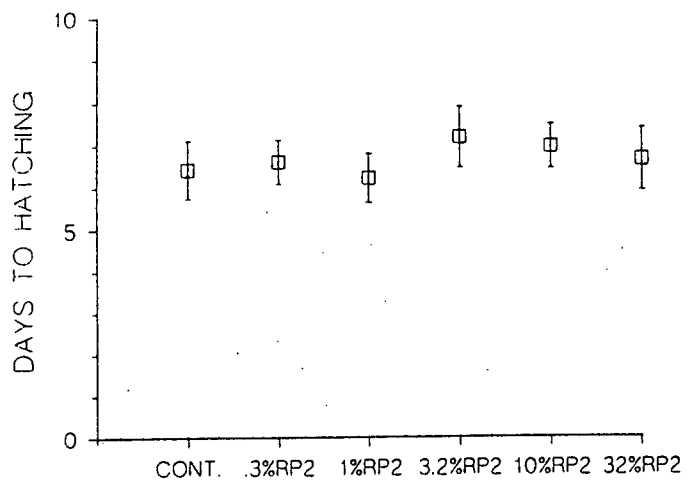


Fig. 3a Trial #1 Days to hatching  
*A. carinata*

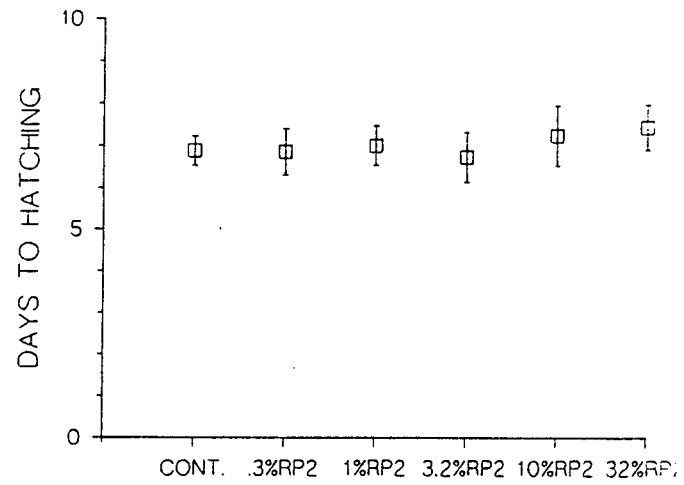


Fig. 3b Trial #1 Days to hatching  
*A. cummingii*

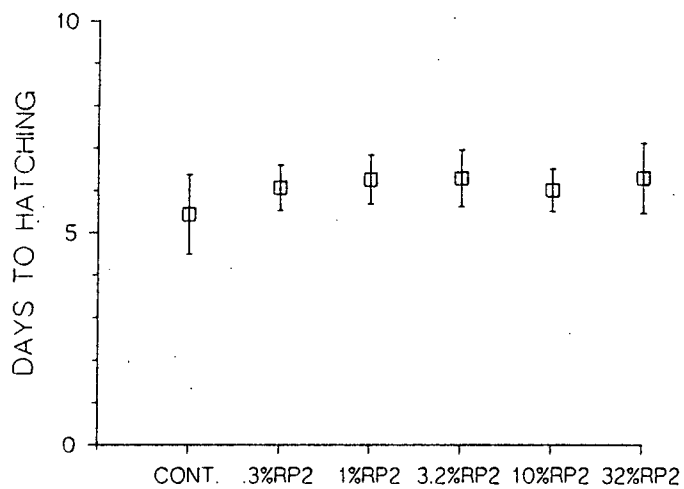


Fig. 3c Trial #1 Days to hatching  
GLYPTOPHYSA sp.

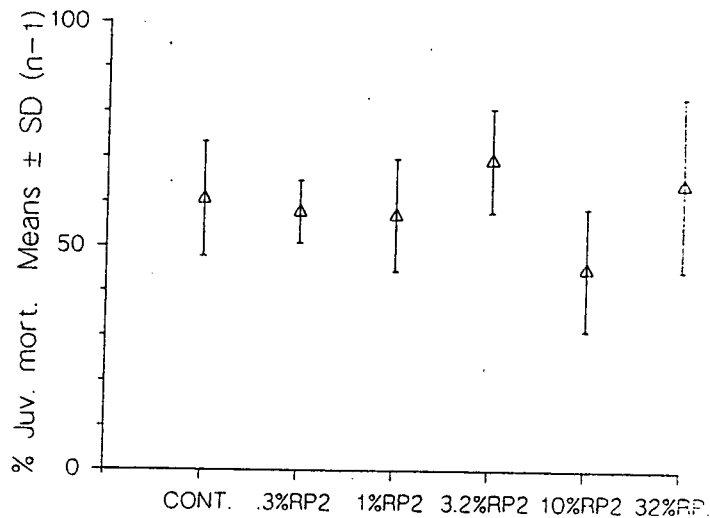


Fig. 3d Trial #1 Percentage juvenile mortality  
*A. carinata*

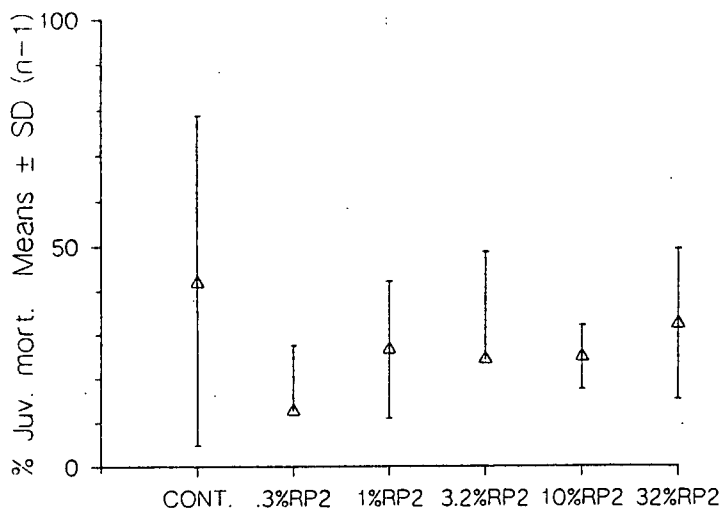


Fig. 3f Trial #1 Percentage juvenile mortality  
GLYPTOPHYSA sp.

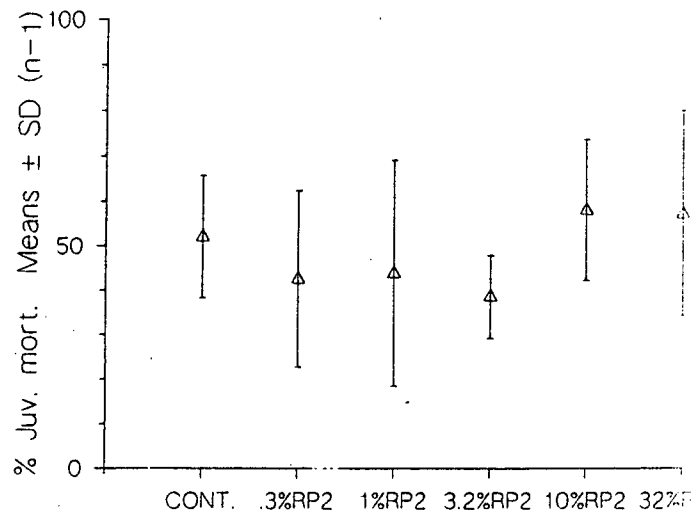


Fig. 3e Trial #1 Percentage juvenile mortality  
*A. cummingii*

**TRIAL #2 COMMENCED 28/2/91**

Trial #2 tested *A. cumingii*, *Gyraulius* sp. and *Lymnaea* sp. The endpoint examined were developmental retardation and embryonic mortality. Tabled data appears in Appendix 3 and full results of statistical analysis appear in Appendix 11.2. Results of analysis of variance performed on pooled replicate data are presented in Table 4a.

**Table 4a. Results of analysis of variance. Trial #2.**

Species	Endpoint	P>F	Conclusion
<b>Developmental retardation</b>			
<i>A. cumingii</i>		0.308	No significant difference
<i>Gyraulius</i> sp.		0.0001	Significantly different
<i>Lymnaea</i> sp.		0.0001	Significantly different
<b>Embryonic mortality</b>			
<i>A. cumingii</i>		0.6490	No significant difference
<i>Gyraulius</i> sp.		0.082	No significant difference
<i>Lymnaea</i> sp.		0.619	No significant difference

Significantly different results were subjected to Dunnett's testing. The results are summarised in Table 4b, and appear in full in Appendix 11.2.c.

**Table 4b. Results of Dunnett's testing. Trial #2.**

Species	Endpoint	NOEC	LOEC
<b>Developmental retardation</b>			
<i>Gyraulius</i> sp.		32% RP2	100% RP2
<i>Lymnaea</i> sp.		32% RP2	100% RP2

From these results it was seen that developmental retardation was a more sensitive endpoint than direct mortalities to the growing embryos. The effects of RP2 water in terms of developmental retardation were apparent at 100% concentration.

<b>TRIAL #3 COMMENCED 13/1/91</b>
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Trial #3 was conducted in a similar manner to Trial #2. In order to relate the results, *A. cumingii* was included, along with *A. carinata* and *Glyptophysa* sp. Observations on developing egg masses were not carried out on days 2 and 3, and the first developmental class included gastrula, trochophore, veliger and hippo stages. Tables results appear in Appendix 4. Analysis of variance was carried out on pooled data. Full results appear in Appendix 11.3. Table 5a presents a summary of these.

**Table 5a Results of analysis of variance. Trial #3.**

Species	Endpoint	P>F	Conclusion
<b>Developmental retardation</b>			
<i>A. carinata</i>		0.0011	Significantly different
<i>A. cumingii</i>		0.027	Significantly different
<i>Glyptophysa</i> sp.		0.0105	Significantly different
<b>Embryonic mortality</b>			
<i>A. carinata</i>	Treatment	0.1310	No significant diff.
	Stage	0.0872	No significant diff.
	Interaction	0.3582	No significant int'ion.
<i>A. cumingii</i>	Treatment	0.2682	No significant diff.
	Stage	0.7719	No significant diff.
	Interaction	0.9734	No significant int'ion.
<i>Glyptophysa</i> sp.	Treatment	0.8022	No significant diff.
	Stage	0.1252	No significant diff.
	Interaction	0.5397	No significant int'ion.
<b>Juvenile mortality</b>			
<i>A. carinata</i>		0.0004	Significantly different
<i>A. cumingii</i>		0.0001	Significantly different
<i>Glyptophysa</i> sp.		0.017	Significantly different

Results of Dunnett's testing appears in Appendix 11.3 and is summarised in Table 5b.

**Table 5b Results of Dunnett's testing. Trial #3.**

Species	Endpoint	NOEC	LOEC
<b>Developmental retardation</b>			
<i>A. carinata</i>		32%	100%
<i>A. cumingii</i>		100%	undefined
<i>Glyptophysa</i> sp.		100%	undefined
<b>Juvenile mortality</b>			
<i>A. carinata</i>		32%	100%
<i>A. cumingii</i>		32%	100%
<i>Glyptophysa</i> sp.		100%	undefined

The species which showed undefined LOECs must derive their significant anova terms from treatments producing shorter hatching periods and lower juvenile mortality than control. For *Glyptophysa* sp., 32% juvenile mortality was significantly different from control, but because 100% mortality was not significantly different, the LOEC remained undefined. Effects are displayed in Figures 4a and 4b. From this trial *A. carinata* was seen to be more sensitive than the two other species tested, although *A. cumingii* was seen to be as sensitive in terms of juvenile mortality.

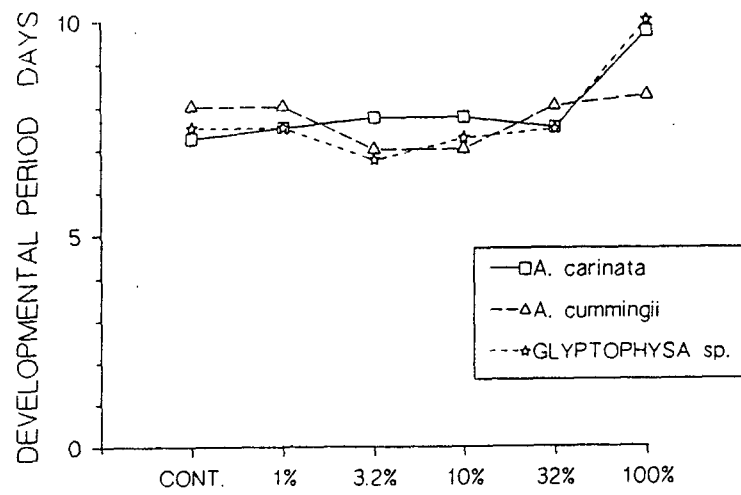


FIG 4A MEAN DEVELOPMENTAL PERIOD TRIAL #3

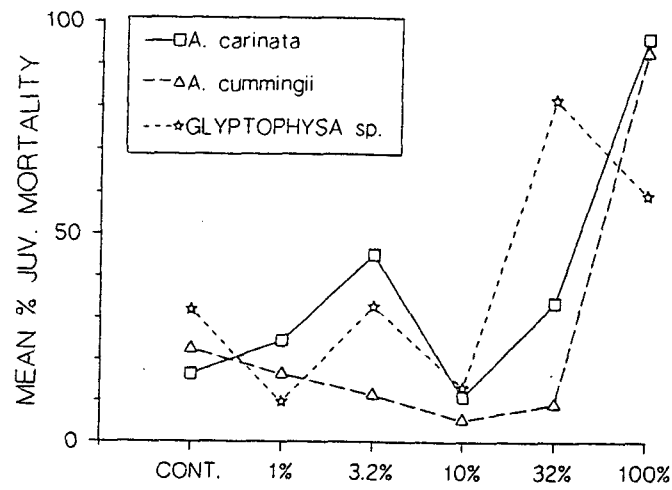


FIG 4B MEAN % JUVENILE MORTALITY TRIAL #3

<b>TRIAL #4 COMMENCED 17/3/91</b>
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Trial #4 was conducted to determine the RP2 concentration at which developmental retardation began. For this, a geometric series of five treatments between 100% and 32% inclusive was used. It was attempted to determine the life stage at which effects commenced. Data is tabulated in Appendix 4. From considering the raw data, it appeared that most of the retardation occurred during the hatchling stage, and that stages leading up to this were not affected in that way. Results of analysis of variance performed on pooled replicate data is presented in Table 6a.

**Table 6a. Results of analysis of variance. Trial #4**

Species	Endpoint	P>F	Conclusion
<b>Period spent pre-hatchling</b>			
<i>A. carinata</i>		0.6992	No significant difference
<i>A. cumingii</i>		0.0087	Significantly different
<b>Period spent as hatchling</b>			
<i>A. carinata</i>		0.033	Significantly different
<i>A. cumingii</i>		0.0008	Significantly different
<b>Period of development (Developmental retardation)</b>			
<i>A. carinata</i>		0.0509	No significant difference
<i>A. cumingii</i>		0.0001	Significantly different
<i>Glyptophysa</i> sp.		0.0067	Significantly different

Although *A. carinata* showed significant difference in analysis of variance, a LOEC was not established by Dunnett's testing. The error term from analysis of variance of period as hatchling (EMSS=2.3665), was large in comparison to the error term of the same parameter of *A. cumingii* (EMSS=0.3665). *A. cumingii* was retarded by the higher concentrations in pre-hatchling and hatchling stages (Dunnett's testing Table 6b.), and *Glyptophysa* sp. was retarded over the full developmental period, but the data was unfit for analysis. (This robust species had a low fertility rate, many eggs degenerated or were infected by a fungus). The means of replicate values are illustrated in Figures 5a, 5b and 5c. Figures 5d to f illustrate the retarding effect at the hatchling stage

Table 6b. Results of Dunnett's testing. Trial #4.

Species	Endpoint	NOEC	LOEC
<b>Period spent pre-hatchling</b>			
<i>A. cumingii</i>		56%	75%
<b>Period spent as hatchling</b>			
<i>A. carinata</i>		100%	undefined
<i>A. cumingii</i>		75%	100%
<b>Developmental period</b>			
<i>A. cumingii</i>		<32%	32%

From the *A. carinata* results developmental retardation was seen to occur at neither the pre-hatchling nor the hatchling embryonic stage, although a strong trend towards retardation during the late stages of development was noted. A large error term associated with the analysis of variance obscured this in the Dunnett's testing. For *A. cumingii*, it is possible to say that effects occurred at both pre-hatchling and hatchling stages. From the analysis it would appear that effects are more pronounced at the earlier developmental stages and that effects over the developmental period are cumulative.

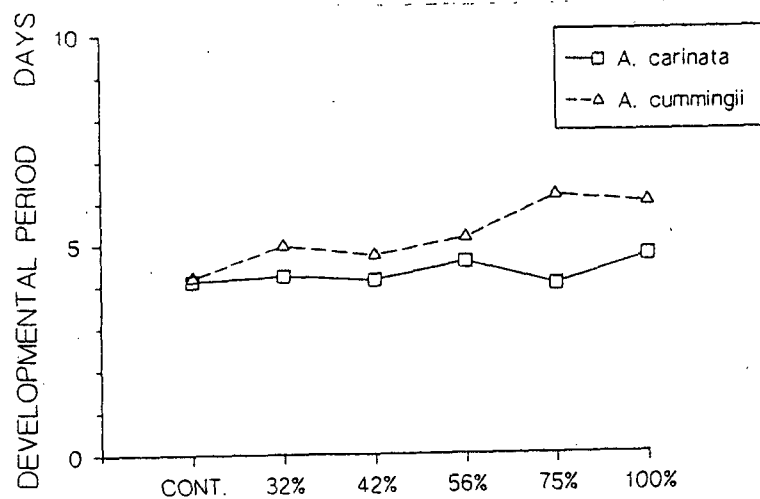


FIGURE 5A PERIOD SPENT PRE-HATCHLING TRIAL #4

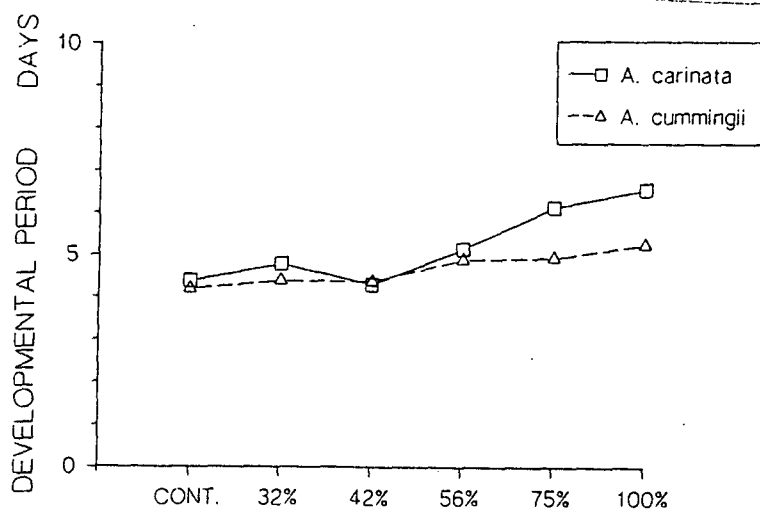


FIGURE 5B PERIOD SPENT AS HATCHLING TRIAL #4

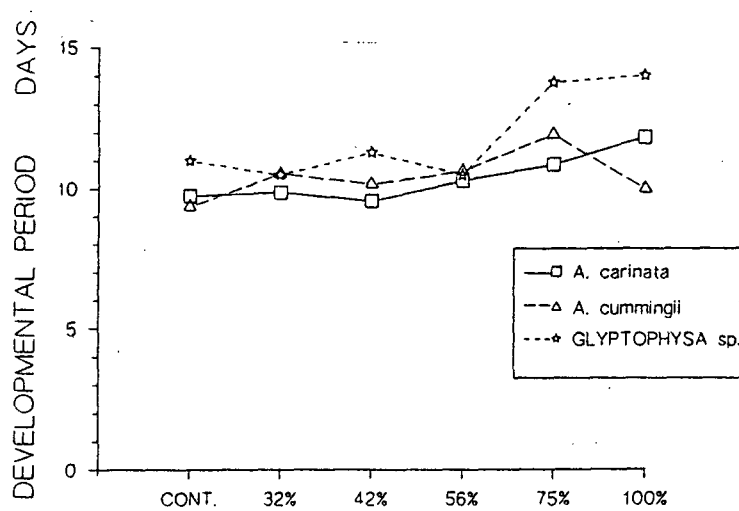


FIGURE 5C DEVELOPMENTAL PERIOD TRIAL #4



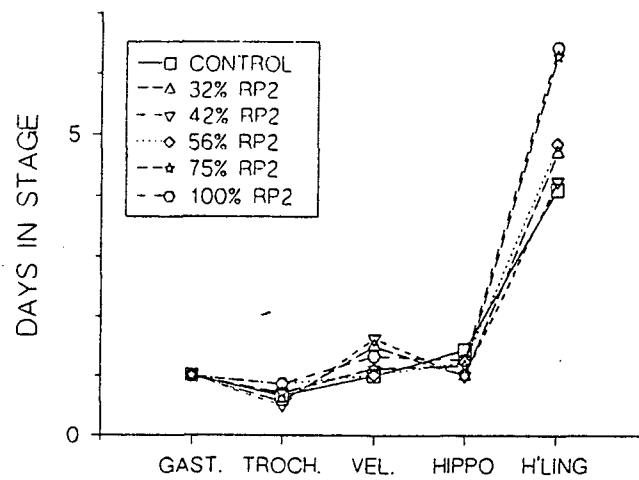


FIG. 5D DEVELOPMENTAL RETARDATION TRIAL #4  
A. carinata

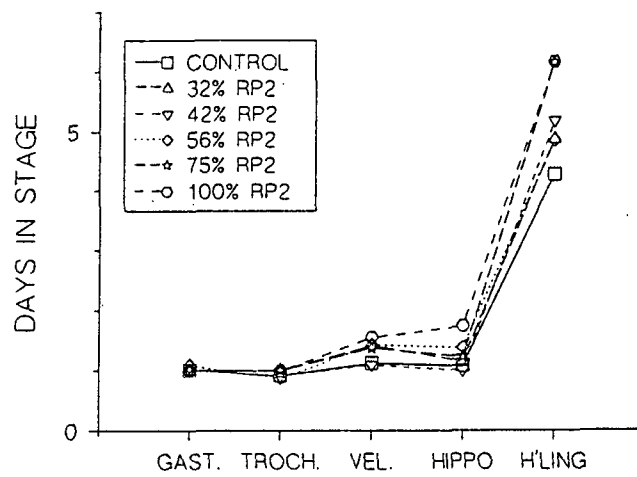


FIG. 5E DEVELOPMENTAL RETARDATION TRIAL #4  
A. cummingii

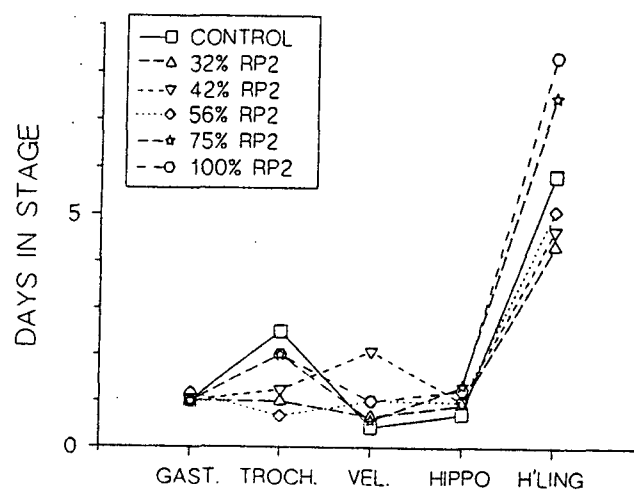


FIG. 5F DEVELOPMENTAL RETARDATION TRIAL #4  
GLYPTOPHYSA sp.

<b>Trial #5 Commenced 17/3/91</b>
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Trial #5 investigated the effects of RP2 water on the fecundity of *A. carinata*, *A. cumingii* and *Glyptophysa* sp. It was postulated that the metabolic load imposed by toxicity would decrease reproductive output. Tabulated data is presented in Appendix 6. Results of analysis of variance performed on pooled replicate data is presented in Table 7a. Figures 6a to 6c display average daily egg mass production means, and Figures 6d to 6f display average daily total egg production. Full statistical results appear in Appendix 11.4.a-d.

**Table 7a. Results of analysis of variance. Trial #5.**

Species	Endpoint	P>F	Conclusion
Average daily egg mass production			
<i>A. carinata</i>		0.001	Significantly different
<i>A. cumingii</i>		0.0001	Significantly different
Average daily egg mass production			
<i>A. carinata</i>		0.0001	Significantly different
<i>A. cumingii</i>		0.0001	Significantly different

Where significant differences were found Dunnett's testing was carried out to find difference from control means. The results appear in Table 7b.

**Table 7b. Results of Dunnett's testing. Trial #5.**

Species	Endpoint	NOEC	LOEC
Average daily egg mass production			
<i>A. carinata</i>		<1% RP2	1% RP2
<i>A. cumingii</i>		3.2% RP2	10% RP2
Average daily egg production			
<i>A. carinata</i>		<1% RP2	1% RP2
<i>A. cumingii</i>		3.2% RP2	10% RP2

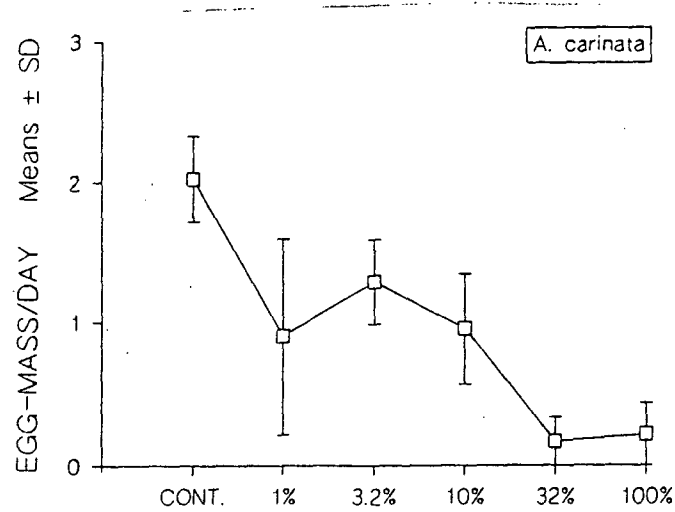


FIG. 6A AVERAGE EGG MASS PRODUCTION TRIAL #5

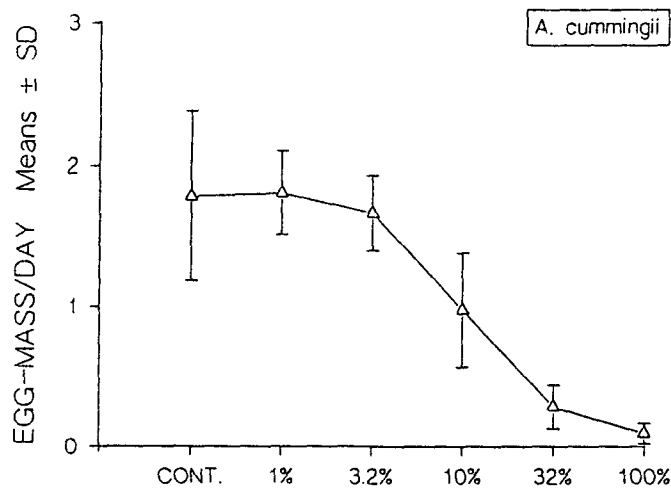


FIG. 6B AVERAGE EGG MASS PRODUCTION TRIAL #5

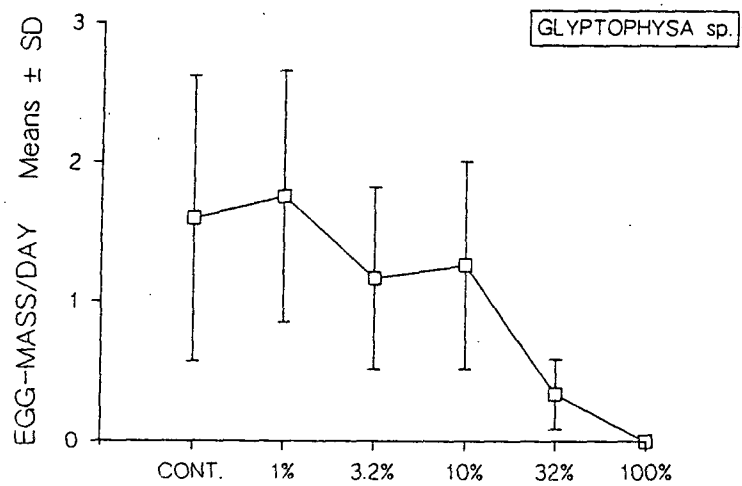


FIG. 6C AVERAGE EGG MASS PRODUCTION TRIAL #5

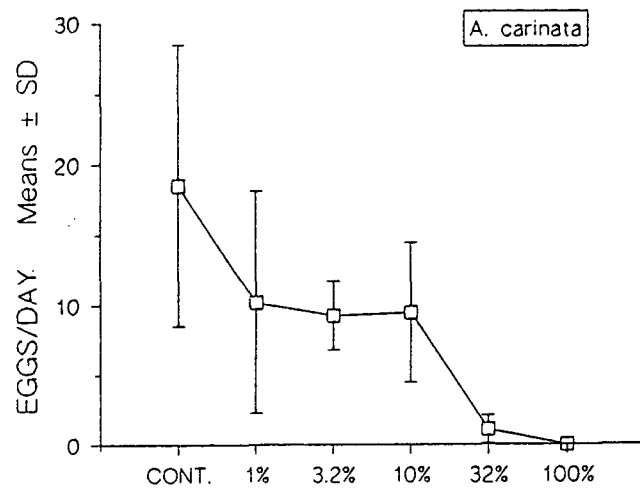


FIG. 6D AVERAGE EGG PRODUCTION TRIAL #5

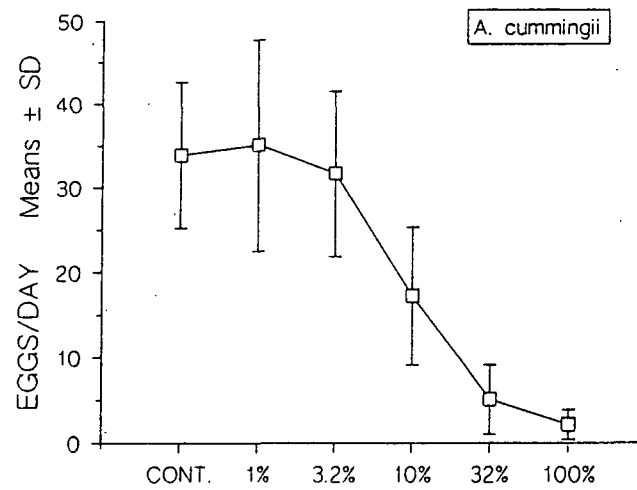


FIG. 6E AVERAGE EGG PRODUCTION TRIAL #5

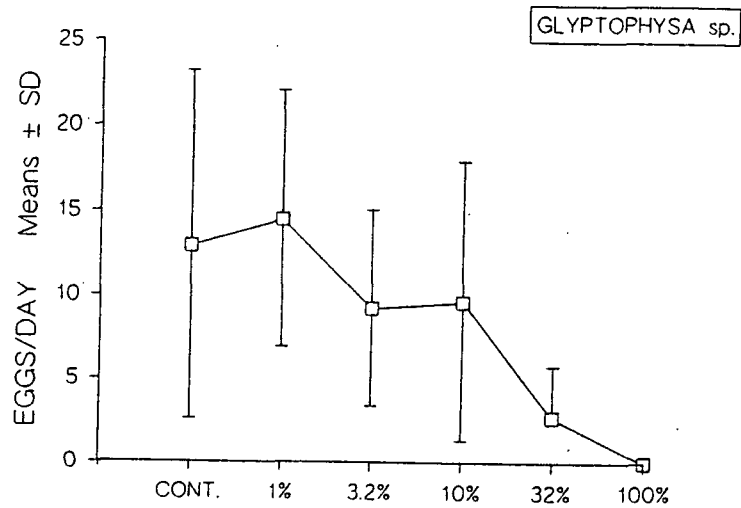


FIG. 6F AVERAGE EGG PRODUCTION TRIAL #5

The results of Trial #5 suggested that the fecundity parameters were sensitive measures of RP2 water effects, and that in terms of these parameters *A. carinata* was the most sensitive of the species examined.

# **TRIAL KNOWN PARENT**

This trial was conducted to investigate whether parentage contributed to the within treatment variation observed in Developmental retardation and Juvenile mortality. The results of analysis of variance performed on data pooled gained from the replicate samples is presented in Table 8a. Raw data appears in Appendix 9a.

Table 8a. Results of analysis of variance. Trial Known parentage.

Species	Endpoint	P>F	Conclusion
% Hatched/laid			
<i>A. carinata</i>		0.6605	No significant difference
<i>A. cumingii</i>		0.2137	No significant difference
% Mortality/hatched			
<i>A. carinata</i>		0.7870	No significant difference
<i>A. cumingii</i>		0.6290	No significant difference
% Mortality/laid			
<i>A. carinata</i>		0.9955	No significant difference
<i>A. cumingii</i>		0.3953	No significant difference

There was no significant difference found between various parameters measured. Although sample numbers were uneven and low (*A. carinata*:4,4,3,3. *A. cumingii*:4,4,3,3,2) the trial indicated that the observed variation in other trials was not due to genetic factor in the population sampled.

The lettuce discs were weighed daily before and after each daily exposure. It was seen to be of little use as an endpoint because of vegetative decay and differences in each disc weight due to internal structure (thick veins etc).

<b>Trial #6. Validation commenced 12/4/91</b>
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Trial #6 involved concurrent testing in the laboratory and at the creekside monitoring station. The differences in these two situations were flow rate, temperature and light. The full results from the creekside station testing were unavailable for analysis, and basic statistics and results of T-testing to determine LOECs and NOECs only were available. Tabulated data is in Appendix 7. Results of analysis of variance are in Table 9a. Full results of statistical analysis are in Appendix 11.6. Egg mass production, egg production, weight loss and control reared juvenile mortality are illustrated in Figures 7a-j.

**Table 9a. Results of analysis of variance. Trial #6 Laboratory results.**

Species	Endpoint	P>F	Conclusion
<b>Daily egg mass laying</b>			
<i>A. carinata</i>		0.0001	Significantly different
<i>A. cumingii</i>		0.0001	Significantly different
<i>Glyptophysa</i> sp.		0.0001	Significantly different
<b>Daily egg laying</b>			
<i>A. carinata</i>		0.0001	Significantly different
<i>A. cumingii</i>		0.0001	Significantly different
<i>Glyptophysa</i> sp.		0.0008	Significantly different
<b>Adult weight change - % loss</b>			
<i>A. carinata</i>		0.1631	No significant diff.
<i>A. cumingii</i>		0.0550	No significant diff.
<i>Glyptophysa</i> sp.		0.0010	Significantly different

pto

**Table 9a. contd Results of analysis of variance. Trial #6 Laboratory results.**

Species	Endpoint	P>F	Conclusion
<b>Adult weight change - % gain</b>			
<i>A. carinata</i>		0.3241	No significant diff.
<i>A. cumingii</i>		0.2405	No significant diff.
<b>Juvenile mortality - exposed egg masses</b>			
<i>A. carinata</i>		0.7027	No significant diff.
<i>A. cumingii</i>		0.0558	No significant diff.
<i>Glyptophysa</i> sp.		0.4196	No significant diff.
<b>Juvenile mortality - control reared egg masses</b>			
<i>A. cumingii</i>		0.3003	No significant diff.

The results of Dunnett's testing following significant differences are in Table 9b.

**Table 9b. Results of Dunnett's testing. Trial #6 Laboratory results.**

Species	Endpoint	LAB.	CREEK
<b>Average daily egg mass production</b>			
<i>A. carinata</i>		10% RP2	1.0% RP2
<i>A. cumingii</i>		3.2% RP2	10% RP2
<i>Glyptophysa</i> sp.		32% RP2	10% RP2
<b>Average daily egg production</b>			
<i>A. carinata</i>		1% RP2	No data
<i>A. cumingii</i>		10% RP2	No data
<i>Glyptophysa</i> sp.		32% RP2	No data
<b>Percentage weight loss</b>			
<i>Glyptophysa</i> sp.		Undefined	No data
<b>Juvenile mortality</b>			
<i>A. carinata</i>		Undefined	1.0% RP2

Parameters which produced significant differences from the control values were average daily egg and egg mass production. The NOEC established for *A. carinata* using egg mass production was higher in this trial than the previous. For *A. cumingii* the same parameter produced a LOEC of 3.2% RP2, as opposed to one of 10% RP2 in the previous trial. This cannot be readily explained as the error terms associated with *A. carinata* are similar for egg mass production in Trials 5 and 6 (Standard error=1.450 Vs 1.365 respectively). If the sampling period was too brief, one would expect a larger error term in the shorter period. The same variable for *A. cumingii* is lower in the shorter trial. For total egg production, the two trials yielded identical NOECs and LOECs for *A. carinata* and *A. cumingii*. In trial #6 *Glyptophysa* sp. was seen to be comparatively insensitive in terms of the fecundity parameters. The weight loss parameters presented some difficulty in analysis. There was generally no consistent trend in either weight loss or weight gain within replicates of any treatments. As well as inaccuracies associated with measurement, individual responses to osmotic and metabolic stress may make this parameter too variable for use in short term biological monitoring.

Juvenile mortality was insensitive and in the lab produced no significant differences between treatments for the three species tested. In this trial only egg mass produced by adults exposed to the toxicant were used, in many samples there were no juveniles produced. Juveniles reared in control water and subsequently exposed did not show significant differences under analysis of variance.

Laboratory and field results were generally similar. Fecundity parameters were higher in the field for *A. carinata* and *A. cumingii*, while *Glyptophysa* sp. showed similar responses in lab and field. In terms of response to RP2 water, similar decreases in fecundity were observed, although only *A. carinata* appeared as sensitive in the field (Figures 7k-p). Juvenile mortality did not appear to correspond well in the two situations, mortalities in the field were well above lab mortalities at the lower concentrations.

LOECs from the lab and creekside trials did not correspond exactly. However, lab results from the previous trial for



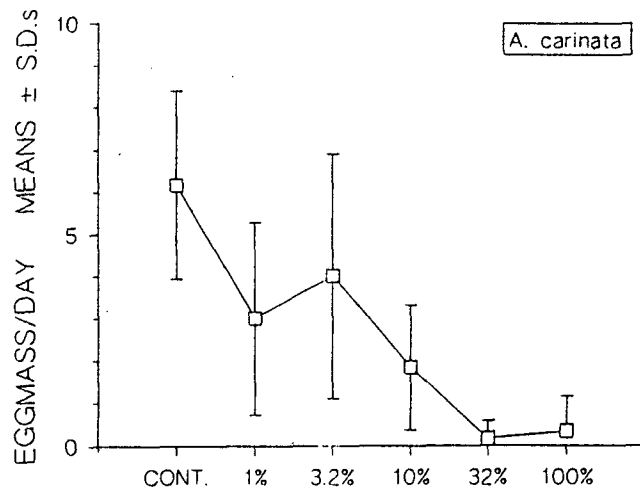


FIGURE 7A AVERAGE DAILY EGGMASS LAYING

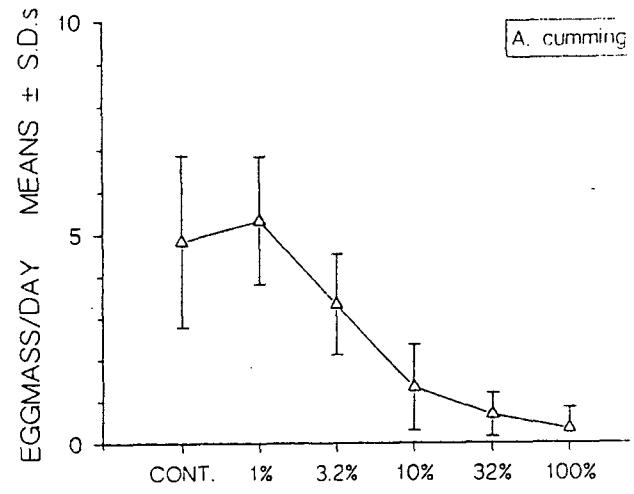


FIGURE 7B AVERAGE DAILY EGGMASS LAYING

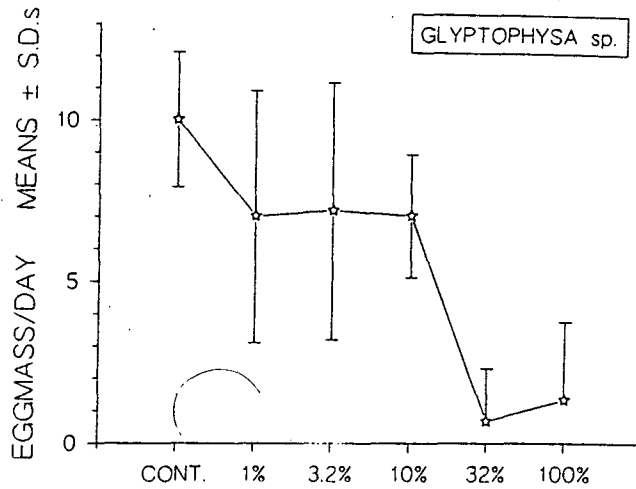


FIGURE 7C AVERAGE DAILY EGGMASS LAYING

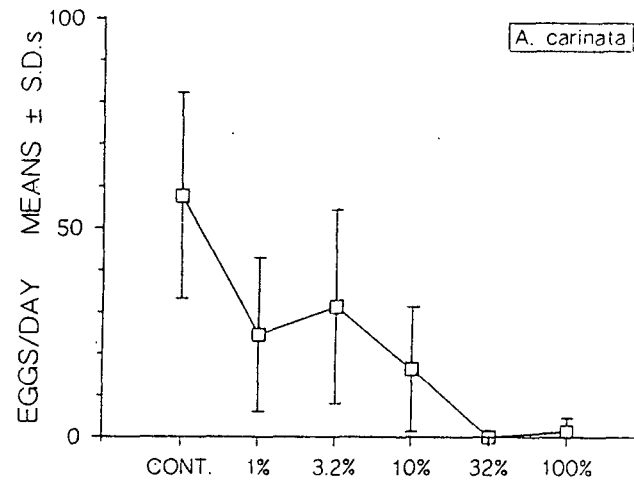


FIGURE 7D AVERAGE DAILY EGG LAYING

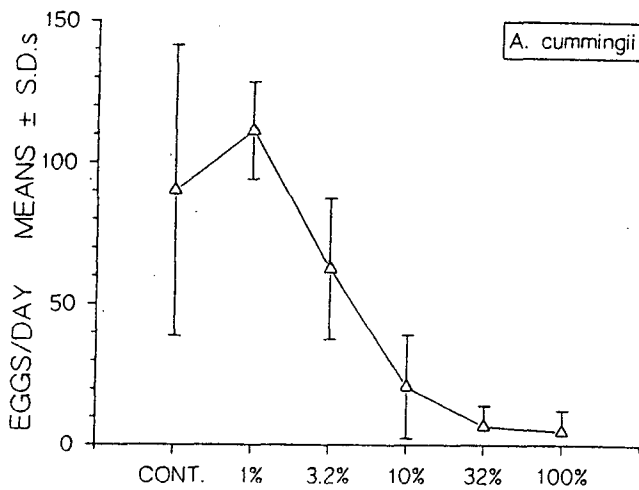


FIGURE 7E AVERAGE DAILY EGG LAYING

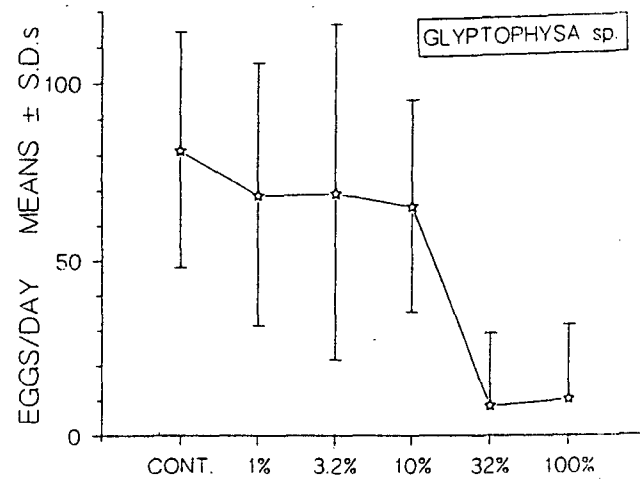


FIGURE 7F AVERAGE DAILY EGG LAYING

TRIAL #6

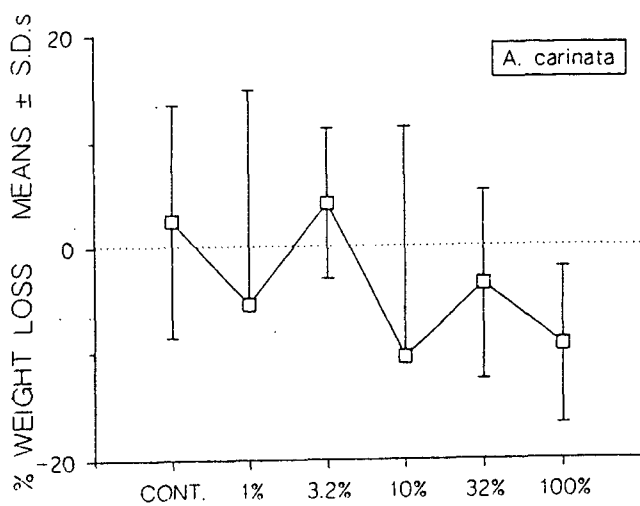


FIGURE 7G AVERAGE WEIGHT LOSS

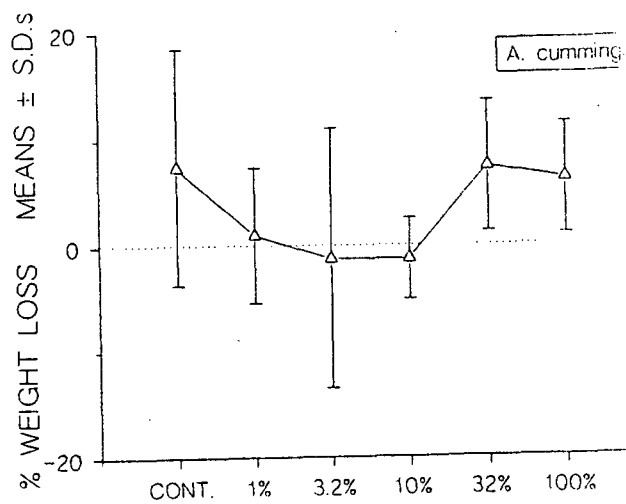


FIGURE 7H AVERAGE WEIGHT LOSS

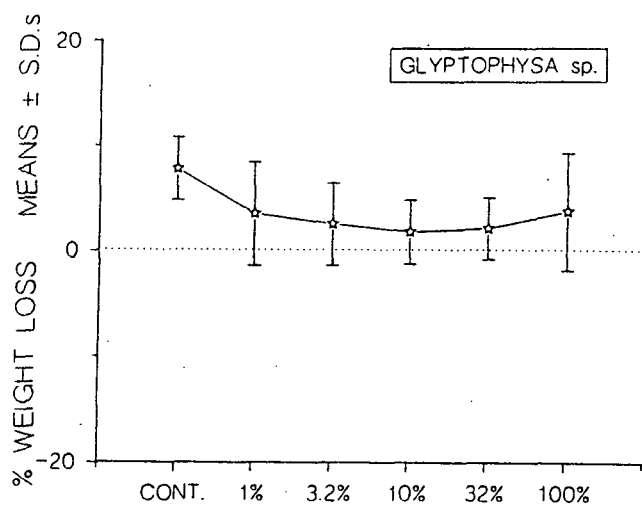


FIGURE 7I AVERAGE WEIGHT LOSS

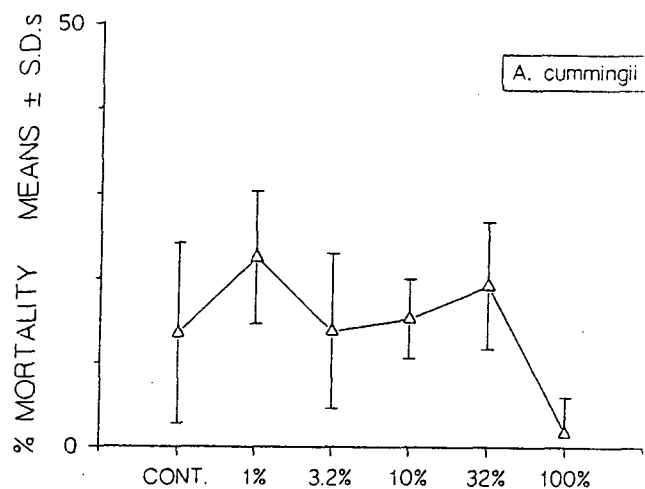


FIGURE 7J CONTROL REARED JUV. MORTALITY

TRIAL #6 LAB/FIELD

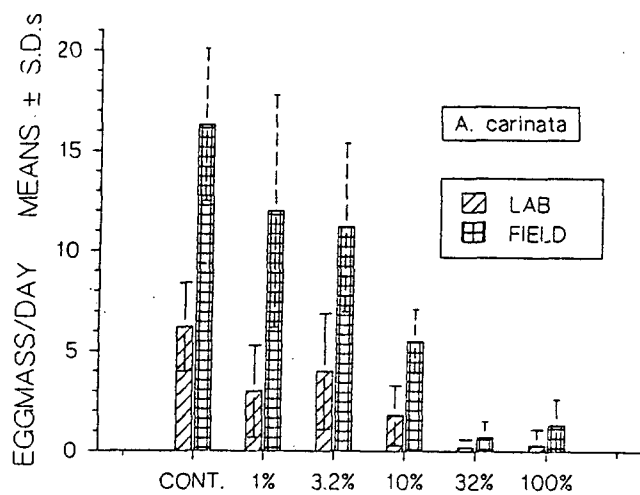


FIGURE 7K AVERAGE DAILY EGGMASS LAYING

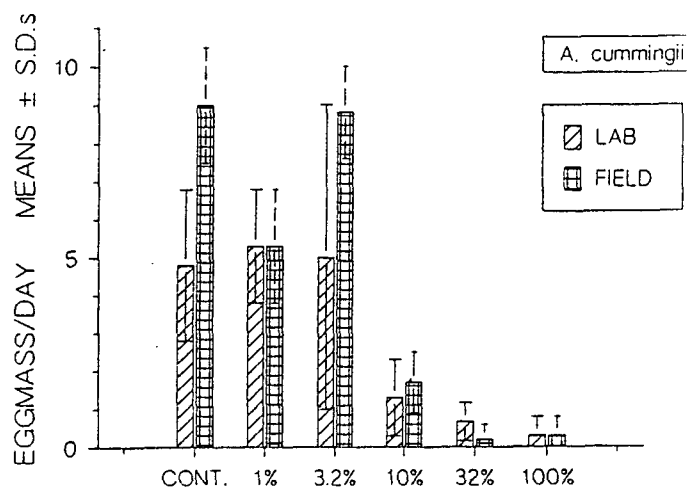


FIGURE 7L AVERAGE DAILY EGGMASS LAYING

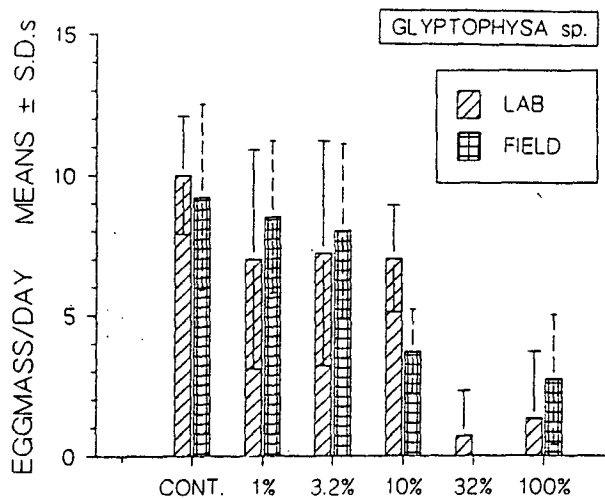


FIGURE 7M AVERAGE DAILY EGGMASS LAYING TRIAL

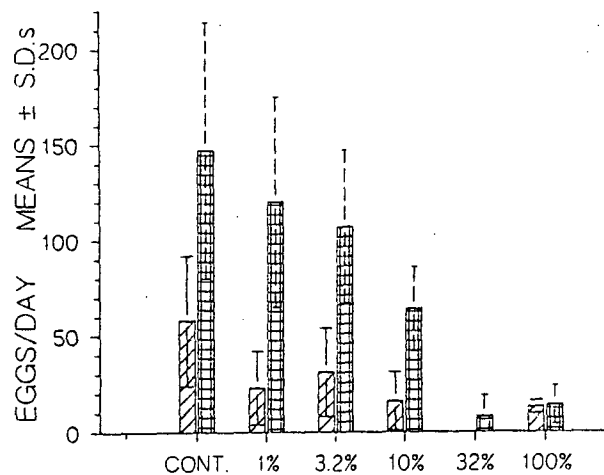


FIGURE 7N AVERAGE DAILY EGG LAYING TRIAL

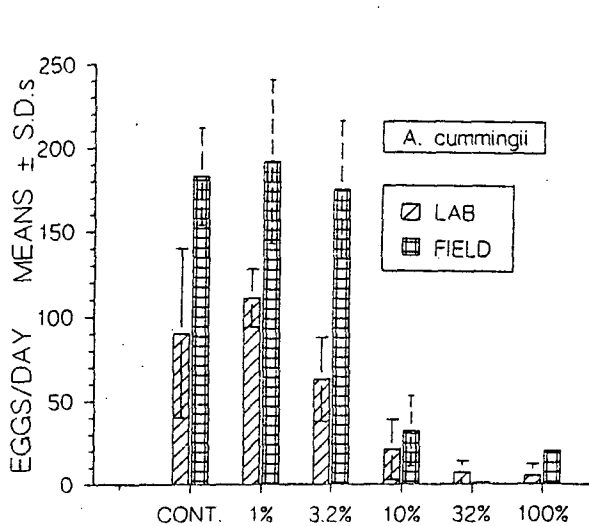


FIGURE 7O AVERAGE DAILY EGG LAYING TRIAL

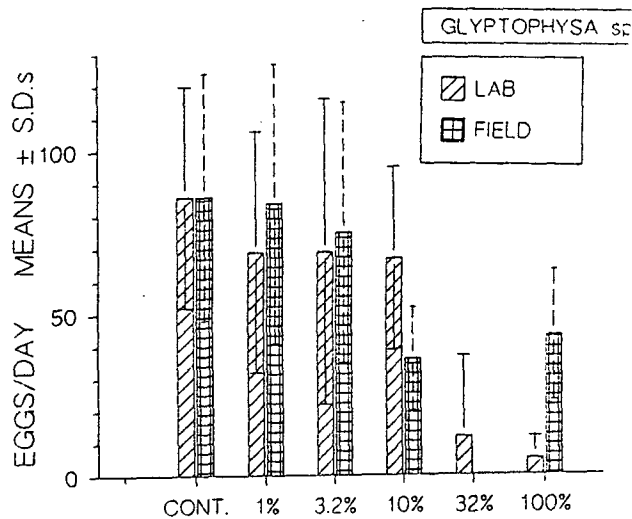


FIGURE 7P AVERAGE DAILY EGG LAYING TRIAL

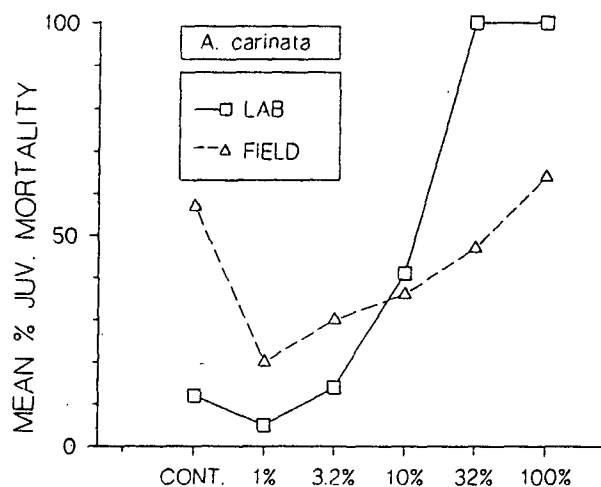


FIGURE 7Q MEAN JUV. MORTALITY TRIAL FROM EXPOSED EGG MASSES

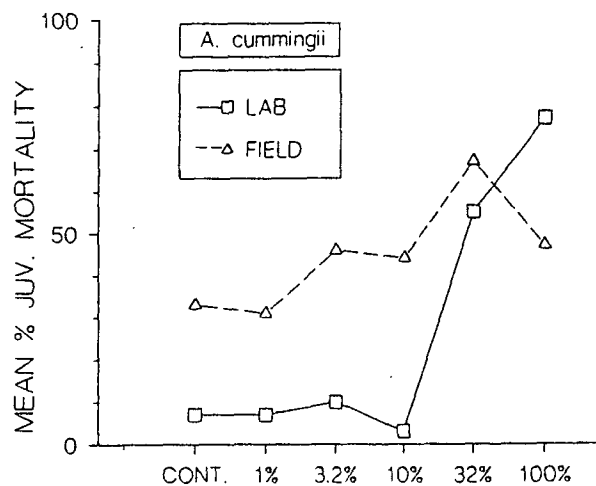


FIGURE 7R MEAN JUV. MORTALITY TRIAL FROM EXPOSED EGG MASSES

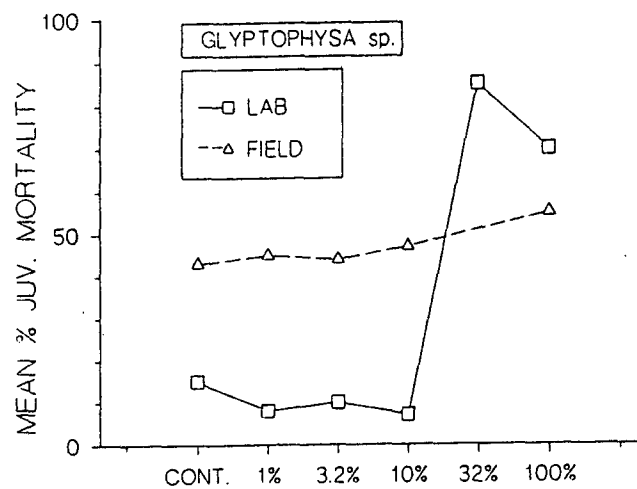


FIGURE 7S MEAN JUV. MORTALITY TRIAL #6 LAB/FIELD FROM EXPOSED EGG MASSES

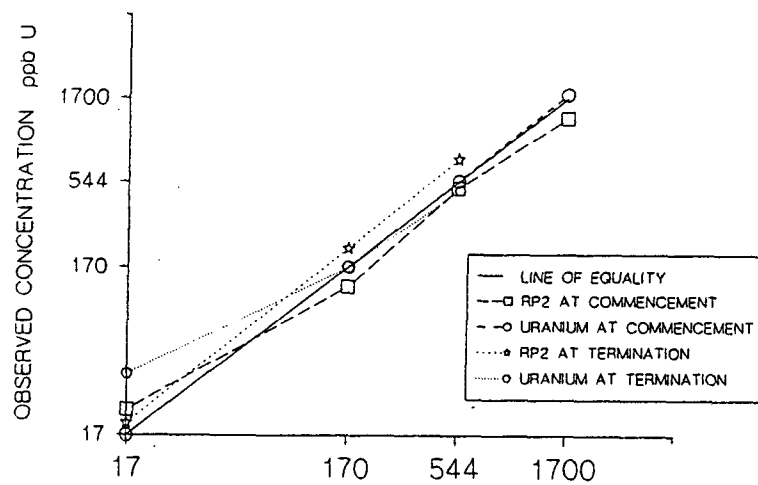


FIGURE 8A NOMINAL URANIUM CONCENTRATIONS ppb U TRIAL #7

<b>Trial #7 Commenced 2/5/91 RP2 - Uranium comparison</b>
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This trial was conducted to compare the effects of uranium solutions of similar concentration to the total uranium content of the RP2 dilutions. An initial 100% RP2 was changed to 32% RP2 after mortality of all adult *A. carinata* after 24 hours exposure to this concentration.

Tabulated data is in Appendix 8. Nominal and observed concentrations of RP2 and uranium treatments are illustrated in Figure 10a. Full Scintrex and GFAAS determinations of uranium and manganese are in Appendix 10e. Results of T-tests between the corresponding RP2 and uranium treatments are in Table 9a.

**Table 10a Results of T-testing. Trial #7.**

Species	Treatment*	P>F (Ho:Var's =)	P>T (Ho:Means =)	Result
<b>Daily egg mass production</b>				
<i>A. carinata</i>	1%	0.499	0.0524	Do not reject Ho.
	10%	1.000	0.0006	Reject Ho.
	32%	0.5345	0.5364	Do not reject Ho.
<i>A. cumingii</i>	1%	0.7599	0.1048	Do not reject Ho.
	10%	0.4650	0.0493	Reject Ho.
	32%	0.5276	0.2560	Do not reject Ho.
<b>Daily egg production</b>				
<i>A. carinata</i>	1%	0.9013	0.5434	Do not reject Ho.
	10%	0.7344	0.2087	Do not reject Ho.
	32%	0.4648	0.3453	Do not reject Ho.

\* The stated treatments (1, 10 and 32%) refer to testing between RP2 and U treatments of nominally identical U concentrations.

pto

Table 10a contd. Results of T-testing. Trial #7.

Species	Treatment *	P>F (Ho:Var's =)	P>T (Ho:Means =)	Result
Daily egg production				
<i>A. cumingii</i>	1%	0.7760	0.3642	Do not reject Ho.
	10%	0.2096	0.0614	Do not reject Ho.
	32%	0.7040	0.5477	Do not reject Ho.

For the endpoint average daily egg masses, both species at the 10% treatment showed significant differences. In both cases the RP2 mean was lower than the U mean. The 10% RP2 uranium concentrations as determined by the Scintrex method varied from commencement to termination (130 and 170 ppb U respectively). The reasons for this were not immediately clear. Discounting contamination, a possible explanation might be coprecipitation of the uranium with manganese, and subsequent concentration as that form in the exposure tanks. The pH levels were seen to vary markedly within treatments. Although a standardised procedure was adopted to measure pH, it is unlikely that were accurate. Analysis of variance showed significant difference between replicates and treatments ( $P>F=0.0002$ ,  $F=7.88$ ). SNK testing grouped together the two uranium treatments and the two RP2 treatments.

<b>TRIAL ALGAE/SIZE</b>
-------------------------

This was conducted for similar reasons as the known parentage trial. The trial sought to establish whether the presence or absence of algae in the vials, or the size of the vials, had any influence on juvenile mortality. Raw data appears in Appendix 9b. Results of analysis of variance appear in Table 11a.

**Table 11a. Results of analysis of variance. Trial Algae/size.**

Species	Endpoint	P>F	Conclusion
<b>Juvenile mortality</b>			
<i>A. carinata</i>			
	Size	0.0008	Significantly different
	+/-Algae	0.8886	No significant difference
	Interaction	0.5477	No significant difference
<i>A. cumingii</i>			
	Size	0.0051	Significantly different
	+/-Algae	0.7327	No significant difference
	Interaction	1.0000	No significant difference

SNK testing produced two groups for each species. The results appear in Table 11b.

**Table 11b. Results of comparisons from SNK testing. Trial Algae/size**

Species	Result	Parameter	Mean	SD
<b>Juvenile mortality</b>				
<i>A. carinata</i>		<sup>3</sup> Large +Algae	66.8%	2.9%
		<sup>3</sup> Large -Algae		
		<sup>3</sup> Small +Algae	30.9%	1.9%
		<sup>3</sup> Small -Algae		
<i>A. cumingii</i>		<sup>3</sup> Large +Algae	77.5%	5.6%
		<sup>3</sup> Large -Algae		
		<sup>3</sup> Small +Algae	25.7%	5.9%
		<sup>3</sup> Small -Algae		

These results indicated that the size of the container did influence juvenile mortality, but could not apportion the observed variation in juvenile mortality of earlier trials to any one factor. Algae was seen to be an unimportant factor in this analysis.

TABLE 12 CRITERIA AND RESPONSE

ENDPOINT	SPECIES						
	<i>A. carinata</i>	<i>G. l. A. cummingtoni</i>	<i>G. abbas</i>	<i>H. pyrophysa</i>	<i>G. yraulsa</i>	<i>e. l. corbis</i>	<i>L. ymnasa</i>
	sp.	sp.	sp.	sp.	sp.	sp.	sp.
<b>Natural</b>							
Egg mass prod.	++	+	-	+	++	-	-
Egg prod.	++	+	-	+	++	-	+
Handling	+	+	-	+	+	-	-
Hatchability		+	+	-	-	--	-
+							
Rearing	+	++	--	+	-	-	+
<b>Trials</b>							
Adult mortality	0	-	nt	-	nt	nt	nt
Dev'l retard'n	0	+	nt	0	+	nt	+
Embryonic mort.	-	-	nt	-	-	nt	-
Egg mass prod.	++	++	nt	+	nt	nt	nt
Egg prod.	++	+	nt	+	nt	nt	nt
Feeding	-	-	nt	-	nt	nt	nt
Juvenile mort.	0	+	nt	-	nt	nt	nt
Weight change	-	-	nt	+	nt	nt	nt
<b>Key</b>							
++	Low variation or sensitivity.		+	Moderate variation or sensitivity.			
0	Indifferent.		nt	Not tested.			
--	High variation or sensitivity.		-	Low variation or sensitivity.			



Table 12 indicates that the fecundity endpoints using *A. carinata* and *A. cumingii* gave the best results in terms of sensitivity and variation. Although *Gyraulus* sp. also showed potential with the fecundity endpoints, handling and rearing this species presented some difficulty. It is possible that these could be overcome. As previously mentioned *Gabbia* sp. and *Helicorbis* sp. were eliminated after the preliminary observations because of high variability in the fecundity parameters, and difficulties in handling and rearing. *Glyptophysa* sp. displayed low fertility and an insensitivity to the toxicant. *Lymnaea* sp. was difficult to handle and laid large egg masses unpredictably. The juveniles appeared to shelter in the egg mass after rupture.

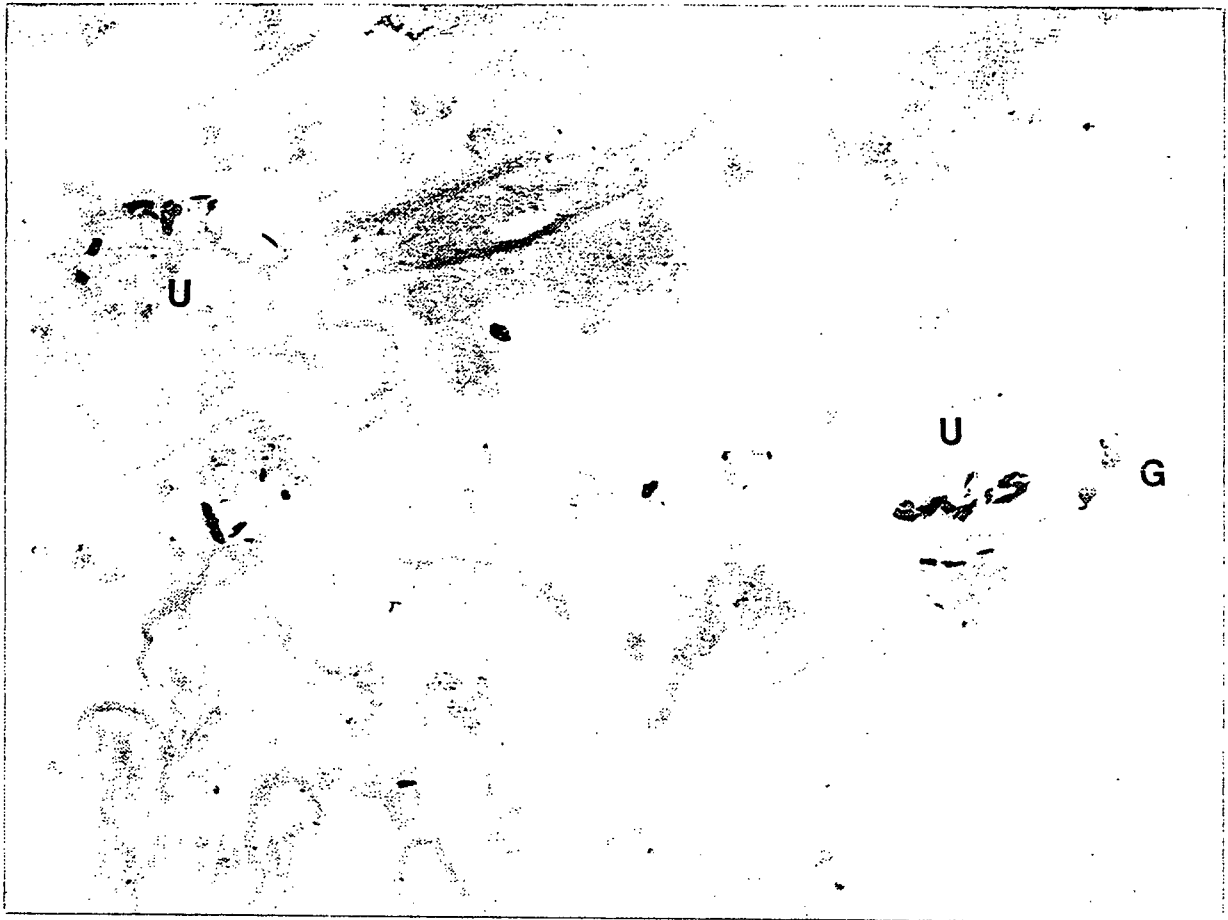
Although some species were not assessed using several endpoints, the success of other species justified this exclusion.



## SEQUESTRATION OF URANIUM

EDAX analysis of unstained sections of *A. carinata* from Trial 5 (exposed to 32% RP2 for seven days) identified uranium in the ovotestis. Other tissues examined which did not show uranium accumulation were the digestive gland (hepatopancreas), mantle, kidney, albumin gland, foot and hermaphroditic duct. In the ovotestis, the uranium was deposited as spindle shaped crystals in association with other electron dense amorphous material. Plates 4a to l show photomicrographs of sections from the ovotestis, mantle and kidney. Sections of the digestive gland were unsuitable for proper analysis because overfixation had made the tissue friable. Figure 1b, 1c and 1d show uranium crystals located in multivesicular bodies of the ciliated epithelium. Plate 4F shows typical electron dense granules in the ovotestis. Plate 4g shows mitosis with no apparent involvement of uranium. Plate 4h shows a multivesicular body at the cell membrane, apparently involved in endocytosis or exocytosis. Plates 4i and 4j show stained and unstained sections of the mantle, the typical electron dense granules are visible in the unstained section. Plate 4k shows a stained section of the kidney. Again normal electron dense granules are present. Plate 4l shows what is probably artifactual uranium crystals associated with the cell membrane. No such deposits were observed in the unstained sections.

The results of the ultrastructural study are somewhat inconclusive. The normal pathway for the deposition of metals in molluscs is via the digestive gland (gastropods) or the kidney (bivalves) (Simkiss and Mason, 1983). It must be assumed that the uranium is taken up, either in solution or associated with a metallothein-like molecule, and deposited in the digestive gland, with which the ovotestis makes close contact. Subsequent movement of the uranium into the digestive gland must then follow.



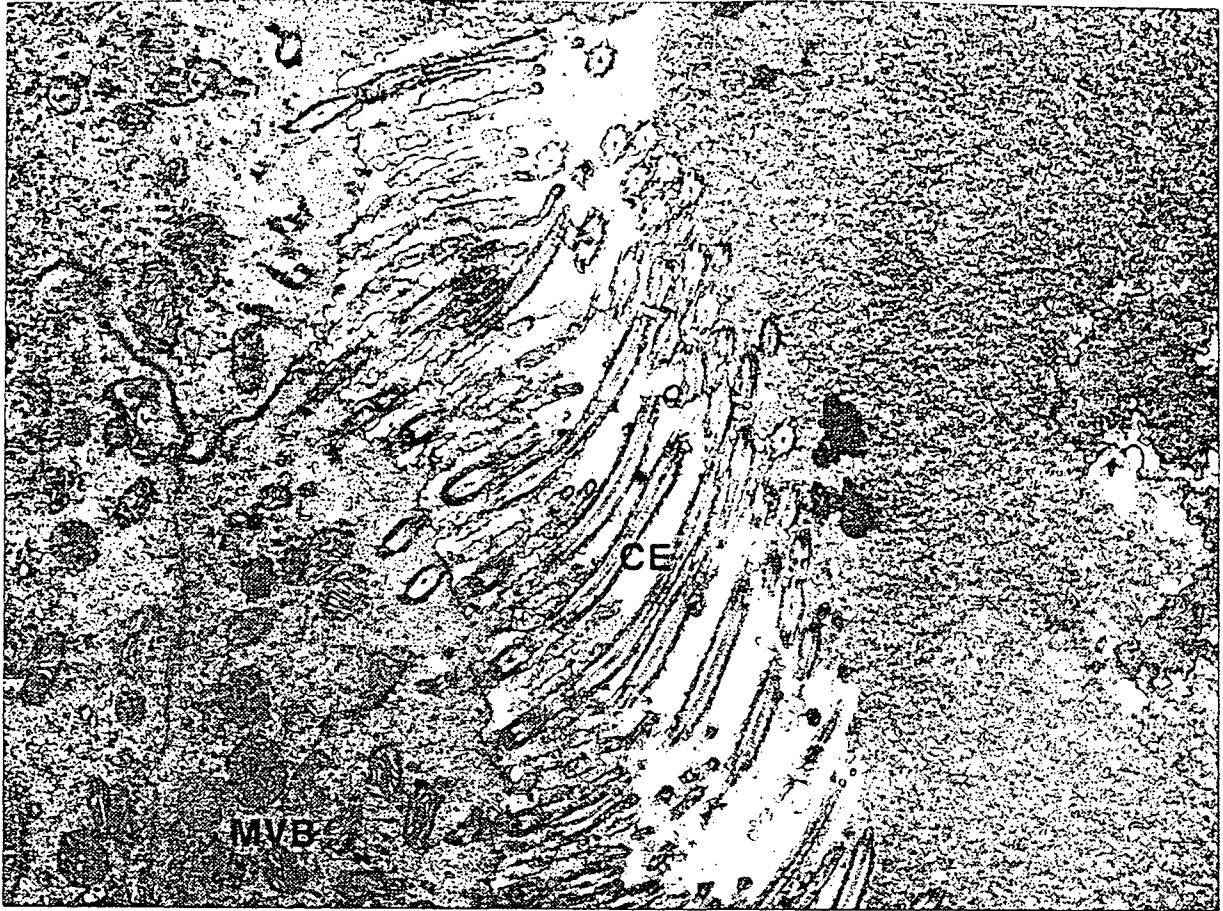
**Plate 4a** Unstained ovotestes : uranium and granules

**X 35,000**



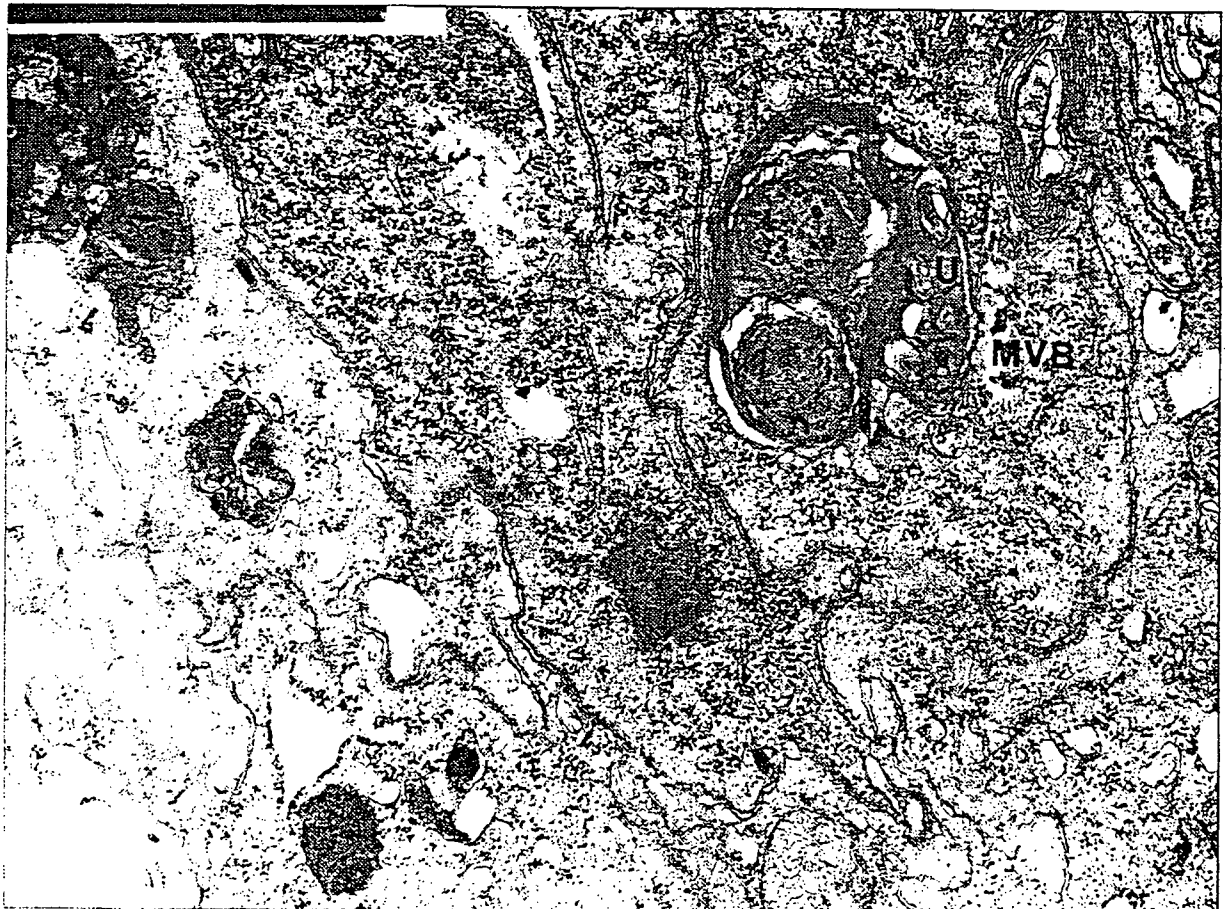
**Plate 4b** Stained ovotestes : uranium in multivesicular bodies

**X 35,000**



**Plate 4c** Uranium containing MVB in ovotestes eplthelia

**X 15,000**



**Plate 4d** Uranium crystals in a multivesicular body

**X 20,000**



Plate 4e

Enlargement of Plate 4a

X 90,000



Plate 4f

Typical granules in the ovotestes

X 15,000

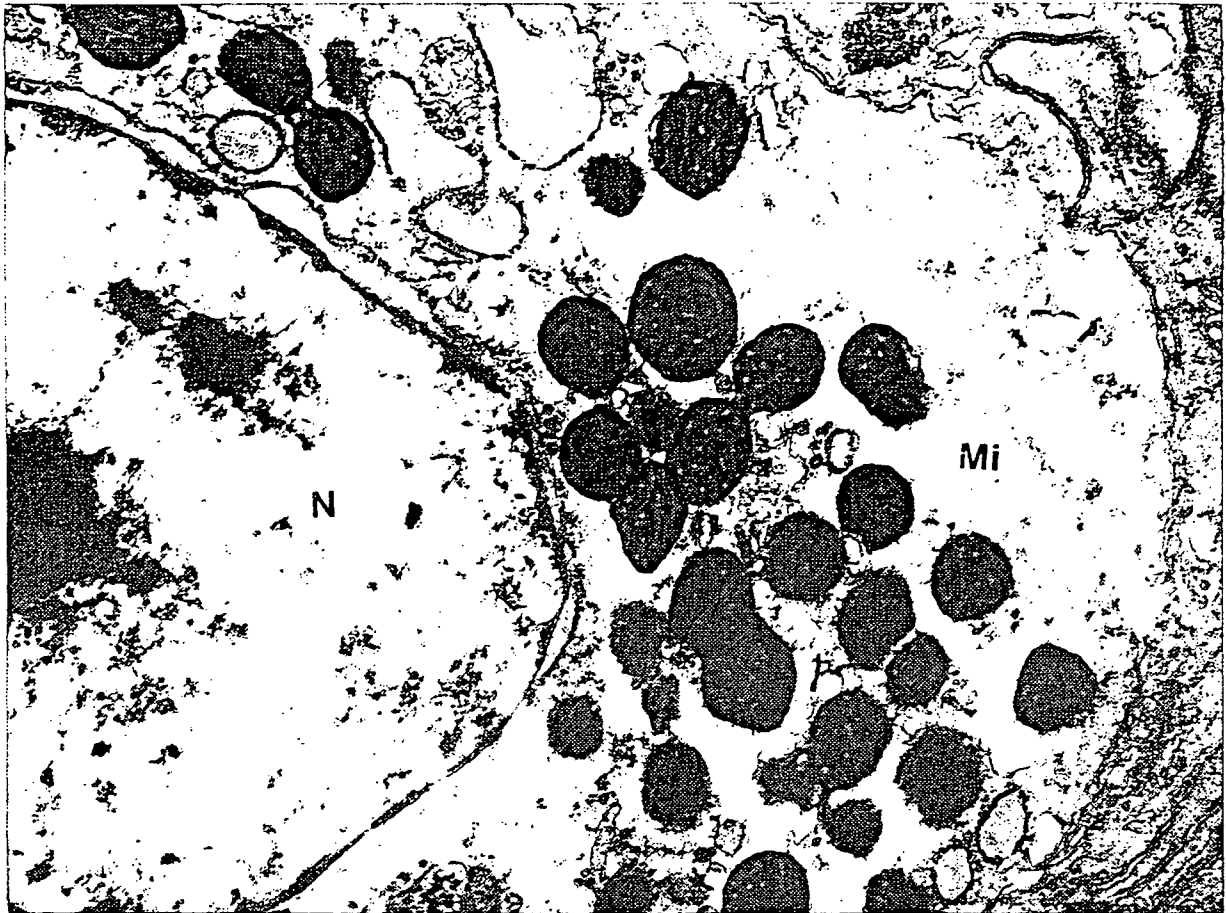


Plate 4g Cell undergoing meiosis : ovotestes

X 20,000

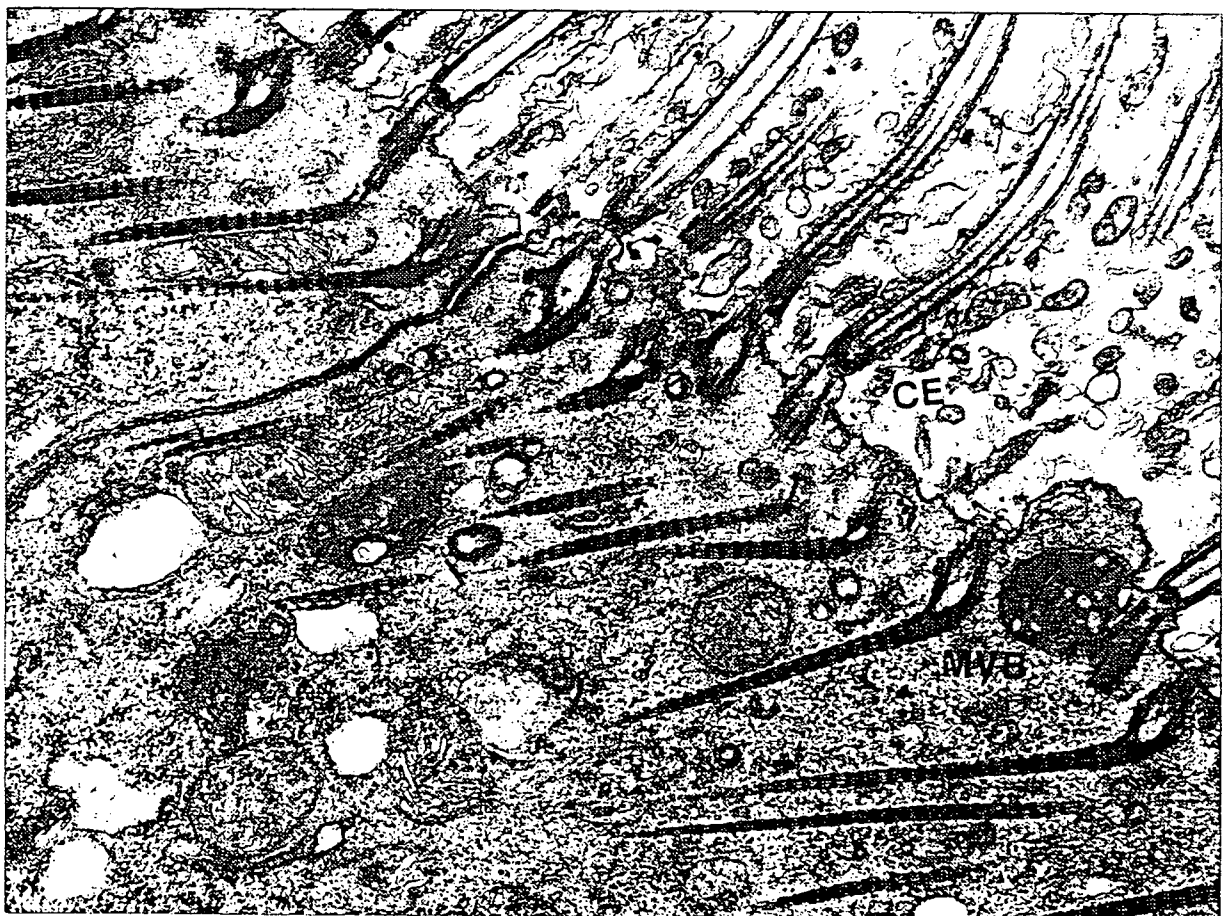


Plate 4h Multivesicular body at cell membrane : ovotestes

X 20,000



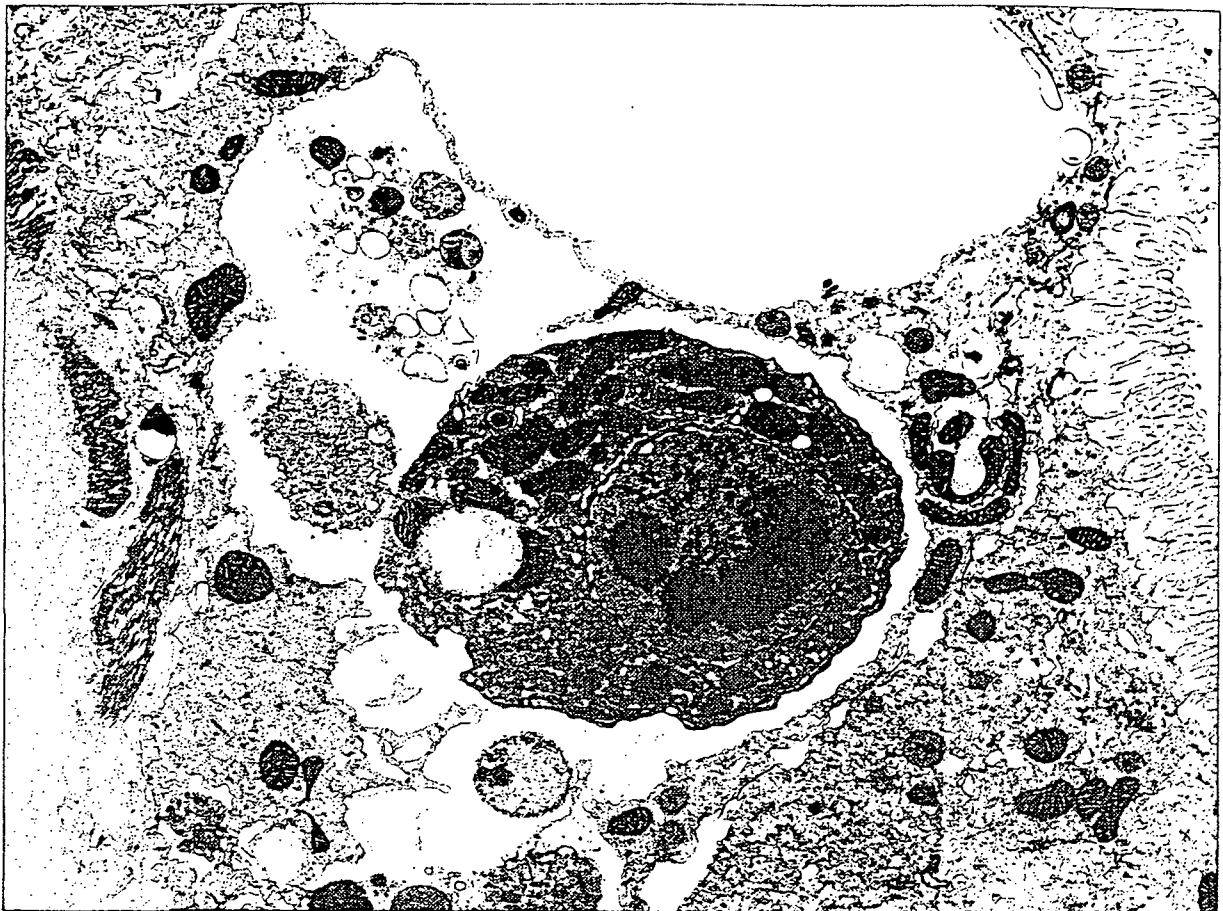


Plate 4i

Mantle

X 10,000

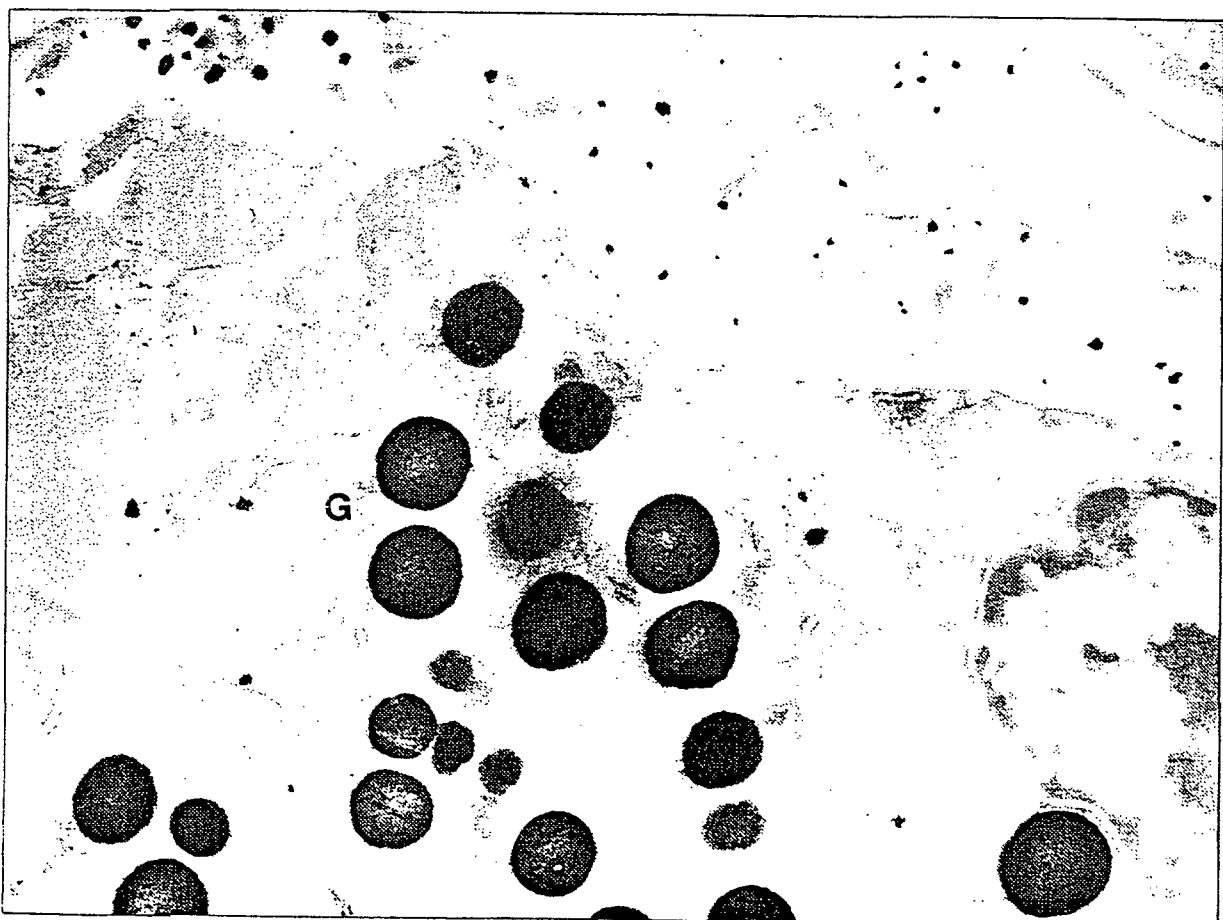


Plate 4j

Mantle

X 20,000



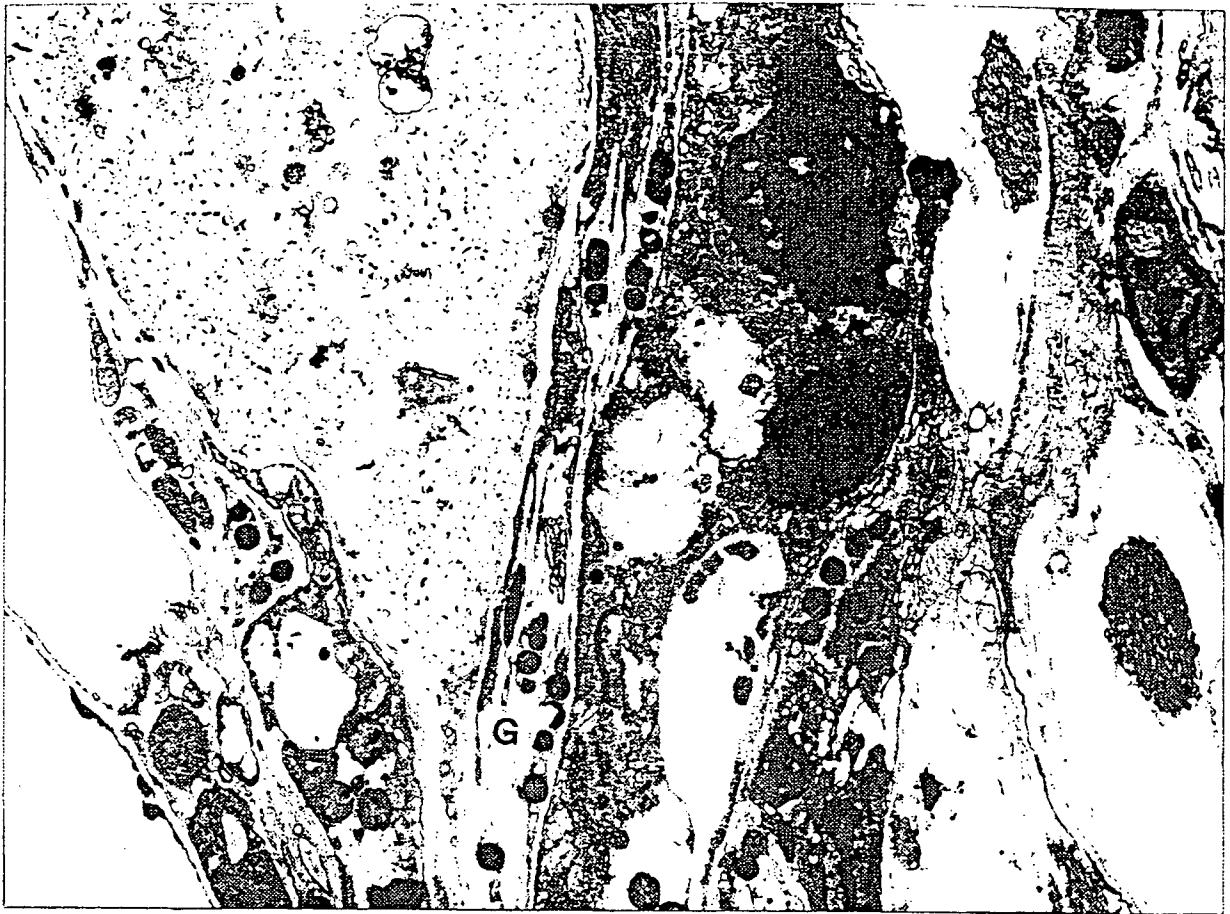


Plate 4k

Kidney

X 8,000

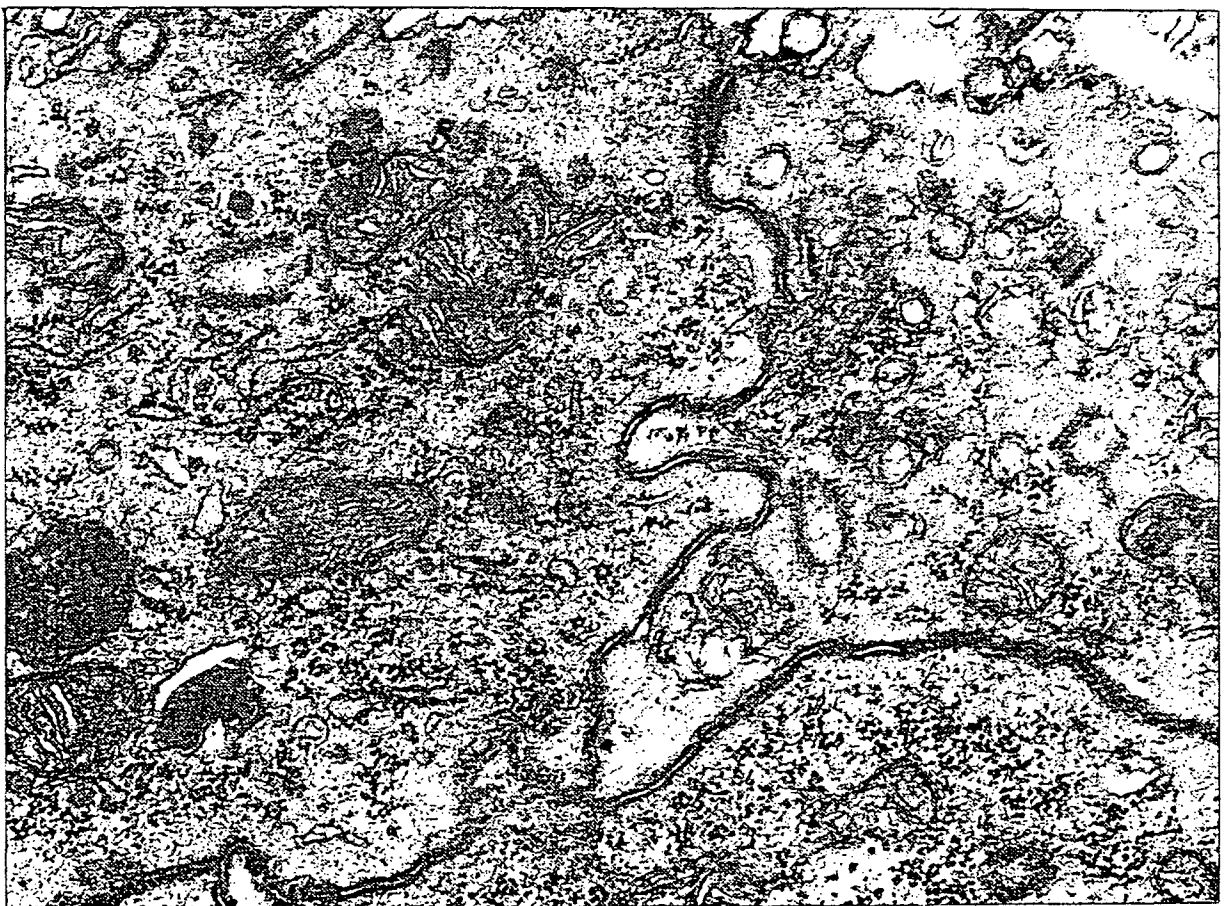


Plate 4l

Uranium in the intercellular space (artifact)

X 20,000

## DISCUSSION

### ENDPOINTS

#### Adult mortality

The mortality of all *A. carinata* adults after 24hrs exposure at 100% RP2 in Trial #7 and of an *A. carinata* individual at 100% RP2 in Trial #6 were the only instances of adult mortality throughout the trials period. Neither of these occurrences could be explained by dissolved oxygen, pH or conductivity levels, and must be due to RP2 toxicity. Generally, adult mortality was insensitive and inappropriate for short term biological monitoring of the RP2 waters using the species tested. All species tested became moribund at 100% RP2, implying a possible use for adults in longer trials.

#### Embryonic mortality

Of five species tested with this endpoint, none showed significant differences between treatments upon analysis of variance, although it appeared that the rate of embryonic breakdown increased with RP2 concentration. The reason for this may be that breakdown usually occurred in one of three ways:

- 1) acute breakdown and death in the transition between stages,
- 2) acute mortality during the later lengthier stages,
- 3) chronic deformation during the veliger and hippo phase (ghosting and hydro-cephaly) protracting a number of days until death.

In cases 1 and 2, the acuteness made for easy quantification and provided the bulk of embryonic mortality data. In case 3, quantification was more difficult and beyond the time resources available. Rupture of the egg mass and the hatching of healthy neonates usually occurred before the death of the deformed embryos. Only results from healthy individuals were considered as data for juvenile mortality. As no observations could be made following egg mass rupture (since the period of juvenile exposure required minimum disturbance) the fate of these deformed individuals was not recorded. Mortality due to deformation would have contributed to embryonic mortality, but might also have been a useful endpoint in its own right. Ravera (1991) states that a high proportion of embryonic deformation is common in cases of heavy metal exposure.

Generally, embryonic mortality as an endpoint was insensitive because it tended to occur regardless of concentration, and may be a response to factors outside RP2 exposure.

### Juvenile mortality

Trials 1, 3, 6 and 7 tested *A. carinata*, *A. cumingii* and *Glyptophysa* sp. in terms of juvenile mortality. Trial 1 involved RP4 water of low toxicity. Trial 3 established LOECs of 32% for *A. carinata* and *A. cumingii*. In Trials 6 and 7 no significant differences were found upon analysis of variance. The literature emphasises the susceptibility of molluscan juveniles to various metals. In these trials juvenile mortality was seen to be variable in response to RP2 concentration. At 100% RP2, Trial 3 mortalities were close to 100% for *A. carinata* and *A. cumingii*, but in Trial 6, a mean mortality of only 5% was recorded for *A. cumingii* at 100% RP2. A possible explanation for this variability might be the pH dependent speciation and toxicity of the uranate ion  $\text{UO}_2^{2+}$ . Magela Ck water made up to 900 ppb uranium and of pH range 6.0 to 7.0, was toxic to *Hydra viridissima*, while the same solution carbonate-buffered to pH 8.5 was not toxic (R. Hyne pers comm. 1991). The dominant complex at pH 8.5 would be the tricarbonatate  $[\text{UO}_2(\text{CO}_3)_3]^{4-}$ , which is membrane impermeable. At lower pHs, the uranate  $\text{UO}_2^{2+}$  or uranyl carbonate  $\text{UO}_2\text{CO}_3$  would predominate. These species would be more membrane permeable and therefore more toxic than the anionic forms (R. Hyne pers comm. 1991).

However this may not suffice to explain the difference in juvenile mortality between Trials 3 and 6 and between the RP2 and U treatments of Trial 7, since the measured pHs are not disparate to the extent described above, unless the juveniles are particularly sensitive to pH changes.

Other possible sources of extraneous juvenile mortality were examined. Known parentage and the presence or absence of algae were seen as unimportant, while an increase in the size of the exposure vial was seen to increase juvenile mortality. It is possible that a "rim-effect" operated, since the circumference of the larger vials was twice that of the smaller. At the termination of several trials, dead juveniles had been observed between the gauze and outside wall of the exposure vial. In such a position, it was possible that they had died by mechanical damage. This complication had been considered early in the trials, but had not been satisfactorily resolved by the end. The problem of large variation in juvenile mortality within treatments, and high control mortality, will continue to receive attention at OSS. In terms of the project, the question of juvenile sensitivity remains to a certain extent unanswered. Despite these uncertainties, low mortalities observed at high RP2 concentrations counted against the use of juvenile mortality as a suitable endpoint.

### **Weight change**

Analysis of variance revealed a significant difference between treatments for *Glyptophysa* sp. but a LOEC was not established by Dunnett's testing. The responses of *A. carinata* and *A. cumingii* in terms of this endpoint were irregular across the treatments and did not show significant differences upon analysis of variance. Osmoregulatory effects arising from the differing conductivities of the treatments might have interacted with intrinsic RP2 toxicity. Because percentile data was considered most the appropriate form for analysis and since some values were negative, arcsine transformation of the whole data set was not possible. This endpoint might be useful in chronic trials which incorporate a longer exposure period, but in these relatively brief trials, the degree of weight change was seen to be insensitive.

### **Developmental retardation**

Trials 2, 3 and 4 investigated developmental retardation as a response to RP2 toxicity. Although this effect was apparent at 100% RP2 during the trials, Dunnett's testing did not always reveal this. The conservative nature of the statistics employed were probably producing type 2 errors, where large error terms associated with analysis of variance made the *q* statistic of Dunnett's test insignificant. This occurred for *A. cumingii* and *Glyptophysa* sp. in Trial 3 and for *A. carinata* in Trial 4. Ravera (1991) states that developmental period of embryos exposed to heavy metals can be up to 4 times the normal period. In these trials, the nature of the observation process (following each egg mass through its full development) meant there a low number (3 to 6) of egg masses available for observation, and as a result large error terms were generated. Even if larger numbers of egg masses were exposed to each treatment, it would still be doubtful whether LOECs below 32% RP2 would be established. In all, developmental retardation was a moderately sensitive endpoint, and repeatability could be increased with a larger sample size.

### **Egg and egg mass production**

The fecundity parameters were seen to be the most sensitive of all endpoints examined. In Trials 5 and 6, LOECs established using egg mass and egg production by *A. carinata* and *A. cumingii* were between 1 and 10% RP2, more or less an order of magnitude more sensitive than any other. However, there existed a large discrepancy between the LOECs established by the egg mass production of *A. carinata* in Trials 5 and 6 (1 and 10% respectively). When daily means were taken it was seen that the selected animals were more productive in Trial 5. This was probably due to the thinning of the *A. carinata* stocks by the time of Trial 6, when smaller, less fecund individuals were used. The 32% LOECs established by *Glyptophysa* sp. highlighted its relative insensitivity. It is possible that the more robust shell of this species mitigated RP2 toxicity. Beeby and Richmond (1989) report the loss of magnesium from the

shell of terrestrial pulmonate *Helix aspersa* following lead exposure and postulate that this loss is part of a heavy metal detoxification mechanism.

Considering the validation exercise Trial 6, Lab and Creek results were in reasonable agreement. The species were more fecund in the creek water. The deterioration of water quality through storage is well documented for waters of the region, and a decrease in the quality of the lab water may have been responsible for the lower fecundity. The effects of storage on RP2 water might include co-precipitation of uranium with manganese (P. Cussons. *pers comm.*, 1991) or some change in the speciation of uranium. The higher flow rate in the creekside trial might also have contributed to this effect. But despite differences in flow rate and possible differences in water quality, effects observed in the lab were observed at the creekside. By considering the LOECs established for the fecundity parameters, it was seen that the lab results of Trials 5 and 6 were validated by the results of the creekside trial.

The uncertainties of the assumption of a normally distributed response because of low sample numbers must be taken into consideration. That consecutive and concurrent trials did establish comparatively low LOECs for *A. carinata* and *A. cumingii* indicated that the assumption of normality was not incorrectly taken.

The results of Trials 5 and 6 strongly indicated that the fecundity parameters for either *A. carinata* or *A. cumingii* were consistent and sensitive endpoints for the biological monitoring of RP2 water release into the Magela Ck system.

#### RP2/URANIUM COMPARISON

The results from this trial were somewhat contradictory. A significant difference existed in the 10% treatments in egg mass production for both *A. carinata* and *A. cumingii*. No difference was found at either 1% or 32% using egg mass production, and no difference was found in any of the three treatments using egg production. Juvenile mortality upon analysis of variance did not indicate significant differences between treatments for *A. carinata* but did indicate a significant interaction between treatments and concentrations for *A. cumingii*. The results indicated a greater toxicity of the 10% RP2 treatments, possibly caused by the lower average pH. In spite of this, the lack of significant difference between uranium and RP2 at 1 and 32% concentrations demonstrated that uranium and RP2 produced similar levels of effects in the species used, indicating that the observed toxicity of RP2 water was due to uranium.

## SEQUESTRATION OF URANIUM IN *A. CARINATA*

Ultrastructural studies showed that uranium had accumulated in the ovotestis of *A. carinata* which had been exposed for 7 days at 32% RP2 during Trial 5. The uranium was deposited in multivesicular bodies of squamous ciliated epithelial cells of the ovotestis. These are toward the opening of the acinis into the hermaphroditic duct. It appeared as spindle shaped crystals associated with other electron dense amorphous material. Some evidence suggested that the multivesicular bodies were involved in either endocytosis or exocytosis. The actual mechanism of the inhibition of egg production by uranium was not elucidated by this study. The sections studied did not include regions toward the bottom of the acinis where germ cell production occurs. Ravera (1991) states that the effects of cadmium on reproduction in *Biomphalaria glabrata* included degeneration of the male germ cells. Female germ cells did not show any alteration.

Other tissues analysed from 32% exposed *A. carinata*, were the kidney, the digestive gland (hepatopancreas), the mantle, the foot, the albumin gland and the hermaphroditic duct. Uranium was not detected in any of these tissues. A variety of metals including gold, lead and zinc was present in the mantle. The absence of uranium from the digestive gland was most probably a spurious result, possibly due to overfixation which resulted the friable sections. This organ is thought to be the site of haemoglobin production and blood processing in the pulmonates. Most of the literature on metal accumulation deals with the digestive gland, and only occasional papers investigate other tissue.

The results obtained conflict with Phillips (1980), who states that accumulation in the gonads is generally low in comparison to that in other tissues. However he cites results from Segar *et al.* (1971) who stated that nickel, cadmium, lead, zinc and chromium were accumulated in the gonads of the mussel *Modiolus modiolus* to a similar or greater extent than in the digestive gland.

Simkiss and Mason (1983) and Heneine *et al.* (1969) state that iron initially appears in the digestive gland and is progressively redistributed to the ovotestis. Johnson *et al.* (1962) report excretion of iron with the eggs of *B. glabrata*. Munzinger and Guarducci (1988) found that zinc is transferred to the embryos of *B. glabrata* from the parent. Developmental retardation and embryonic mortality greater than control were observed in eggs laid in control water from previously exposed adults.

The initial uptake of uranium is thought to occur in conjunction with calcium extraction from the water column (R. Jeffree *pers comm.* 1991). In the metabolic analogue hypothesis, various

metals are taken up along with calcium, especially in soft waters. (Jeffree & Simpson, 1984). Metals dissolved or transported in the blood are then removed from the animal's general metabolism by precipitation due to differential solubilities, and accumulate as granules which are not necessarily excreted. The toxicity of the xenobiotic metals is thus reduced by compartmentalisation away from the general metabolism (R. Jeffree *pers comm.* 1991). If this were the sole mechanism operating to deal with uranium in *A. carinata*, the uranium should be present in association with immobilised granules in the digestive gland or mantle, and not appear in tissues away from the major blood vessels, such as the ovotestis.

Although the results of microstructural analysis were tentative, the fact that uranium was located in the ovotestis indicated that the depression of fecundity was directly caused by a toxic effect of uranium on that organ. This reinforces the case for the fecundity endpoints of *A. carinata* and *A. cumingii* as endpoints for biological monitoring of RP2 waters.

## CONCLUSION

Of the 7 species examined, *A. carinata* and *A. cumingii* were most sensitive to RP2 water. When fecundity parameters were used as endpoints, LOECs of between 1 and 10% RP2 were established for these species. This was roughly equivalent to uranium concentrations of 17 to 170ppb total uranium. The endpoints embryonic mortality, juvenile mortality and adult mortality were seen to be insensitive or too variable for use in biological monitoring. Developmental retardation was a moderately sensitive endpoint and established LOECs at 32% RP2.

Other species tested were excluded on various grounds, including rearing and handling difficulties, insensitivity and variability in the measured parameter or response.

Uranium solutions of similar concentration to RP2 treatments were seen to be equivalently toxic. The depressed fecundity effect of RP2 water on *A. carinata* and *A. cumingii* was attributed to uranium accumulation in the ovotestis.

Since the fecundity parameters were linked directly to the accumulation of uranium in the snail tissue, the case is strong for the use of this endpoint in biological monitoring of RP2 release into the Magela Ck system using *A. carinata* or *A. cumingii*.



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## Appendix 1.a.1.

PRELIMINARY OBSERVATION No.1 COMMENCED 19/1/91 A. carinata NUMBERS OF EGGS LAID PER DAY PAGE 1					
	REP.1	REP.2	REP.3	REP.4	
19/1/91	20	19	21	15	
			7	19	
20/1/91	19	16	5	21	
			9		
			13		
21/1/91	20	15	5	17	
		14	22	16	
		18		23	
22/1/91	27	11	16	17	
	19	19		17	
	17	13		17	
	8	3		15	
	17			12	
23/1/91	0	0	0	0	
EGGMASS PER DAY	2.8	3	2.8	2.4	
MEAN	4.09	3.46	1.79	1.95	
STD	146	115	63.9	81.2	
CV					
EGGS PER DAY					
MEAN	29.4	25.6	21.4	37.8	
STD	33.8	20.4	13.9	30.3	
CV	115	79.9	65.1	80.2	
EGGS PER EGGMASS					
MEAN	18.4	14.2	11.8	17.2	
STD	5.24	5.02	4.65	2.99	
CV	28.5	35.3	54.7	17.4	
TOTAL EGGS PER EGGMASS (USING EACH DAY REPLICATE)					
MEAN					29.6
STD					14.2
CV					47.8
EGG MASS MEAN ACROSS REPS.	19/1/91	20/1/91	21/1/91	22/1/91	23/1/91
STD	2.5	1.75	2.25	7	0.25
CV	1.29	1.5	0.96	2.94	0.5
	51.6	85.7	42.6	0	200
EGG MEANS ACROSS REPLICATES	25.3	23	37.5	57	0
STD	7.09	8.91	16.8	32.7	0
CV	28.1	38.7	44.9	57.3	
CUMMULATIVE EGGMASS TOTAL	10	17	26	54	55
MEAN ACROSS CUMM'IVE REPS.	2.5	2.215	2.17	3.38	2.75
STD	1.29	1.36	1.19	2.73	2.75
CV	51.6	63.9	55.1	80.9	100
CUMMULATIVE EGG TOTAL	101	193	343	571	571
MEAN ACROSS CUMM'IVE REPS.	25.6	24.15	28.6	35.7	28.6
STD	7.09	7.55	12.5	22.1	24.5
CV	28.1	31.3	43.8	62	85.9

## Appendix 1.a.2.

PRELIMINARY OBSERVATION No.1 COMMENCED 19/1/91 A. cummingii NUMBERS OF EGGS LAID PER DAY PAGE 1								
	REP.1	REP.2	REP.3	REP.4	REP.5	REP.6	REP.7	
19/1/91	15	0	45	0	21	44	39	0
						33		
20/1/91	18	12	31	22	24	38	50	
	18	6	43	47		44	57	
	11	48	29	39				
	9	12	39					
	15		31					
21/1/91	23	10	47	52	13	40	44	
	18	10	45	21	19			
	15	16	51	25	26			
	8	12	41	25				
	18	13						
22/1/91	22	16	54	24	12	45	50	
		15	39	50				
				21				
				20				
				9				
				20				
				41				
				54				
23/1/91	30	15	38	38	12	47	43	
	23	16	37	48	19	35	43	
	28	15	41		6	39	50	
	25		41		24	24	46	
			34		26	40	37	
			54		17	27	58	
			45		12	28	44	
			45		13	23		
			36			32		
EM/DAY								
MEAN	3.2	3.2	4.2	3.2	3.1	3.2	2.2	
STD	2.05	1.92	3.27	2.95	2.85	3.35	2.77	
CV	64	60.1	68.1	92.2	91.8	105	126	
EGGS/DAY								
MEAN	61.2	39.4	184	106	55.4	115	104	
STD	41	25.6	126	85.7	53.6	108	126	
CV	67	59.8	68.4	80.7	96.7	94.1	121	
EGGS/EM								
MEAN	19.1	12.3	41.3	31.8	18.5	35.5	47.5	
STD	5.86	3.05	6.78	14.8	6.14	8.26	6.3	
CV	30.7	24.8	16.4	46.5	33.2	23.3	13.3	
TOTAL EGGS PER EGGMASS (USING EACH DAY REPLICATE)								
MEAN								29.6
STD								14.2
CV								47.8
EGG MASS MEANS ACROSS REPS.	19/1/91	20/1/91	21/1/91	22/1/91	23/1/91			
STD	1	3.14	3.14	2.57	6.14			
CV	1.15	1.57	2.27	2.64	3.13			
	115	50	72.2	103	51			
EGG MEANS ACROSS REPLICATES	32.3	86.1	84.6	57	199			
STD	44.1	49.8	45.5	32.7	137			
STD AS % OF MEAN	137	57.8	53.8	57.3	68.8			
CUMMULATIVE EGGMASS TOTAL	7	29	53	71	114			
MEAN ACROSS CUMM'IVE REPS.	1	2.07	2.52	2.54	3.26			
STD	1.15	1.73	1.83	2.01	2.66			
CV	115	83.5	72.7	79.2	87.1			
CUMMULATIVE EGG TOTAL	226	829	1421	1934	3327			
MEAN ACROSS CUMM'IVE REPS.	32.3	59.2	67.1	69.1	95.1			
STD	44.1	53.1	51	57.2	93.2			
CV	137	89.7	75.4	82.8	98.1			

## Appendix 1.a.3.

PRELIMINARY OBSERVATION No.1 COMMENCED 19/1/91 GLYPTOPHYSA SP. NUMBERS OF EGGS LAID PER DAY PAGE 1				
	REP.1	REP.2	REP.3	REP.4
19/1/91	0	9	4	4
		9		
		9		
		8		
20/1/91	0	12	12	7
		10	12	9
		10	9	5
		10		
21/1/91	5	12	4	8
		12		4
		12		
		11		
		13		
		10		
		9		
22/1/91	1	9	11	11
	9	13	13	13
		10	13	9
		7	11	3
		11	1	9
		12		8
		12		9
		13		
		13		
		11		
		14		
		9		

PRELIMINARY OBSERVATION No.1 COMMENCED 19/1/91  
GLYPHOPHYS A SP. NUMBERS OF EGGS LAID PER DAY PAGE 2

	REP.1	REP.2	REP.3	REP.4
EGG MASS PER DAY				
MEAN	0.6	6	1.8	3.2
STD	0.89	4.64	1.1	2.28
CV	149	77.3	60.8	71.3
EGGS PER DAY				
MEAN	3	64.4	18.8	22.8
STD	4.47	52.3	21	21.6
CV	149	81.2	112	93.2
EGGS PER EGG MASS				
MEAN	5	10.1	8.54	7.13
STD	4	3.2	4.41	2.25
CV	80	30	51.6	35.8

TOTAL EGGS PER EGG MASS (USING EACH DAY REPLICATE)

MEAN	8.95
STD	3.23
CV	36.1

	19/1/91	20/1/91	21/1/91	22/1/91	23/1/91
EGG MASS MEAN ACROSS REPS.	1.5	2.5	3	1.25	6.25
STD	1.73	1.73	3.37	1.26	4.99
CV	115	69.3	112	101	80
EGG MEANS ACROSS REPLICATES	10.8	24	27.8	8.5	65.3
STD	16.4	18.2	41.7	8.06	56
STD AS % OF MEAN	151	75.7	150	94.9	85.8
CUMULATIVE EGG MASS TOTAL	6	16	28	33	58
MEAN ACROSS CUMM'IVE REPS.	1.5	2	2.33	2.06	2.9
STD	1.73	1.65	2.27	2.06	3.21
CV	115	84.5	97.3	101	111
CUMULATIVE EGG TOTAL	43	139	250	284	545
MEAN ACROSS CUMM'IVE REPS.	10.8	17.4	20.8	17.8	27.3
STD	16.28	17.5	26.3	23.5	36.2
CV	151	101	126	132	133

Appendix 1.a.4.

PRELIMINARY OBSERVATION No.1 COMMENCED 19/1/91 PAGE 1  
CYRAULUS SP. NUMBERS OF EGGS LAID PER DAY

19/1/91	REP. 1	REP. 2	REP. 3	REP. 4
	3	3	9	0
	5		4	
			5	
			4	
20/1/91	5	4	13	4
	2	4	11	4
	4	8	5	5
	4	6	6	7
	11	5	5	7
	6	8	9	8
	1	6		8
	3	4		4
	5			9
	4			8
	3			
	7			
	10			
21/1/91	9	7	5	4
	6	3	13	3
	2	7	5	8
		5	10	7
		6	12	5
			5	9
			9	8
			2	8
				10
				5
				4
				9
				4
				10
				4
22/1/91	9	5	10	6
	6	5	7	6
	5	5	12	9
	7	9	5	2
	7	8	5	4
	4	6	6	10
	8	6		10
	9			10
	7			9
23/1/91	8	5	3	5
	8	6	5	9
	7	5	5	9
	7	7	6	7
	5	4	12	5
	6	9	7	5
	12	9	11	5
	4	9	6	4
	8	12	5	10
	11	4		4
	4	9		8
				4
				6
EGGMASS PER DAY				
MEAN	8.32	6.2	6.6	9.8
STD	13.8	3.42	1.95	6.14
CV	166	55.2	29.5	62.7
EGGS PER DAY				
MEAN	50.6	42	47.4	65
STD	35.2	38.1	58.1	39.8
CV	69.6	75.6	75.6	61.2
EGGS PER EGGMASS				
MEAN	6.17	6.16	7.18	7.52
STD	2.7	2.16	3.14	5.84
CV	43.8	35.3	43.7	77.6
TOTAL EGGS PER EGGMASS (USING EACH DAY REPLICATE)				
MEAN				6.36
STD				2.64
CV				41.5

CYRAULUS SP.

	19/1/91	20/1/91	21/1/91	22/1/91	23/1/91
EGG MASS MEAN ACROSS REPS.	1.75	9.5	7.75	8	12
STD	1.71	3.42	5.25	1.83	2.5
CV	97.6	36	67.8	22.8	20.4
EGG MEANS ACROSS REPLICATES	8.25	58.25	51	56.3	82.5
STD	9.74	13.8	36.5	13.7	16.1
CV	118	23.8	71.5	24.3	19.5
CUMULATIVE EGG MASS TOTAL	7	45	76	108	156
MEAN ACROSS CUMM'IVE REPS.	1.75	5.63	6.33	6.75	7.8
STD	1.71	4.84	4.84	4.3	4.49
CV	97.6	86	86	63.7	57.6
CUMULATIVE EGG TOTAL	33	266	470	695	1025
MEAN ACROSS CUMM'IVE REPS.	8.25	33.3	39.2	43.3	51.3
STD	9.74	28.9	31.2	28.4	30.6
CV	118	87	79.6	65.5	59.7

Appendix 1.a.5.

PRELIMINARY OBSERVATION No.1 COMMENCED 19/1/91  
HELIOCORBIS SP. NUMBERS OF EGGS LAID PER DAY PAGE 1

19/1/91	REP.1	REP.2	REP.3	REP.4	
	6	0	3	6	
	5			8	
				3	
20/1/91	0	0	5	2	
			2		
			2		
21/1/91	5	4	4	4	
		6	6	5	
			7	6	
			5	6	
				7	
22/1/91	2	4	3	5	
	4	4	4	6	
	5	4	5	6	
	5	4	2	5	
		5	2	4	
			2	3	
			3	4	
			6	5	
			3	6	
			4	8	
				8	
				8	
23/1/91	5	6	5	7	
	4	4	7	7	
	4	4	1	7	
	3	5	5	7	
	7	3	6	3	
	4	5	3	6	
	5	3	1	5	
	6		3		
MINARY OBSERVATION No.1 COMMENCED 19/1/91 PAGE 3					
CORBIS SP. DAILY AND CUMULATIVE STATISTICS					
	19/1/91	20/1/91	21/1/91	22/1/91	23/1/91
MASS MEAN ACROSS REPS.	1.5	1.5	4	7.75	7.75
	1.29	1.28	1.41	3.86	0.96
	86.1	86.1	35.4	49.8	12.4
MEANS ACROSS REPLICATES	7.75	2.75	16.3	35	36.8
	7.72	4.27	10.6	23.4	4.99
	99.6	155	65.2	66.9	13.6
LATIVE EGGMASS TOTAL	6	12	28	59	90
ACROSS CUMM'IVE REPS.	1.5	1.5	2.33	3.69	4.5
	1.29	1.2	1.72	3.32	3.41
	86.1	80	73.9	90.1	75.8
LATIVE EGG TOTAL	31	42	107	247	396
ACROSS CUMM'IVE REPS.	7.75	5.25	8.92	15.4	19.7
	7.72	6.36	9.26	17.6	18
	99.6	121	104	114	91.4

PRELIMINARY OBSERVATION No.1 COMMENCED 19/1/91  
HELIOCORBIS SP. NUMBERS OF EGGS LAID PER DAY PAGE 2

	REP.1	REP.2	REP.3	REP.4
EGGMASS PER DAY				
MEAN	4.2	2.8	5.4	5.6
STD	2.49	3.11	3.91	4.44
CV	59.3	111	72.4	75.3
EGGS PER DAY				
MEAN	14	12.2	21.1	31.4
STD	14.7	13.2	15.2	25.2
CV	105	1.8	71.6	80.2
EGGS PER EGGMASS				
MEAN	4.67	4.36	3.81	5.61
STD	1.23	0.93	1.92	1.69
CV	26.5	21.3	50.4	30.1
TOTAL EGGS PER EGGMASS (USING EACH DAY REPLICATE)				
MEAN				4.7
STD				1.67
CV				35.6

## Appendix 1.b.1.

PRELIMINARY OBSERVATION No.2 COMMENCED 30/1/91  
ENDPOINT:DEVELOPMENTAL CHARACTERISTICS PAGE 1  
SPECIES A. carinata

RAW DATA

	REPLICATE No. 1 DEVELOPMENTAL STAGE					H'LING	N'NATE
	GASTRULA	TROCH	VEL	HIPPO			
DAY 1							
DAY 2		14					
DAY 3		14 21 20					
DAY 4			14 15 16 14	23 23 21 20			
DAY 5				21 20 14	17 15 16 14 15 23 23 16		
DAY 6					13 18 17 15 16 14 15 16 23 23 21 20 14		
DAY 7					13 18 17 15 16 14 15 23 16 23 21 20 14		
DAY 8					13 18 14 15 23 16 23	17 15 16 21 20 14	
DAY 9					13 18 14 15 16 23 17 15 16 21 20 14		

PRELIMINARY OBSERVATION No.2 COMMENCED 30/1/91  
ENDPOINT:DEVELOPMENTAL CHARACTERISTICS PAGE 2  
SPECIES A. carinata

PERCENTILE  
DATA

	REPLICATE No. 1 DEVELOPMENTAL STAGE					H'LING	N'NATE
	GASTRULA	TROCH	VEL	HIPPO			
DAY 1	100						
DAY 2		100					
DAY 3		100					
DAY 4			40	60			
DAY 5				28	72		
DAY 6					100		
DAY 7					100		
DAY 8					57	43	
DAY 9						100	

PRELIMINARY OBSERVATION No.2 COMMENCED 30/1/91  
ENDPOINT:DEVELOPMENTAL CHARACTERISTICS PAGE 1  
SPECIES A. carinata

RAW DATA

	REPLICATE No. 2 DEVELOPMENTAL STAGE					H'LING	N'NATE
	GASTRULA	TROCH	VEL	HIPPO			
DAY 1							
DAY 2							
DAY 3		16 18					
DAY 4			16 16 18				
DAY 5			17 12	15 12 8 6 13	10 20 17 17 12 12 18		
DAY 6				17 10 15	12 20 10 20 17 12 12 18 15 16 16		
DAY 7					12 20 10 20 17 17 10 18 16	13 15 16	
DAY 8					17 20	12 20	
DAY 9						12 20 10 20 17 17	

PRELIMINARY OBSERVATION No.2 COMMENCED 30/1/91  
ENDPOINT:DEVELOPMENTAL CHARACTERISTICS PAGE 2  
SPECIES A. carinata

PERCENTILE  
DATA

	REPLICATE No. 2 DEVELOPMENTAL STAGE					H'LING	N'NATE
	GASTRULA	TROCH	VEL	HIPPO			
DAY 1	100						
DAY 2							
DAY 3		100					
DAY 4			100				
DAY 5			11	40	49		
DAY 6				15	85		
DAY 7					76	24	
DAY 8					24	76	
DAY 9						100	



PRELIMINARY OBSERVATION No.2 COMMENCED 30/1/91  
ENDPOINT:DEVELOPMENTAL CHARACTERISTICS PAGE 1  
SPECIES A. carinata

RAW DATA

	GASTRULA	TROCH	REPLICATE No. 3 DEVELOPMENTAL STAGE			H'LING	N'NATE
			VEL	HIPPO			
DAY 1							
DAY 2		12 13					
DAY 3			13 13 8 12 13				
DAY 4				15 12 6 8 13			
DAY 5				15 13 6 13	13 2 9 11 3		
DAY 6					13 2 8 15 13 11 4 6 13	16 13	
DAY 7							16 13 15 13
DAY 8							13 9 15 13

DAY 8	19 16 15 13 18	11 17 16 11 17 7 14
DAY 9		16 19 15 13 18

PRELIMINARY OBSERVATION No.2 COMMENCED 30/1/91  
ENDPOINT:DEVELOPMENTAL CHARACTERISTICS PAGE 2  
SPECIES A. carinata

PERCENTILE  
DATA

	GASTRULA	TROCH	REPLICATE No. 4 DEVELOPMENTAL STAGE			H'LING	N'NATE
			VEL	HIPPO			
DAY 1	100						
DAY 2		100					
DAY 3		28	72				
DAY 4			30	70			
DAY 5				80	20		
DAY 6					100		
DAY 7					96	4	
DAY 8					47	53	
DAY 9							100

PRELIMINARY OBSERVATION No.2 COMMENCED 30/1/91  
ENDPOINT:DEVELOPMENTAL CHARACTERISTICS PAGE 2  
SPECIES A. carinata

PERCENTILE  
DATA

	GASTRULA	TROCH	REPLICATE No. 3 DEVELOPMENTAL STAGE			H'LING	N'NATE
			VEL	HIPPO			
DAY 1	100						
DAY 2		100					
DAY 3			100				
DAY 4				100			
DAY 5				55	45		
DAY 6					100		
DAY 7					63	37	
DAY 8							100
DAY 9							

PRELIMINARY OBSERVATION No.2 COMMENCED 30/1/91  
ENDPOINT:DEVELOPMENTAL CHARACTERISTICS PAGE 1  
SPECIES A. cummingii

RAW DATA

	GASTRULA	TROCH	REPLICATE No. 1 DEVELOPMENTAL STAGE			H'LING	N'NATE
			VEL	HIPPO			
DAY 1							
DAY 2	34 25 13						
DAY 3	13	34 25					
DAY 4		27 31 13	33 34 25				
DAY 5			25 13	30 27 33 35 26 34			
DAY 6				26 30 25	35 33 34 25		
DAY 7					13 34 25 13 25 26 30 33		34
DAY 8							34 25 13 34 25 26 30 33

PRELIMINARY OBSERVATION No.2 COMMENCED 30/1/91  
ENDPOINT:DEVELOPMENTAL CHARACTERISTICS PAGE 1  
SPECIES A. carinata

RAW DATA

	GASTRULA	TROCH	REPLICATE No. 4 DEVELOPMENTAL STAGE			H'LING	N'NATE
			VEL	HIPPO			
DAY 1							
DAY 2		7 17 14					
DAY 3		11 7	16 17 14				
DAY 4			13 18	15 16 11 17 7 14			
DAY 5				15 13 18 16 11 17	11 17		
DAY 6					19 11 16 17 15 13 18 16 11 17 7 14		
DAY 7						7	
DAY 8							19 16 11 17 15 13 18 16 11 17 14

PRELIMINARY OBSERVATION No.2 COMMENCED 30/1/91  
ENDPOINT:DEVELOPMENTAL CHARACTERISTICS PAGE 2  
SPECIES A. cummingii

PERCENTILE  
DATA

	GASTRULA	TROCH	REPLICATE No. 1 DEVELOPMENTAL STAGE			H'LING	N'NATE
			VEL	HIPPO			
DAY 1	100						
DAY 2	100						
DAY 3	18	72					
DAY 4		44	56				
DAY 5			18	72			
DAY 6				37	63		
DAY 7					85	15	
DAY 8							100
DAY 9							

## Appendix 1.b.2.

PRELIMINARY OBSERVATION No.2 COMMENCED 30/1/91  
ENDPOINT:DEVELOPMENTAL CHARACTERISTICS PAGE 1  
SPECIES A. cummingii

RAW DATA

	REPLICATE No. 2 DEVELOPMENTAL STAGE					
	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY 1						
DAY 2						
DAY 3		28				
DAY 4		25	27 31 30			
DAY 5				28 34 30 25 31 26		
DAY 6				26	28 34 30 25 31	
DAY 7					28 34 30 25 31 26	
DAY 8						28 34 30 25 31 26

PRELIMINARY OBSERVATION No.2 COMMENCED 30/1/91  
ENDPOINT:DEVELOPMENTAL CHARACTERISTICS PAGE 2  
SPECIES A. cummingii PERCENTILE  
DATA

	REPLICATE No. 2 DEVELOPMENTAL STAGE					
	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY 1	100					
DAY 2						
DAY 3		100				
DAY 4		22	78			
DAY 5				100		
DAY 6				15	85	
DAY 7					100	
DAY 8						100

PRELIMINARY OBSERVATION No.2 COMMENCED 30/1/91  
ENDPOINT:DEVELOPMENTAL CHARACTERISTICS PAGE 1  
SPECIES A. cummingii

RAW DATA

	REPLICATE No. 3 DEVELOPMENTAL STAGE					
	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY 1						
DAY 2						
DAY 3		36 27				
DAY 4			24 31 36 27 41			
DAY 5				43 24 41 31 36 27		
DAY 6				31 27	43 24 31 46	24
DAY 7					43 24 41 31 36 27	24
DAY 8						36 27 31 41 24 43 24

PRELIMINARY OBSERVATION No.2 COMMENCED 30/1/91  
ENDPOINT:DEVELOPMENTAL CHARACTERISTICS PAGE 2  
SPECIES A. cummingii PERCENTILE  
DATA

	REPLICATE No. 3 DEVELOPMENTAL STAGE					
	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'N
DAY 1	100					
DAY 2						
DAY 3		100				
DAY 4			100			
DAY 5				100		
DAY 6					26	63
DAY 7						89
DAY 8						100

PRELIMINARY OBSERVATION No.2 COMMENCED 30/1/91  
ENDPOINT:DEVELOPMENTAL CHARACTERISTICS PAGE 1  
SPECIES A. cummingii

RAW DATA

	REPLICATE No. 4 DEVELOPMENTAL STAGE					
	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY 1						
DAY 2						
DAY 3		43				
DAY 3			43 34			
DAY 4				36 43 43 34		
DAY 5					43 43 36 36	
DAY 6					43 43 36 36	
DAY 7						43 43 36 34

PRELIMINARY OBSERVATION No.2 COMMENCED 30/1/91  
ENDPOINT:DEVELOPMENTAL CHARACTERISTICS PAGE 2  
SPECIES A. cummingii PERCENTILE  
DATA

	REPLICATE No. 4 DEVELOPMENTAL STAGE					
	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY 1	100					
DAY 2						
DAY 3		100				
DAY 4			100			
DAY 5				100		
DAY 6					100	
DAY 7					100	
DAY 8						100

## Appendix 1.b.3.

PRELIMINARY OBSERVATION No. 2 COMMENCED 30/1/91 RAW DATA  
ENDPOINT: DEVELOPMENTAL CHARACTERISTICS PAGE 1  
SPECIES GABBIA SP.

	GASTRULA	TROCH	REPLICATE 1 DEVELOPMENTAL STAGE				H'LING	N'NATE
			VEL	HIPPO				
DAY 1	1							
DAY 2	1							
DAY 3		1						
DAY 4		1						
DAY 5			1 1 2 1					
DAY 6			1	1 1 2 1				
DAY 7				1 1 1 1 1 1 2				
DAY 8				1 1 1 1				
DAY 9				1 2	1 1 1 1 2			
DAY 10					1 1 1 1 2			
DAY 11					1 1 1 1 1 2			
DAY 12					1 1 1 2			
DAY 13					1 1 1 2			
TERMINATION								

PRELIMINARY OBSERVATION No. 2 COMMENCED 30/1/91  
ENDPOINT: DEVELOPMENTAL CHARACTERISTICS PAGE 2  
SPECIES GABBIA SP. PERCENTILE DATA

	GASTRULA	TROCH	REPLICATE 1 DEVELOPMENTAL STAGE				H'LING	N'NATE
			VEL	HIPPO				
DAY 1	100							
DAY 2								
DAY 3		100						
DAY 4		100						
DAY 5			100					
DAY 6			14	86				
DAY 7				100				
DAY 8				100				
DAY 9				14	86			
DAY 10					100			
DAY 11					100			
DAY 12					100			
DAY 13					100			
TERMINATION								

PRELIMINARY OBSERVATION No. 2 COMMENCED 30/1/91 RAW DATA  
ENDPOINT: DEVELOPMENTAL CHARACTERISTICS PAGE 1  
SPECIES GABBIA SP.

	GASTRULA	TROCH	REPLICATE 3 DEVELOPMENTAL STAGE				H'LING	N'NATE
			VEL	HIPPO				
DAY 1								
DAY 2								
DAY 3								
DAY 4								
DAY 5		1						
DAY 6			1 1					
DAY 7			1 1 1 1 1 1 1					
DAY 9			1	1				
DAY 10			1 1 1 1 1 1 1				1	
DAY 11			1 1 1 1 1 1 1				1	
DAY 12			1 1 1 1 1 1					
DAY 13			1 1					1
TERMINATION			1 1 1					

PRELIMINARY OBSERVATION No. 2 COMMENCED 30/1/91  
ENDPOINT: DEVELOPMENTAL CHARACTERISTICS PAGE 2  
SPECIES GABBIA SP. PERCENTILE DATA

	GASTRULA	TROCH	REPLICATE 3 DEVELOPMENTAL STAGE				H'LING	N'NATE
			VEL	HIPPO				
DAY 1	100							
DAY 2								
DAY 3								
DAY 4								
DAY 5		100						
DAY 6			100					
DAY 7				100				
DAY 8				100				
DAY 9				87	13			
DAY 10				87				13
DAY 11				86				14
DAY 12				86				14
DAY 13				83				17
TERMINATION								

## Appendix 1.b.4.

PRELIMINARY OBSERVATION No. 2 COMMENCED 30/1/91 RAW DATA  
ENDPOINT: DEVELOPMENTAL CHARACTERISTICS PAGE 1  
SPECIES GLYPTOPHYSA SP.

	GASTRULA	TROCH	REPLICATE 1 DEVELOPMENTAL STAGE			H'LING	N'NATE
			VEL	HIPPO			
DAY 1							
DAY 2	11						
DAY 3		7 11					
DAY 4			8 7				
DAY 5				6 7			
DAY 6				1	5 1	13 12 5	
DAY 7					1	4 6 13 5 1	
DAY 8						13 6 4 5 1	

PRELIMINARY OBSERVATION No. 2 COMMENCED 30/1/91  
ENDPOINT: DEVELOPMENTAL CHARACTERISTICS PAGE 2  
SPECIES GLYPTOPHYSA SP. PERCENTILE DATA

	GASTRULA	TROCH	REPLICATE 1 DEVELOPMENTAL STAGE			H'LING	N'NATE
			VEL	HIPPO			
DAY 1	100						
DAY 2	100						
DAY 3		100					
DAY 4			100				
DAY 5				100			
DAY 6					9	16	81
DAY 7						3	97
DAY 8							100

PRELIMINARY OBSERVATION No. 2 COMMENCED 30/1/91 RAW DATA  
ENDPOINT: DEVELOPMENTAL CHARACTERISTICS PAGE 1  
SPECIES GLYPTOPHYSA SP.

	GASTRULA	TROCH	REPLICATE 3 DEVELOPMENTAL STAGE			H'LING	N'NATE
			VEL	HIPPO			
DAY 1							
DAY 2	2						
DAY 3		2					
DAY 4			2				
DAY 5					2		
DAY 6					2 3 3	10	
DAY 7					2	2	
DAY 8					3	10	
DAY 9					2 2 3 2 10	2 2 3 2 10	

PRELIMINARY OBSERVATION No. 2 COMMENCED 30/1/91  
ENDPOINT: DEVELOPMENTAL CHARACTERISTICS PAGE 2  
SPECIES GLYPTOPHYSA SP. PERCENTILE DATA

	GASTRULA	TROCH	REPLICATE 3 DEVELOPMENTAL STAGE			H'LING	N'NATE
			VEL	HIPPO			
DAY 1							
DAY 2	100						
DAY 3		100					
DAY 4			100				
DAY 5					100		
DAY 6					44	56	
DAY 7					29	71	
DAY 8					12	88	

PRELIMINARY OBSERVATION No. 2 COMMENCED 30/1/91 RAW DATA  
ENDPOINT: DEVELOPMENTAL CHARACTERISTICS PAGE 1  
SPECIES GLYPTOPHYSA SP.

	GASTRULA	TROCH	REPLICATE 4 DEVELOPMENTAL STAGE			H'LING	N'NATE
			VEL	HIPPO			
DAY 1							
DAY 2		1					
DAY 3		1	1				
DAY 4		1	1				
DAY 5		1		1			
DAY 6				1	2	1	
DAY 7					1	2	
DAY 8						1	
DAY 9						1	1

PRELIMINARY OBSERVATION No. 2 COMMENCED 30/1/91  
ENDPOINT: DEVELOPMENTAL CHARACTERISTICS PAGE 2  
SPECIES GLYPTOPHYSA SP. PERCENTILE DATA

	GASTRULA	TROCH	REPLICATE 4 DEVELOPMENTAL STAGE			H'LING	N'NATE
			VEL	HIPPO			
DAY 1							
DAY 2		100					
DAY 3		50	50				
DAY 4		50	50				
DAY 5		50		50			
DAY 6		25		50	25		
DAY 7				25	75		
DAY 8				50	50		
DAY 9					100		

PRELIMINARY OBSERVATION No. 2 COMMENCED 30/1/91 RAW DATA  
ENDPOINT: DEVELOPMENTAL CHARACTERISTICS PAGE 1  
SPECIES GYTRAUUS SP.

	GASTRULA	TROCH	REPLICATE 1 DEVELOPMENTAL STAGE			H'LING	N'NATE
			VEL	HIPPO			
DAY 1							
DAY 2	9						
DAY 3			9 6 9 11	9			
DAY 4					7 9 6 9 11 9 13 8		
DAY 5					9 6 9 11	6 13 8 11	
DAY 6						6 9 6 9 11 8 8 13 5 13 8 3	7
DAY 7						5 9 6 9 11 13 6	5 3 7 7 5
DAY 8						13 6	9 6 9 11 5
DAY 9							13

## Appendix 1.b.5.

PRELIMINARY OBSERVATION No. 2 COMMENCED 30/1/91  
ENDPOINT: DEVELOPMENTAL CHARACTERISTICS PAGE 2  
SPECIES CYRAULUS SP. PERCENTILE DATA

	GASTRULA	REPLICATE 1 DEVELOPMENTAL STAGE				H'LING	N'NATE
		TROCH	VEL	HIPPO			
DAY 1	100						
DAY 2	100						
DAY 3			80	20			
DAY 4				100			
DAY 5				44	56		
DAY 6					94	6	
DAY 7					69	31	
DAY 8					32	68	
DAY 9						100	

PRELIMINARY OBSERVATION No. 2 COMMENCED 30/1/91  
ENDPOINT: DEVELOPMENTAL CHARACTERISTICS PAGE 2  
SPECIES CYRAULUS SP. PERCENTILE DATA

	GASTRULA	TROCH	REPLICATE 2 DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY 1	100					
DAY 2		100				
DAY 3		13	87			
DAY 4				83	17	
DAY 5				28	72	
DAY 6					92	8
DAY 7					59	41
DAY 8						100

PRELIMINARY OBSERVATION No. 2 COMMENCED 30/1/91  
ENDPOINT: DEVELOPMENTAL CHARACTERISTICS PAGE 1  
SPECIES CYRAULUS SP. RAW DATA

	GASTRULA	TROCH	REPLICATE 2 DEVELOPMENTAL STAGE				H'LING	N'NATE
			VEL	HIPPO				
DAY 1								
DAY 2		7 9 9						
DAY 3			7 9 11 9 9 6 8					
DAY 4				7 9 9 5 10 7 8 8 9 9 9 8	12 8			
DAY 5				9 9 5 8 10 7 1	7 9 9 10 11 8 11 7 7 8 12 8 9 9 6			
DAY 6					7 9 9 9 5 8 10 7 6 12 8		9 7	
DAY 7					9 7 10 11 11 8 7 6 10 8 8	7 9 9 9 11 7 8 6 9 10 8 7 11 3		
DAY 8								9 5 8 10 9 3 9 10 11

	GASTRULA	TROCH	REPLICATE 3 DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY 1						
DAY 2	11 9	12				
DAY 3		11 9 8 8 9 9	12 4 11			
DAY 4				11 9 12 6 8 11 9 10 10 10 9 8		
DAY 5				11 8 9	9 12 6 8 11 9 12 10 9 8 9 4 9 9	
DAY 6					6 8 9 11 9 8 10 11 9 12 12 10 10	9 9
DAY 7					9 8 11 12 6 8 8 11 9 10 9	9 12 10 10 8 9 8 9
DAY 8					6 8 8 11 4	11 9 12 9 10 9 9 4
DAY 9						6 9 8 11 4

PRELIMINARY OBSERVATION No. 2 COMMENCED 30/1/91  
ENDPOINT: DEVELOPMENTAL CHARACTERISTICS PAGE 2  
SPECIES CYRAULUS PERCENTILE DATA

	REPLICATE 3 DEVELOPMENTAL STAGE					
	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY 1	100					
DAY 2	62	38				
DAY 3		65	35			
DAY 4				90	10	
DAY 5				20	80	
DAY 6					81	9
DAY 7					55	45
DAY 8					29	71
DAY 9						100

PRELIMINARY OBSERVATION No. 2 COMMENCED 30/1/91  
ENDPOINT: DEVELOPMENTAL CHARACTERISTICS PAGE 1  
SPECIES LYMNEAE SP. RAW DATA

	REPLICATE 2 DEVELOPMENTAL STAGE					
	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY 1						
DAY 2						
DAY 3			42			
DAY 4			42			
DAY 5				35 42		
DAY 6				35 60	42	
DAY 7					35 42 60 124	
DAY 8					35 42 60 124	
DAY 9					35 42 60 124	
DAY 10					60	35 42 124
DAY 11					60	
DAY 12					60	
DAY 13						60

PRELIMINARY OBSERVATION No. 2 COMMENCED 30/1/91  
ENDPOINT: DEVELOPMENTAL CHARACTERISTICS PAGE 2  
SPECIES LYMNEAE SP. PERCENTILE DATA

	REPLICATE 2 DEVELOPMENTAL STAGE					
	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY 1	100					
DAY 2						
DAY 3			100			
DAY 4			100			
DAY 5				100		
DAY 6				69	31	
DAY 7					100	
DAY 8					100	
DAY 9					100	
DAY 10					30	70
DAY 11					30	
DAY 12					30	
DAY 13					30	

PRELIMINARY OBSERVATION No. 2 COMMENCED 30/1/91  
ENDPOINT: DEVELOPMENTAL CHARACTERISTICS PAGE 1  
SPECIES LYMNEAE SP. RAW DATA

	REPLICATE 3					
	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY 1						
DAY 2						
DAY 3						
DAY 4					46 34	
DAY 5					46 34 30	
DAY 6					46	

30  
57  
32  
124

DAY 7	46 34 30 57 32 124	
DAY 8	46 34 30	57 32 124
DAY 9	46 34 30	
DAY 10		46 34 30

PRELIMINARY OBSERVATION No. 2 COMMENCED 30/1/91  
ENDPOINT: DEVELOPMENTAL CHARACTERISTICS PAGE 2  
SPECIES LYMNEAE SP. PERCENTILE DATA

	REPLICATE 3 DEVELOPMENTAL STAGE					
	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY 1	100					
DAY 2						
DAY 3						
DAY 4					100	
DAY 5					100	
DAY 6					100	
DAY 7					100	
DAY 8					34	66
DAY 9					34	
DAY 10						34

PRELIMINARY OBSERVATION No. 2 COMMENCED 30/1/91  
ENDPOINT: DEVELOPMENTAL CHARACTERISTICS PAGE 1  
SPECIES LYMNEAE SP. RAW DATA

	REPLICATE 4 DEVELOPMENTAL STAGE					
	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY 1						
DAY 2						
DAY 3			76			
DAY 4			76			
DAY 5				76 30 48		
DAY 6				30	76 48 46	
DAY 7					76 30 48 46 55 57	
DAY 8					76 30 48 46 55 57	
DAY 9					76 30 48 55	46 57
DAY 10					76 30 48	55
DAY 11					76 48	30
DAY 12					76	48
DAY 13						76

## Appendix 1.b.6.

PRELIMINARY OBSERVATION No. 2 COMMENCED 30/1/91  
ENDPOINT: DEVELOPMENTAL CHARACTERISTICS PAGE 2  
SPECIES LYMNEAE SP. PERCENTILE DATA

	GASTRULA	TROCH	REPLICATE 4 DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY 1	100					
DAY 2						
DAY 3			100			
DAY 4			100			
DAY 5				100		
DAY 6				15	85	
DAY 7					100	
DAY 8					100	
DAY 9					67	33
DAY 10					74	26
DAY 11					81	19
DAY 12					61	39
DAY 13						100

PRELIMINARY OBSERVATION No. 2 COMMENCED 30/1/91  
ENDPOINT: DEVELOPMENTAL EMBRYONIC MORTALITY  
SPECIES A. carinata RAW DATA

	GASTRULA	TROCH	REPLICATES 1, 2, 3, 4.		H'LING	N'NATE
			VEL	HIPPO		
REP 1		1				
REP 2	2		3	5		
REP 3	5	2	25	37		
REP 4						
PERCENTILE DATA						
	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
REP 1		4.4				
REP 2	1.2		1.7	2.9		
REP 3	3.6	1.5	18.2	27		
REP 4						
TOTAL	7	3	28	42	0	0
PERCENT.	8.8	3.8	35	52.5	0	0

PRELIMINARY OBSERVATION No. 2 COMMENCED 30/1/91  
ENDPOINT: DEVELOPMENTAL EMBRYONIC MORTALITY  
SPECIES A. cumingii RAW DATA

	GASTRULA	TROCH	REPLICATES 1, 2, 3, 4.		H'LING	N'NATE
			VEL	HIPPO		
REP 1		2	4		1	
REP 2		2	3			
REP 3		8	2	2		
REP 4	4		1			
PERCENTILE DATA						
	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
REP 1		0.9	1.8		0.4	
REP 2		1	3			
REP 3		3	1	1		
REP 4	2		11			
TOTAL	4	12	10	2	1	0
PERCENT.	13.8	41.4	34.5	6.9	3.4	0

PRELIMINARY OBSERVATION No. 2 COMMENCED 30/1/91  
ENDPOINT: DEVELOPMENTAL EMBRYONIC MORTALITY  
SPECIES GABBLA SP. RAW DATA

	GASTRULA	TROCH	REPLICATE 1, 3		H'LING	N'NATE
			VEL	HIPPO		
REP 1		2				
REP 3		1	1	3		
PERCENTILE DATA						
	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
REP 1		22				
REP 3		10	10	30		
TOTAL	0	3	1	3	0	0
PERCENT.	0	42.9	14.3	42.9	0	0

PRELIMINARY OBSERVATION No. 2 COMMENCED 30/1/91  
ENDPOINT: DEVELOPMENTAL EMBRYONIC MORTALITY  
SPECIES GLYPTOPHYSA SP. RAW DATA

	GASTRULA	TROCH	REPLICATE 1, 2, 3, 4.		H'LING	N'NATE
			VEL	HIPPO		
REP 1		21	2	6	7	
REP 2		39	1			
REP 3		18			1	
REP 4		6			2	
PERCENTILE DATA						
	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
REP 1		32	3	9	11	
REP 2		98	2			
REP 3		50			2.8	
REP 4		60			20	
TOTAL	0	84	3	6	10	0
PERCENT.	0	81.2	2.9	5.8	9.7	0

PRELIMINARY OBSERVATION No. 2 COMMENCED 30/1/91  
ENDPOINT: DEVELOPMENTAL EMBRYONIC MORTALITY  
SPECIES CYRAULUS SP. RAW DATA

	GASTRULA	TROCH	REPLICATE 1, 2, 3, 4777.		H'LING	N'NATE
			VEL	HIPPO		
REP 1		33	2	8	1	
REP 2		1	11	30	4	
REP 3		8			2	
REP 4						
PERCENTILE DATA						
	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
REP 1		20.8	1.3	5	0.6	
REP 2		0.4	4.7	12.8	1.7	
REP 3		4			1	
REP 4						
TOTAL	0	42	13	38	7	0
PERCENT.	0	42	13	38	7	0

NOTE: LYMNEAE HAD NO OBSERVABLE EMBRYONIC MORTALITY

## Appendix 1.c.1.

PRELIMINARY OBSERVATIONS No. 2 COMMENCED 25/1/91

A. carinata NUMBERS OF EGGS LAID PER DAY PAGE 1

	REP.1	REP.2	REP.3	REP.4
25/1/91	18 13	12	0	16 19
26/1/91	16 15 17	20 17 10	16 9 13	17 11
27/1/91	23 23 15 17 14	18 15 12 17	15 18 13	18 13 15
28/1/91	21 20	16	8 15 13	16 11
29/1/91	14	18 18	17 13	7 14 17

EGG MASS PER DAY

MEAN	2.6	2.2	2	2.4
STD	1.52	1.3	1.22	0.55
STD % OF MEAN	58.3	59.3	61.2	22.8

EGGS PER DAY

MEAN	45.2	34.6	27.4	34.8
STD	29.1	21	15.6	7.99
STD % OF MEAN	64.4	60.8	57	22.4

EGGS PER EGG MASS

MEAN	17.4	15.7	13.7	14.5
STD	3.4	3.13	3.23	3.48
STD % OF MEAN	19.6	19.9	23.6	24

TOTAL EGGS PER EGG MASS (USING EACH DAY/REPLICATE)

MEAN				15.3
STD				3.51
STD % OF MEAN				22.8

PRELIMINARY OBSERVATIONS No. 2 COMMENCED 25/1/91 PAGE 2  
A. carinata DAILY AND CUMULATIVE STATISTICS

	25/1/91	26/1/91	27/1/91	28/1/91	29/1/91
EGG MASS MEANS ACROSS REPS.	1.25	2.75	3.5	2	2
STD	0.96	0.5	1.29	0.82	0.82
STD AS % OF MEAN	76.7	18.1	36.9	40.8	40.8
EGG MEAN ACROSS REPS.	19.5	40.3	58.3	30	29.5
STD	16.4	9.32	25.4	11	10.9
STD AS % OF MEAN	84.2	23.2	43.7	36.7	36.9
CUMM'IVE VE EGG TOTAL	5	16	30	38	46
MEAN ACROSS CUMM'IVE REPS.	1.25	2	2.5	2.38	2.3
STD	0.96	1.07	1.31	1.2	1.13
STD AS % OF MEAN	76.6	53.5	52.6	50.7	49.1
CUMMULATIVE EGG TOTAL	78	239	472	592	710
MEAN ACROSS CUMM'IVE REPS.	19.5	29.9	39.3	37	35.5
STD	16.4	16.6	23.4	21	19.4
STD AS % OF MEAN	84.2	55.6	59.5	56.9	54.7

## Appendix 1.c.2.

PRELIMINARY OBSERVATIONS No. 2 COMMENCED 25/1/91

A. cumingi NUMBERS OF EGGS LAID PER DAY PAGE 1

	REP.1	REP.2	REP.3	REP.4
25/1/91	0	0	26	0
26/1/91	37 26	34 28	45	37 43
27/1/91	35 31 38	31 33 25	31 41 32	32
28/1/91	0	28	27 36	4
29/1/91	13 25 35	0	0	0

EGG MASS PER DAY

MEAN	1.6	1.2	1.4	1
STD	1.52	1.3	1.14	1
STD % OF MEAN	94.8	109	81.4	100

EGGS PER DAY

MEAN	48	35.8	47.6	31.8
STD	46.4	39.2	39.2	33.8
STD % OF MEAN	96.6	109	82.4	106

EGGS PER EGG MASS

MEAN	30	29.8	34	31.8
STD	8.4	3.4	7.07	16.2
STD % OF MEAN	28	11.5	20.8	51

TOTAL EGGS PER EGG MASS (USING EACH DAY/REPLICATE)

MEAN				31.4
STD				8.9
STD % OF MEAN				28.4

PRELIMINARY OBSERVATIONS No. 2 COMMENCED 25/1/91 PAGE 2  
A. cumingi DAILY AND CUMULATIVE STATISTICS

	25/1/91	26/1/91	27/1/91	28/1/91	29/1/91
EGG MASS MEAN ACROSS REPS.	0.25	1.75	2.5	1.25	0.75
STD	0.5	0.5	1	0.96	1.5
STD AS % OF MEAN	200	28.6	40	76.6	200
EGG MEAN ACROSS REPS.	6.5	62.5	82.3	34.5	18.25
STD	13	14.3	34.2	27.1	36.5
STD AS % OF MEAN	200	22.9	41.6	78.6	200
CUMM'IVE EGG MASS TOTAL	1	8	18	23	26
MEAN ACROSS CUMM'IVE REPS.	0.25	1	1.5	1.44	1.3
STD	0.5	0.93	1.17	1.09	1.17
STD AS % OF MEAN	200	92.6	77.8	76	90.3
CUMMULATIVE EGG TOTAL	26	276	605	743	816
MEAN ACROSS CUMM'IVE REPS.	6.5	34.5	50.4	46.4	40.8
STD	13	32.5	39.3	36.5	37.3
STD AS % OF MEAN	200	94.2	77.9	78.5	91.5

## Appendix 1.c.3.

PRELIMINARY OBSERVATIONS No. 2 COMMENCED 25/1/91

GABBIA SP. NUMBERS OF EGGS LAID PER DAY PAGE 1

	REP.1	REP.2	REP.3	REP.4
25/1/91	1	0	0	1
26/1/91	1 2 1 1	0	1 1 1 1	0
27/1/91	1	0	1	0
28/1/91	1	0	1	0
29/1/91	0	0	0	0

EGG MASS PER DAY

MEAN	1.4	0	1.8	0.2
STD	1.52	0	2.95	0.45
STD % OF MEAN	108		164	224

EGGS PER DAY

MEAN	1.6	0	1.8	0.2
STD	1.95	0	2.95	0.45
STD % OF MEAN	122		164	224

EGGS PER EGG MASS

MEAN	1.14		1	1
STD	0.38		0	0
STD % OF MEAN	33		0	0

TOTAL EGGS PER EGG MASS (USING EACH DAY/REPLICATE)

MEAN				1.06
STD				0.24
STD % OF MEAN				22.9

PRELIMINARY OBSERVATIONS No. 2 COMMENCED 25/1/91 PAGE 2  
GABBIA SP. DAILY AND CUMULATIVE STATISTICS

	25/1/91	26/1/91	27/1/91	28/1/91	29/1/91
EGG MASS MEAN ACROSS REPS.	0.5	2.75	0.5	0.5	0
STD	0.57	3.4	0.58	0.58	0
STD AS % OF MEAN	115	124	115	115	
EGG MEAN ACROSS REPLICATES	0.5	3	0.5	0.5	0
STD	0.58	3.56	0.58	0.58	0
STD AS % OF MEAN	115	119	115	115	
CUMM'IVE EGG MASS TOTAL	2	13	15	17	17
MEAN ACROSS CUMM'IVE REPS.	0.5	1.63	1.25	1.06	0.85
STD	0.58	2.56	2.14	1.88	1.73
STD AS % OF MEAN	115	158	171	177	203
CUMMULATIVE EGG TOTAL	2	14	16	18	18
MEAN ACROSS CUMM'IVE REPS.	0.5	1.75	1.33	1.3	9
STD	0.58	2.71	2.27	2	1.83
STD AS % OF MEAN	115	155	170	177	204



# Appendix 1.c.4.

PRELIMINARY OBSERVATIONS No. 2 COMMENCED 25/1/91

GLYPTOPHYSA SP. NUMBERS OF EGGS LAID PER DAY PAGE 1

	REP.1	REP.2	REP.3	REP.4
25/1/91	9 15 13 10	16 15 15 11	13 10 11	8
26/1/91	0	0	0	0
27/1/91	0	0	0	0
28/1/91	7	0	0	11
29/1/91	11	0	2	1
EGGMASS PER DAY				
MEAN	1.2	0.6	0.8	0.6
STD	1.64	1.34	1.3	0.55
STD % OF MEAN	137	224	163	91.3

EGGS PER DAY				
MEAN	13	9.2	7.2	4
STD	19.6	20.6	15	5.15
STD % OF MEAN	151	224	208	129
EGGS PER EGGMASS				
MEAN	10.8	15.3	9	6.67
STD	2.86	0.58	4.83	5.13
STD % OF MEAN	26.4	3.8	53.7	77
TOTAL EGGS PER EGGMASS (USING EACH DAY/REPLICATE)				
MEAN				10.44
STD				4.83
STD % OF MEAN				42

PRELIMINARY OBSERVATIONS No. 2 COMMENCED 25/1/91 PAGE 2  
GLYPTOPHYSA DAILY AND CUMULATIVE STATISTICS

	25/1/91	26/1/91	27/1/91	28/1/91	29/1/91
EGG MASS MEAN ACROSS REPS.	2.75	0	0	0.5	75
STD	1.26	0	0	0.58	0.5
STD AS % OF MEAN	45.8			115	56.7
EGG MEAN ACROSS REPLICATES	33.8	0	0	4.5	3.5
STD	18.2	0	0	5.45	5.07
STD AS % OF MEAN	53.8			121	145
CUMM'IVE EGG MASS TOTAL	2.75	11	11	13	15
MEAN ACROSS CUMM'IVE REPS.	2.75	1.38	0.92	0.81	0.75
STD	1.25	1.69	1.51	1.33	1.21
STD AS % OF MEAN	45.8	123	164	163	161
CUMMULATIVE EGG TOTAL	135	135	135	153	167
MEAN ACROSS CUMM'IVE REPS.	33.8	16.9	11.3	9.56	8.35
STD	18.2	21.6	19.1	16.8	15.3
STD AS % OF MEAN	53.8	128	170	176	183

## Appendix 1.c.5.

PRELIMINARY OBSERVATIONS No. 2 COMMENCED 25/1/91

GYRAULUS SP. NUMBERS OF EGGS LAID PER DAY PAGE 1

	REP.1	REP.2	REP.3	REP.4
25/1/91	10 8 8 15 8	9 10 8 8 7 8	9 6 9 8	11
26/1/91	11	10 11 8 7 7	9 4 10 9	11
27/1/91	13 8 8 8 9	10 7 8 8 12	5 12 10 10 9 8	11 8 9
28/1/91	6 9 11 6 9 11	12 9 9 8 6	6 8 8 11 10 9	12
29/1/91	9	7 9 9	11 9 12	11

EGGMASS PER DAY				
MEAN	3.4	5.4	4.6	1.6
STD	2.3	1.52	1.34	0.89
STD % OF MEAN	67.7	28.1	29.2	55.9

EGGS PER DAY				
MEAN	31.8	47	40.4	16.8
STD	20.3	13.9	11.5	7.79
STD % OF MEAN	63.8	29.6	28.5	46.3

EGGS PER EGGMASS				
MEAN	9.35	8.7	8.78	10.5
STD	2.3	1.56	2.04	1.31
STD % OF MEAN	24.8	18	23.2	12.5

TOTAL EGGS PER EGGMASS (USING EACH DAY/REPLICATE)				
MEAN				9.1
STD				1.84
STD % OF MEAN				20.2

PRELIMINARY OBSERVATIONS No. 2 COMMENCED 25/1/91 PAGE 2  
GYRAULUS SP. DAILY AND CUMULATIVE STATISTICS

	25/1/91	26/1/91	27/1/91	28/1/91	29/1/91
EGG MASS MEAN ACROSS REPS.	3.75	3.25	5.25	4.5	2
STD	2.06	2.22	1.71	2.38	1.15
STD AS % OF MEAN	55	68.2	32.5	53	57.7
EGG MEAN ACROSS REPLICATES	33.5	29.8	47.5	40	19.3
STD	16.7	18.3	14.5	19	11.1
STD AS % OF MEAN	50	61.5	30.6	47.6	57.6
CUMM'IVE EGG MASS TOTAL	15	28	49	67	75
MEAN ACROSS CUMM'IVE REPS.	3.75	3.5	4.08	4.2	3.75
STD	2.06	2	2.02	2.04	2.07
STD AS % OF MEAN	55	57.1	49.5	48.7	55.3
CUMMULATIVE EGG TOTAL	134	253	443	603	680
MEAN ACROSS CUMM'IVE REPS.	33.5	31.6	36.9	37.7	34
STD	16.7	16.3	17	16.9	17.4
STD AS % OF MEAN	50	51.7	46	44.9	51.2

## Appendix 1.c.6.

PRELIMINARY OBSERVATIONS No. 2 COMMENCED 25/1/91

LYMNEAE SP. NUMBERS OF EGGS LAID PER DAY PAGE 1

	REP.1	REP.2	REP.3	REP.4
25/1/91	0	97	34 57 124	55 57
26/1/91	0	60	30	46
27/1/91	0	35 124	0	48 30
28/1/91	0	0	0	0
29/1/91	0	42	0	0
EGGMASS PER DAY				
MEAN	0	1	0.8	1
STD		0.71	1.3	1
STD % OF MEAN		71	163	100
EGGS PER DAY				
MEAN	0	71.6	48.6	47.2
STD		60	92.8	49
STD % OF MEAN		83.9	191	104
EGGS PER EGGMASS				
MEAN		71.6	122	47.2
STD		37.9	130	10.7
STD % OF MEAN		52.9	107	22.6
TOTAL EGGS PER EGGMASS (USING EACH DAY/REPLICATE)				
MEAN				59.8
STD				32.2
STD % OF MEAN				53.9

PRELIMINARY OBSERVATIONS No. 2 COMMENCED 25/1/91 PAGE 2  
LYMNEAE SP. DAILY AND CUMULATIVE STATISTICS

	25/1/91	26/1/91	27/1/91	28/1/91	29/1/91
EGG MASS ACROSS REPS.	1.5	0.75	1	0	25
STD	1.29	0.5	1.15	0	0.5
STD AS % OF MEAN	86.1	66.7	116		200
EGG MEAN ACROSS REPLICATES	106	34	59.3	0	10.5
STD	87.2	25.8	76	0	21
STD AS % OF MEAN	82.6	75.8	128		200
CUMM'IVE EGG MASS TOTAL	6	9	13	13	14
MEAN ACROSS CUMM'IVE REPS.	1.5	1.125	1.08	0.81	0.7
STD	1.29	0.99	1	0.98	0.92
STD AS % OF MEAN	86.1	88.1	92	12.7	132
CUMMULATIVE EGG TOTAL	422	558	795	795	837
MEAN ACROSS CUMM'IVE REPS.	106	69.8	66.3	49.7	41.9
STD	87.2	70.7	69.2	66.2	61.6
STD AS % OF MEAN	82.6	101	104	133	147

PRELIMINARY OBSERVATION No. 2 COMMENCED 30/1/91  
ENDPOINT: DEVELOPMENTAL EMBRYONIC MORTALITY  
SPECIES GABBIA SP. RAW DATA

	GASTRULA	TROCH	REPLICATE 1, 3		H'LING	N'NATE
			VEL	HIPPO		
REP 1		2				
REP 3		1	1	3		
PERCENTILE DATA						
	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
REP 1		22				
REP 3		10	10	30		
TOTAL	0	3	1	3	0	0
PERCENT.	0	42.9	14.3	42.9	0	0

PRELIMINARY OBSERVATION No. 2 COMMENCED 30/1/91  
ENDPOINT: DEVELOPMENTAL EMBRYONIC MORTALITY  
SPECIES GLYPTOPHYSA SP. RAW DATA

	GASTRULA	TROCH	REPLICATE 1, 2, 3, 4.		H'LING	N'NATE
			VEL	HIPPO		
REP 1		21	2	6	7	
REP 2		39	1			
REP 3		18			1	
REP 4		6			2	
PERCENTILE DATA						
	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
REP 1		32	3	9	11	
REP 2		98	2			
REP 3		50			2.8	
REP 4		60			20	
TOTAL	0	84	3	6	10	0
PERCENT.	0	81.2	2.9	5.8	9.7	0

PRELIMINARY OBSERVATION No. 2 COMMENCED 30/1/91  
ENDPOINT: DEVELOPMENTAL EMBRYONIC MORTALITY  
SPECIES GYRAULUS SP. RAW DATA

	GASTRULA	TROCH	REPLICATE 1, 2, 3, 4, 777.		H'LING	N'NATE
			VEL	HIPPO		
REP 1		33	2	8	1	
REP 2		1	11	30	4	
REP 3		8			2	
REP 4						
PERCENTILE DATA						
	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
REP 1		20.8	1.3	5	0.6	
REP 2		0.4	4.7	12.8	1.7	
REP 3		4			1	
REP 4						
TOTAL	0	42	13	38	7	0
PERCENT.	0	42	13	38	7	0

NOTE: LYNCEAE HAD NO OBSERVABLE EMBRYONIC MORTALITY

## Appendix 1.d.

PRELIMINARY OBSERVATION No. 2 COMMENCED 30/1/91  
ENDPOINT: DEVELOPMENTAL EMBRYONIC MORTALITY  
SPECIES A. carinata RAW DATA

	GASTRULA	TROCH	REPLICATES 1, 2, 3, 4.		H'LING	N'NATE
			VEL	HIPPO		
REP 1		1				
REP 2	2		3	5		
REP 3	5	2	25	37		
REP 4						
PERCENTILE DATA						
	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
REP 1		4.4				
REP 2	1.2		1.7	2.9		
REP 3	3.6	1.5	18.2	27		
REP 4						
TOTAL	7	3	28	42	0	0
PERCENT.	8.8	3.8	35	52.5	0	0

PRELIMINARY OBSERVATION No. 2 COMMENCED 30/1/91  
ENDPOINT: DEVELOPMENTAL EMBRYONIC MORTALITY  
SPECIES A. cumingii RAW DATA

	GASTRULA	TROCH	REPLICATES 1, 2, 3, 4.		H'LING	N'NATE
			VEL	HIPPO		
REP 1		2	4		1	
REP 2		2	3			
REP 3		8	2	2		
REP 4	4		1			
PERCENTILE DATA						
	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
REP 1		0.9	1.8		0.4	
REP 2		1	3			
REP 3		3	1	1		
REP 4	2		11			
TOTAL	4	12	10	2	1	0
PERCENT.	13.8	41.4	34.5	6.9	3.4	0

# Appendix 2.a.1.

TRIAL 1 COMMENCED 18/1/91 PAGE 1  
ENDPOINT: DEVELOPMENTAL PERIOD DAYS  
SPECIES *A. carinacea*

TREATMENT	CONTROL 1		CONTROL 2	
	A	B	A	B
EGGMASS				
A	7	6	6	6
B	7	5	7	6
C	6			7
D	7			
E	7			
MEAN	6.8	5.5	6.5	6.33
STD	0.45	0.71	0.71	0.58

TREATMENT	0.3% RP2		0.3% RP2	
	A	B	A	B
EGGMASS				
A	7	6	7	6
B	6	7	7	7
C	6			7
MEAN	6.33	6.5	7	6.67
STD	0.58	0.71	0	0.58

TREATMENT	1% RP2		1% RP2	
	A	B	A	B
EGGMASS				
A	6	6	6	7
B	6	7	6	7
C	6	6	7	6
D	6			
E	5			
MEAN	5.8	6.33	6.33	6.67
STD	0.45	0.58	0.58	0.58

TREATMENT	3.2% RP2		3.2% RP2	
	A	B	A	B
EGGMASS				
A	7	7	7	8
B	6	8	7	7
C		8		7
D		6		
E		8		
MEAN	6.5	7.4	7	7.33
STD	0.71	0.89	0	0.58

TREATMENT	10% RP2		10% RP2	
	A	B	A	B
EGGMASS				
A	7	8	6	6
B	7	7	7	6
C	7	7		
D		7		
E		7		
MEAN	7	7.2	6.5	6
STD	0	0.45	0.71	0

TREATMENT	32% RP2		32% RP2	
	A	B	A	B
EGGMASS				
A	7	7	7	7
B	7	5	7	6
C		5		
D		7		
E		7		
F		7		
MEAN	7	6.33	7	6.5
STD	0	1.03	0	0.45

## Appendix 2.a.2.

TRIAL 1 COMMENCED 18/1/91 PAGE 1  
ENDPOINT: DEVELOPMENTAL PERIOD DAYS  
SPECIES *A. cummingii*

TREATMENT	CONTROL 1		CONTROL 2	
	A	B	A	B
EGGMASS				
A	7	7	7	7
B		7	7	7
C		6		
MEAN	7	6.67	7	7
STD		0.58	0	0

TREATMENT	0.3% RP2		0.3% RP2	
	A	B	A	B
EGGMASS				
A	7	6	7	7
B	7	6	7	7
C	8	7		
D	7	6		
E		7		
MEAN	7.25	6.4	7	7
STD	0.5	0.55	0	0

TREATMENT	1% RP2		1% RP2	
	A	B	A	B
EGGMASS				
A	7	7	8	7
B	7	6	7	7
C	7			7
MEAN	7	6.5	7.5	7
STD	0	0.71	0.71	0

TREATMENT	3.2% RP2		3.2% RP2	
	A	B	A	B
EGGMASS				
A	7	7	8	7
B	6	7	7	7
C	7	7		
D	7			
E	6			
F	6			
G	6			
H	6			
MEAN	6.38	7	8.5	7
STD	0.52	0	0.71	0

TREATMENT	10% RP2		10% RP2	
	A	B	A	B
EGGMASS				
A	7	7	8	7
B		8	8	7
C		6		
MEAN	7	7	8	7
STD		1	0	0

TREATMENT	32% RP2		32% RP2	
	A	B	A	B
EGGMASS				
A	7	8	8	7
B	7	7	8	8
C		7		
MEAN	7	7.33	8	7.5
STD	0	0.58	0	0.71

## Appendix 2.a.3.

TRIAL 1 COMMENCED 18/1/91 PAGE 1  
ENDPOINT: DEVELOPMENTAL PERIOD DAYS  
SPECIES *GLYPHOPHYSA* SP.

TREATMENT	CONTROL 1		CONTROL 2	
	A	B	A	B
EGGMASS				
A	5	5	7	6
B	4	4	6	6
C		6		6
D		5		6
E		4		
F		6		
MEAN	4.5	5	6.5	6
STD	0.71	0.89	0.71	0

TREATMENT	0.3% RP2		0.3% RP2	
	A	B	A	B
EGGMASS				
A	5	6	5	6
B	6	6	6	6
C	6	7	6	7
D	6	6		6
E		7		6
F		7		6
MEAN	6.75	6.6	5.67	6.17
STD	0.5	0.55	0.58	0.41

TREATMENT	1% RP2		1% RP2	
	A	B	A	B
EGGMASS				
A	7	6	6	5
B	7	5	6	6
C	7		6	6
D	7		6	
E	7			
F	6			
G	6			
MEAN	6.71	5.5	6	6
STD	0.49	0.71	0	0

TREATMENT	3.2% RP2		3.2% RP2	
	A	B	A	B
EGGMASS				
A	6	7	6	5
B	7	7	6	6
C	7	7	6	5
D				
E	6			
F	6			
G	6			
H	6			
MEAN	6.38	7	6	5.67
STD	0.52	0	0	0.58

TREATMENT	10% RP2		10% RP2	
	A	B	A	B
EGGMASS				
A	6	6	5	6
B	6		6	6
C		7	5	6
D		6		
E		6		
F		6		
G		6		
H		7		
I		6		
MEAN	6	6.22	5.67	6
STD	0	0.44	0.58	0

TREATMENT	32% RP2		32% RP2	
	A	B	A	B
EGGMASS				
A	7	7	6	5
B	7	6	7	6
C		6		5
D		7		5
E		7		
F		7		
MEAN	7	6.67	6.5	5.25
STD	0	0.52	0.71	0.5

# Appendix 2.b.1.

TRIAL 1 DATE 18/2/91  
ENDPOINT: JUVENILE MORTALITY EXPECTED: FROM NUMBERS OF  
SPECIES A. carinata NEONATES HATCHED

TREATMENT	REPLICATE 1			REPLICATE 2		
	LIVING	DEAD	% MORT	LIVING	DEAD	% MORT
CONTROL	9	11	55	54	44	45
0.3% RP2	21	38	64	12	19	61
1% RP2	10	58	85	15	14	48
3.2% RP2	7	24	77	27	38	58
10% RP2	17	8	32	42	39	48

32% RP2	26	53	67	17	11	39
---------	----	----	----	----	----	----

TREATMENT	REPLICATE 1			REPLICATE 2		
	LIVING	DEAD	% MORT	LIVING	DEAD	% MORT
CONTROL	10	23	70	12	31	72
0.3% RP2	16	15	48	22	31	58
1% RP2	11	31	74	20	18	47
3.2% RP2	20	31	61	6	25	81
10% RP2	22	13	37	16	27	63
32% RP2	6	36	86	13	23	64

# Appendix 2.b.2.

TRIAL 1 DATE 18/2/91  
ENDPOINT: JUVENILE MORTALITY EXPECTED: FROM NUMBERS OF  
SPECIES A. cunningii NEONATES HATCHED

TREATMENT	REPLICATE 1			REPLICATE 2		
	LIVING	DEAD	% MORT	LIVING	DEAD	% MORT
CONTROL	15	17	53	26	63	71
0.3% RP2	51	59	54	35	5	13
1% RP2	10	25	71	38	30	50
3.2% RP2	21	7	25	68	51	43
10% RP2	15	18	55	32	51	61
32% RP2	18	24	65	10	64	86

TREATMENT	REPLICATE 1			REPLICATE 2		
	LIVING	DEAD	% MORT	LIVING	DEAD	% MORT
CONTROL	32	23	43	27	19	41
0.3% RP2	21	25	54	28	27	49
1% RP2	36	28	44	27	3	10
3.2% RP2	30	20	40	22	28	56
10% RP2	36	23	39	10	33	77
32% RP2	26	19	42	38	21	36

# Appendix 2.b.3.

TRIAL 1 DATE 18/2/91  
ENDPOINT: JUVENILE MORTALITY OBSERVED: NUMBERS RECORDED  
SPECIES *Glyptophysa* sp. AT TERMINATION

TREATMENT	REPLICATE 1			REPLICATE 2		
	LIVING	DEAD	% MORT	LIVING	DEAD	% MORT
CONTROL	9	15	63	1	5	83
0.3% RP2	26	0	0	15	0	0
1% RP2	5	2	29	6	2	25
3.2% RP2	4	6	60	4	1	20
10% RP2	28	11	28	6	1	14
32% RP2	2	12	14	25	14	36

TREATMENT	REPLICATE 1			REPLICATE 2		
	LIVING	DEAD	% MORT	LIVING	DEAD	% MORT
CONTROL	8	1	11	9	1	10
0.3% RP2	28	7	20	21	9	30
1% RP2	14	1	7	6	5	45
3.2% RP2	9	1	10	14	1	7
10% RP2	7	3	30	8	3	27
32% RP2	12	4	25	6	7	54

# Appendix 2.c.

TRIAL 11 COMMENCED 18/2/91  
PHYSICO-CHEMICAL DATA

DISSOLVED OXYGEN mg/L												
	TREATMENT											
	CON.	CON.	0.3%RP2	0.3%RP2	1%RP2	1%RP2	3.2%RP2	3.2%RP2	10%RP2	10%RP2	32%RP2	32%RP2
DAY1	7.7	7	6.9	7	6.8	6.8	6.9	6.4	6.8	6.3	6.3	7
DAY2	6.9	7.7	7.6	7.1	7.4	7.7	7.1	7.6	7.1	7.6	7.4	7.7
DAY3	6.8	7.3	6.8	6.9	6.6	6.3	6.8	6.6	6.6	6.8	7	7.1
DAY4	7.1	7.1	7.2	7	6.9	7.1	6.9	6.9	7.1	7.1	7.1	7.1
DAY5	7.2	7.1	6.8	9.4	7	6.9	7.1	7.3	7.1	6.9	7	7.2
DAY6	7	6.9	6.9	7.1	7.2	6.9	7	7.1	6.8	7.1	7.1	7
DAY7	6.9	7.2	7	6.9	6.8	6.9	7	6.6	6.7	7.1	7	6.8
pH												
	TREATMENT											
	CON.	CON.	0.3%RP2	0.3%RP2	1%RP2	1%RP2	3.2%RP2	3.2%RP2	10%RP2	10%RP2	32%RP2	32%RP2
DAY1	6.47	6.44	6.52	6.46	6.59	6.38	6.48	6.52	6.42	6.46	6.74	6.74
DAY2	6.38	6.33	6.43	6.33	6.57	6.52	6.44	6.56	6.42	6.56	6.81	6.75
DAY3	6.47	6.4	6.5	6.49	6.37	6.4	6.55	6.43	6.57	6.48	6.78	6.68
DAY4	6.36	6.42	6.43	6.38	6.53	6.42	6.56	6.44	6.51	6.49	6.73	6.72
DAY5	6.46	6.41	6.46	6.47	6.46	6.49	6.55	6.51	6.57	6.54	6.83	6.84
DAY6	6.07	6.44	6.44	6.52	6.59	6.45	6.59	6.54	6.42	6.54	6.78	6.82
DAY7	6.34	6.25	6.41	6.48	6.58	6.56	6.5	6.47	6.52	6.48	6.83	6.75
CONDUCTIVITY uS/cm												
	TREATMENT											
	CON.	CON.	0.3%RP2	0.3%RP2	1%RP2	1%RP2	3.2%RP2	3.2%RP2	10%RP2	10%RP2	32%RP2	32%RP2
DAY1	19.7	20.4	26.2	25.2	36.8	38.8	73.4	74.2	178	177	466	461
DAY2	17.7	17.9	23.7	24.6	34	35.6	71.2	72	174	173	452	456
DAY3	19.1	20	24.1	23.9	35.6	36.8	71.7	70.8	174	173	451	452
DAY4	18.3	18.3	24.1	23.9	36.4	36.4	70	70	173	173	456	456
DAY5	18.2	20.8	23.9	24.3	36.3	36.1	70.7	70.3	175	174	461	460
DAY6	18.1	17.9	23.5	23.9	36.1	35.8	70.4	70.1	173	174	460	459
DAY7	20.3	20.9	24.1	24.8	36	36.1	70.5	71	175	175	461	461

## Appendix 3.a.1.

TRIAL #2 COMMENCED 28/2/91  
 ENDPOINT DEVELOPMENTAL CHARACTERISTICS PAGE 1  
 EGG NUMBERS RAW DATA  
 SPECIES A. cummingii

TREATMENT: CONTROL		REPLICATE 1		SAMPLE A		
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	21					
DAY2		21				
DAY3				21		
DAY4				5	16	
DAY5					21	
DAY6					21	
DAY7						21

TREATMENT: CONTROL		REPLICATE 1		SAMPLE B		
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	42					
DAY2		42				
DAY3			42			
DAY4			21	21		
DAY5					41	
DAY6					41	
DAY7					41	
DAY8						41

TREATMENT: CONTROL		REPLICATE 2		SAMPLE A		
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	23					
DAY2		22				
DAY3				21		
DAY4					21	
DAY5					21	
DAY6					21	
DAY7					21	
DAY8						21

TREATMENT: CONTROL		REPLICATE 2		SAMPLE B		
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	38					
DAY2		34				
DAY3			34			
DAY4					31	
DAY5					31	
DAY6					31	
DAY7					31	
DAY8						31

TREATMENT: 1% RP2		REPLICATE 1		SAMPLE A		
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	38					
DAY2		37				
DAY3				37		
DAY4					37	
DAY5					37	
DAY6					37	
DAY7					37	
DAY8						37

TREATMENT: 1% RP2		REPLICATE 1		SAMPLE B		
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	22					
DAY2		2	20			
DAY3		1		21		
DAY4					21	
DAY5					21	
DAY6					21	
DAY7					21	
DAY8						21

TREATMENT: 1% RP2		REPLICATE 2		SAMPLE A		
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	24					
DAY2		24				
DAY3				24		
DAY4					24	
DAY5					24	
DAY6					24	
DAY7					24	
DAY8						24

TREATMENT: 1% RP2		REPLICATE 2		SAMPLE B		
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	33					
DAY2	2	31				
DAY3				31		
DAY4					31	
DAY5					31	
DAY6					31	
DAY7					31	
DAY8						31

TREATMENT: 3.2% RP2		REPLICATE 1		SAMPLE A		
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	22					
DAY2		22				
DAY3				22		
DAY4				22		
DAY5					22	
DAY6					22	
DAY7						22

TREATMENT: 3.2% RP2		REPLICATE 1		SAMPLE B		
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	30					
DAY2	2	28				
DAY3				29		
DAY4					29	
DAY5					29	
DAY6					29	
DAY7					29	
DAY8						29

TREATMENT: 3.2% RP2		REPLICATE 2		SAMPLE A		
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	27					
DAY2		27				
DAY3		1	26			
DAY4					27	
DAY5					27	
DAY6					27	
DAY7					27	
DAY8						27

TREATMENT: 3.2% RP2		REPLICATE 2		SAMPLE B		
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	29					
DAY2		29				
DAY3				27		
DAY4					27	
DAY5					27	
DAY6					27	
DAY7						27

TRIAL #2 COMMENCED 29/2/91  
 ENDPOINT DEVELOPMENTAL CHARACTERISTICS PAGE 7  
 EGG NUMBERS RAW DATA  
 SPECIES A. cunninghamii

TREATMENT: 10% RP2		REPLICATE 1		SAMPLE A		
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	34					
DAY2		34				
DAY3		14	14			
DAY4			4	24		
DAY5					24	
DAY6					24	
DAY7					24	
DAY8					24	
DAY9						24

TREATMENT: 10% RP2		REPLICATE 1		SAMPLE B		
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	25					
DAY2		25				
DAY3				25		
DAY4					25	
DAY5					25	
DAY6					25	
DAY7					25	
DAY8						25

TREATMENT: 10% RP2		REPLICATE 2		SAMPLE A		
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	25					
DAY2		25				
DAY3				25		
DAY4					25	
DAY5					25	
DAY6					25	
DAY7						25

TREATMENT: 10% RP2		REPLICATE 2		SAMPLE B		
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	31					
DAY2		23				
DAY3		17				
DAY4			4	13		
DAY5				4	13	
DAY6				4	13	
DAY7					13	
DAY8						13

TREATMENT: 32% RP2		REPLICATE 1		SAMPLE A		
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	31					
DAY2		31				
DAY3				31		
DAY4					31	
DAY5					31	
DAY6					31	
DAY7					31	
DAY8						31

TREATMENT: 32% RP2		REPLICATE 1		SAMPLE B		
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	26					
DAY2		25				
DAY3				24		
DAY4					24	
DAY5					24	
DAY6					24	
DAY7					24	
DAY8						24

TREATMENT: 32% RP2		REPLICATE 2		SAMPLE A		
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	33					
DAY2			31			
DAY3				31		
DAY4					31	
DAY5					31	
DAY6					31	
DAY7						31

TREATMENT: 32% RP2		REPLICATE 2		SAMPLE B		
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	28					
DAY2		28				
DAY3		2		23		
DAY4						23
DAY5						23
DAY6						23
DAY7						23
DAY8						

TREATMENT: 100% RP2		REPLICATE 1		SAMPLE A		
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	22					
DAY2		22				
DAY3			21			
DAY4					21	
DAY5					21	
DAY6					21	
DAY7					21	
DAY8					21	
DAY9						21

TREATMENT: 100% RP2		REPLICATE 1		SAMPLE B		
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	31					
DAY2		20				
DAY3				20		
DAY4						
DAY5					20	
DAY6					20	
DAY7					20	
DAY8						20

TREATMENT:100% RP2		REPLICATE 2		SAMPLE A		
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	29					
DAY2		29				
DAY3			29			
DAY4					29	
DAY5					29	
DAY6					29	
DAY7					29	
DAY8					29	
DAY9						29

TREATMENT: 100% RP2		REPLICATE 2		SAMPLE B		
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	26					
DAY2		21				
DAY3				21		
DAY4					21	
DAY5					21	
DAY6					21	
DAY7					21	
DAY8					21	
DAY9					21	
DAY10					21	
DAY11						21

TRIAL #2 COMMENCED 28/2/91  
ENDPOINT DEVELOPMENTAL CHARACTERISTICS PAGE 1  
EGG NUMBERS RAW DATA  
SPECIES CYRAULUS SP.

TREATMENT: CONTROL	REPLICATE 1			SAMPLE B		
	GASTRULA	TROCH	DEVELOPMENTAL STAGE			
			VEL	HIPPO	H'LING	N'NATE
DAY1	3 6 6 4					
DAY2		3 6 6 4				
DAY3				3 6 6 4		
DAY4					3 6 6 4	
DAY5						3 6 6 4

TREATMENT: CONTROL		REPLICATE 2		SAMPLE 8	
		DEVELOPMENTAL STAGE			
	GASTRULA	TROCH	VEL	HIPPO	H'LING
					N'NATE
DAY1	5				
	8				
	6				
	4				
	5				
DAY2		4			
		8			
		5			
		4			
		5			
DAY3				4	
				8	
				5	
				4	
				5	
DAY4					4
					8
					5
					4
					4
					5
DAY5					
					4
					8
					5
					4

TREATMENT: 1% RP2	REPLICATE 1			SAMPLE B		
	GASTRULA	TROCH	DEVELOPMENTAL STAGE			
			VEL	HIPPO	H'LING	N'NATE
DAY1	2 5 1 5 4					
DAY2	1	2 5  5 4				
DAY3			1	2 5  5 4		
DAY4					2 5 1 5 4	
DAYS					2 5 1 5 4	
DAY6						2 5 1 5 4

TREATMENT: 1	RP2	REPLICATE 2	SAMPLE 8
	GASTRULA	DEVELOPMENTAL STAGE	
		VEL	H'LING
		HIPPO	N'NATE
DAY1	5 4 5 5 6 3 5 5 4		
DAY2		5 4 5 5 3 5 5 4	6
DAY3			5 1 5 5 6 3 4 1 5

DAY4

5  
4  
5  
5  
6  
3  
5  
5  
4

DAY5

5  
4  
5  
5  
6  
3  
5  
5  
4

DAY6

5  
4  
5  
5  
6  
3  
5  
5  
4

TREATMENT: 3.2% RP2      REPLICATE 1      SAMPLE A

	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	3 5 5 7					
DAY2		3 5 5 6				
DAY3			3 1	3 2 5 5		
DAY4					3 5 5 6	
DAY5						3 5 5 6

TREATMENT: 3.2% RP2      REPLICATE 1      SAMPLE B

	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	8 4 4 3 5					
DAY2		8 4 4 3 5				
DAY3			3 1 2 1	8 1 3 1 4		
DAY4					8 4 4 3 5	
DAY5						8 4 3
DAY6						4 5

TREATMENT: 3.2% RP2      REPLICATE 2      SAMPLE A

	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	6 4 6 7					
DAY2		6 4 6 7				
DAY3			1 2	5 4 6 5		
DAY4					5 4 6 7	
DAY5						5 4
DAY6						6 7

TREATMENT: 3.2% RP2      REPLICATE 2      SAMPLE B

	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	3 7 4 5 5 3					
DAY2		3	7 4			
DAY3				3 2	7 4 3 5 3	
DAY4						2 7 4 5 4 3

DAY5

2  
7  
4  
5  
4

TREATMENT: 10% RP2      REPLICATE 1      SAMPLE A

	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	6 3 4 6 3					
DAY2		3 4 1 3	6 5			
DAY3					6 3 4 6 3	
DAY4						6 3 4 6 3
DAY5						6 3 4 6 3
DAY6						6 3 4 6 3

TREATMENT: 10% RP2      REPLICATE 1      SAMPLE B

	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	9 7 7 6 8					
DAY2		7 7 5 6	9 1			
DAY3				9 7 6 6 8		
DAY4					9 7 7 5 8	
DAY5					9 7	
Day6						7 5 8 9 7



TREATMENT: 10% RP2		REPLICATE 2		SAMPLE A	
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		N'NATE
			VEL	HIPPO	
DAY1	3 3 7 6 3				
DAY2		3 3 7 6 3			
DAY3				3 3 7 6 3	
DAY4					3 3 7 6 3
DAY5					3 3 7 6 3
DAY6					3 3 6 6 3

TREATMENT: 10% RP2		REPLICATE 2		SAMPLE B	
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		N'NATE
			VEL	HIPPO	
DAY1	6 6 4 6 8				
DAY2		6 6		4 6 4	
DAY3		1		5 6 4 6 8	
DAY4					5 6 4 6 8
DAY5					5 6 4 6 8
DAY6					5 6

TREATMENT: 32% RP2		REPLICATE 1		SAMPLE A	
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		N'NATE
			VEL	HIPPO	
DAY1	7 4 1 2 4 4				
DAY2		7 4 1 2		4 4	
DAY3				7 4 1 2 3 4	
DAY4					7 4 1 2 4 4 4
DAY5					7 4 1 2 4 4
DAY6					7 4 1 2 4 4

TREATMENT: 32% RP2		REPLICATE 1		SAMPLE B	
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		N'NATE
			VEL	HIPPO	
DAY1	4 5 5 5				
DAY2		5 4			
DAY3				5 4	
DAY4					5 4
DAY5					5 4
DAY6					5

TRIAL #2  
ENDPOINT  
SPECIES

COMMENCED 28/2/91  
DEVELOPMENTAL CHARACTERISTICS  
EGG NUMBERS  
CYTULUS SP.

PAGE 19  
RAW DATA

TREATMENT: 32% RP2		REPLICATE 2		SAMPLE A	
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		N'NATE
			VEL	HIPPO	
DAY1	6 5 5 4				
DAY2		6 5 5			
DAY3		5 4			
DAY4				6 5 5 4	
DAY5					6 5 5 4
DAY6					5 5 5 4

TREATMENT: 32% RP2		REPLICATE 2		SAMPLE B	
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		N'NATE
			VEL	HIPPO	
DAY1	6 5 2 3 4 3				
DAY2			6 5 2 3 4 3		
DAY3				6 5 2 3 4 3	
DAY4					6 5 1 3 4 2
DAY5				1	6 5 1 3 4 3
DAY6					5 1 3 4 3

	GASTRULA	TROCH	REPLICATE 1			
			SAMPLE A			
			DEVELOPMENTAL STAGE			
			VEL	HIPPO	H'LING	N'NATE
DAY1	7 8 5					
DAY2		7 8 5				
DAY3				7 8 5		
DAY4					7 8 5	
DAY5					7 8 5	
DAY6					7 8 5	
DAY7					7 8 5	
DAY8					7 8 4	
DAY9					7 8 1	
DAY10						7 8

DAY6	6 6 3 6 4
DAY7	6 6 3 6 4
DAY8	6 6 3 6 4
DAY9	6 3
DAY10	1 3 3 1
DAY11	

	GASTRULA	TROCH	REPLICATE 1			
			SAMPLE B			
			DEVELOPMENTAL STAGE			
			VEL	HIPPO	H'LING	N'NATE
DAY1	3 5 4 2 2					
DAY2	5 2 2	3 4				
DAY3				3 5 4 2 2		
DAY4					3 5 4	
DAY5					2 2	
DAY6					3 5 4 2 2	
DAY7					3 5 4 2 2	
DAY8					3 4 4 2 2	
DAY9					1	4 2
DAY 10						1 2 2

	GASTRULA	TROCH	REPLICATE 2			
			SAMPLE B			
			DEVELOPMENTAL STAGE			
			VEL	HIPPO	H'LING	N'NATE
DAY1	7 6 3 4 8					
DAY2		7 6 3 4 8				
DAY3				1	6 6 3 1	
DAY4						7 6 3 4 8
DAY5						7 6 3 4 8
DAY6						7 6 3 4 8
DAY7						7 6 3 4 8
DAY8						6 3 4 8 3

	GASTRULA	TROCH	REPLICATE 2			
			SAMPLE A			
			DEVELOPMENTAL STAGE			
			VEL	HIPPO	H'LING	N'NATE
DAY1	6 6 3 6 4					
DAY2				6 6 3 6		
DAY3					6 6 3 6	
DAY4					6 6 3 6 4	
DAY5					6 6 3 6 4	

## Appendix 3.a.3.

TRIAL #2 COMMENCED 28/2/91  
 ENDPOINT DEVELOPMENTAL CHARACTERISTICS PAGE 1  
 SPECIES EGG NUMBERS RAW DATA  
 LYMNEAE SP.

TREATMENT: CONTROL REPLICATE 1 SAMPLE A  
 DEVELOPMENTAL STAGE

	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	52					
DAY2		23	27			
DAY3		7		45		
DAY4					45	
DAY5					45	
DAY6						45

TREATMENT: CONTROL REPLICATE 1 SAMPLE B  
 DEVELOPMENTAL STAGE

	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	88					
DAY2		40	48			
DAY3		6		82		
DAY4		5		13	70	
DAY5					81	
DAY6					81	
DAY7						81

TREATMENT: CONTROL REPLICATE 2 SAMPLE A  
 DEVELOPMENTAL STAGE

	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	28					
DAY2		28				
DAY3			14	14		
DAY4					28	
DAY5					28	
DAY6					28	
DAY7						28

TREATMENT: CONTROL REPLICATE 2 SAMPLE B  
 DEVELOPMENTAL STAGE

	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	85					
DAY2		85				
DAY3				85		
DAY4					82	
DAY5					82	
DAY6					82	
DAY7						82

TREATMENT: 1% RP2 REPLICATE 1 SAMPLE A  
 DEVELOPMENTAL STAGE

	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	55					
DAY2			54			
DAY3				54		
DAY4					54	
DAY5					54	
DAY6					54	
DAY7						54

TREATMENT: 1% RP2 REPLICATE 1 SAMPLE B  
 DEVELOPMENTAL STAGE

	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	58					
DAY2		20	37			
DAY3		10		47		
DAY4		2		5	50	
DAY5				7	50	
DAY6				7	50	
DAY7					3	50
DAY8						3

TREATMENT: 1% RP2 REPLICATE 2 SAMPLE A  
 DEVELOPMENTAL STAGE

	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	44					
DAY2		22	22			
DAY3			8	36		
DAY4					36	
DAY5					36	
DAY6					36	
DAY7					3	33
DAY8						3

TREATMENT: 1% RP2 REPLICATE 2 SAMPLE B  
 DEVELOPMENTAL STAGE

	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	76					
DAY2		38	38			
DAY3		16		60		
DAY4					70	
DAY5					70	
DAY6					70	
DAY7					70	
DAY8						70

TREATMENT: 3.2% RP2 REPLICATE 1 SAMPLE A  
 DEVELOPMENTAL STAGE

	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	87					
DAY2		57				
DAY3		11	46			
DAY4			12		45	
DAY5				12	45	
DAY6				12	45	
DAY7				12		45
DAY8				0		

TREATMENT: 3.2% RP2 REPLICATE 1 SAMPLE B  
 DEVELOPMENTAL STAGE

	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	54					
DAY2		52				
DAY3				52		
DAY4					52	
DAY5					52	
DAY6					52	
DAY7					33	19
DAY8						33

TREATMENT: 3.2% RP2 REPLICATE 2 SAMPLE A  
 DEVELOPMENTAL STAGE

	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	43					
DAY2		43				
DAY3			1	42		
DAY4				43		
DAY5				43		
DAY6				43		
DAY7					43	

TREATMENT: 3.2% RP2 REPLICATE 2 SAMPLE B  
 DEVELOPMENTAL STAGE

	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	20					
DAY2			10	10		
DAY3			3	17		
DAY4					17	
DAY5					17	
DAY6					17	
DAY7						17

TREATMENT: 10% RP2		REPLICATE 1		SAMPLE A	
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		N'NATE
			VEL	HIPPO	
DAY1	46				
DAY2		26	20		
DAY3			6	40	
DAY4			3	4	39
DAY5			3	4	39
DAY6					39
DAY7					39

TREATMENT: 10% RP2		REPLICATE 1		SAMPLE B	
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		N'NATE
			VEL	HIPPO	
DAY1	43 23				
DAY2		43 23			
DAY3			5 5	38 18	
DAY4			5	8	35 18
DAY5			5	8	35 18
DAY6					37 18
DAY7					37 18
DAY8					37 18

TREATMENT: 10% RP2		REPLICATE 2		SAMPLE A	
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		N'NATE
			VEL	HIPPO	
DAY1	70				
DAY2		70			
DAY3			70		
DAY4				70	
DAY5					70
DAY6					70
DAY7					2 68
DAY8					2

TREATMENT: 10% RP2		REPLICATE 2		SAMPLE B	
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		N'NATE
			VEL	HIPPO	
DAY1	42				
DAY2		20	22		
DAY3			6	36	
DAY4				8	34
DAY5					34
DAY6					28
DAY7					28

TREATMENT: 32% RP2		REPLICATE 1		SAMPLE A	
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		N'NATE
			VEL	HIPPO	
DAY1	81				
DAY2		81			
DAY3		2		79	
DAY4					76
DAY5					76
DAY6					76
DAY7					26 50
DAY8					26

TREATMENT: 32% RP2		REPLICATE 1		SAMPLE B	
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		N'NATE
			VEL	HIPPO	
DAY1	35				
DAY2		32			
DAY3			1	31	
DAY4				2	30
DAY5				1	30
DAY6					30
DAY7					6 24
DAY8					6

TREATMENT: 32% RP2		REPLICATE 2		SAMPLE A	
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		N'NATE
			VEL	HIPPO	
DAY1	89				
DAY2		46	32		
DAY3				32	
DAY4					32
DAY5					32
DAY6					32
DAY7					32

TREATMENT: 32% RP2		REPLICATE 2		SAMPLE B	
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		N'NATE
			VEL	HIPPO	
DAY1	36				
DAY2		36			
DAY3			5	31	
DAY4					31
DAY5					31
DAY6					31
DAY7					31
DAY8					31

TREATMENT: 100% RP2		REPLICATE 1		SAMPLE A	
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		N'NATE
			VEL	HIPPO	
DAY1	45				
DAY2		45			
DAY3			20	25	
DAY4				1	44
DAY5					45
DAY6					45
DAY7					45
DAY8					45
DAY9					45
DAY10					45
DAY12					35

TERMINATION

TREATMENT: 100% RP2		REPLICATE 1		SAMPLE B	
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		N'NATE
			VEL	HIPPO	
DAY1	69				
DAY2		69			
DAY3			2	67	
DAY4			1	1	67
DAY5					65
DAY6					65
DAY7					65
DAY8					65
DAY9					65
DAY10					65
DAY12					39 6

TERMINATION

TREATMENT: 100% RP2		REPLICATE 2		SAMPLE A	
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		N'NATE
			VEL	HIPPO	
DAY1	68				
DAY2		38	30		
DAY3		17		51	
DAY4		17			51
DAY5			1		51
DAY6					51
DAY7					51
DAY8					51
DAY9					51
DAY10					51
DAY12					51

TERMINATION

TREATMENT: 100% RP2		REPLICATE 2		SAMPLE B	
GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
		VEL	HIPPO		
DAY1	50				
DAY2		30	20		
DAY3		6		44	
DAY4			6	44	
DAY5				43	
DAY6				42	
DAY7				42	
DAY8				42	
DAY9				42	
DAY10				42	
DAY12				32	
TERMINATION					

TREATMENT		32% RP2 #1		32% RP2 #2	
EGGMASS		A	B	A	B
		5	5	6	5
A		5	6	6	6
B		5	6	6	6
C		5	6	6	6
D		5	6	6	6
E		6	6	5	5
F					
MEAN		5.2	5.8	5.67	5.5
STD		0.45	0.45	0.52	0.71

TREATMENT		100% RP2 #1		100% RP2 #2	
EGGMASS		A	B	A	B
		10	9	10	7
A		9	9	10	8
B		10	9	10	8
C			10	10	8
D			8	10	7
E					7
F					
MEAN		9.67	9	10	7.5
STD		0.58	0.71	0	0.55

### Appendix 3.b.

TRIAL 2 DATE 1/3/91  
 ENDPOINT: DEVELOPMENTAL RETARDATION DAYS TO EGG CAPSULE RUPTURE  
 SPECIES A. cummingii

TREATMENT	REPLICATE 1		REPLICATE 2	
	A	B	A	B
CONTROL	7	7	8	8
1% RP2	7	7	8	8
3.2% RP2	7	8	8	7
10% RP2	8	9	7	8
32% RP2	8	8	7	8
100% RP2	9	8	9	10

TRIAL 2 DATE 1/3/91  
 ENDPOINT: DEVELOPMENTAL RETARDATION DAYS TO EGG CAPSULE RUPTURE  
 SPECIES LYMNEAE SP.

TREATMENT	REPLICATE 1		REPLICATE 2	
	A	B	A	B
CONTROL	7	7	8	8
1% RP2	7	7	8	8
3.2% RP2	7	7	7	7
10% RP2	7	8	7	7
32% RP2	7	7	7	8
100% RP2	14	13	13	13

TRIAL 2 DATE 1/3/91  
 DEVELOPMENTAL RETARDATION DAYS TO EGG CAPSULE RUPTURE  
 SPECIES GYRAULUS SP.

TREATMENT	CONT. #1		CONT. #2	
	A	B	A	B
EGGMASS				
A	5	6	5	5
B	5	6	5	5
C	5	6	5	6
D	5	6	5	6
E	5	6	5	6
MEAN	5	6	5	5.6
STD	0	0	0	0.55

TREATMENT	1% RP2 #1		1% RP2 #2	
	A	B	A	B
EGGMASS				
A	6	5	6	5
B	6	5	6	6
C	6	5	6	6
D	6	6		6
E	5	6		
F		6		
G		6		
H		6		
I		6		
MEAN	5.8	5.67	6	5.75
STD	0.45	0.5	0	0.5

TREATMENT	3.2% RP2 #1		3.2% RP2 #2	
	A	B	A	B
EGGMASS				
A	5	5	5	5
B	5	5	5	5
C	5	5	5	5
D	5	6	6	5
E		6	6	5
F				5
G				5
MEAN	5	5.4	5.4	5
STD	0	0.55	0.5	0

TREATMENT	10% RP2 #1		10% RP2 #2	
	A	B	A	B
EGGMASS				
A	5	5	5	5
B	5	5	5	5
C	5	5	6	5
D	6	6	6	6
E	6	6	6	6
MEAN	5.4	5.6	5.6	5.4
STD	0.55	0.55	0.55	0.55

# Appendix 3.c.

TRIAL#2 COMMENCED 28/2/91  
PHYSICO-CHEMICAL DATA

	CONDUCTIVITY uSem/cm		TREATMENT									
	CON A	CON B	1% A	1% B	3.2% A	3.2% B	10% A	10% B	32% A	32% B	100% A	100% B
DAY1	25.9	27.7	42.2	42.2	71.1	70.7	159	160	409	413	1080	1080
DAY2	26.2	29.5	47.7	45.2	73.2	72.6	170	171	427	433	1100	1100
DAY3	27	38.7	42.7	43	74.7	77.9	173	173	437	438	1100	1100
DAY4	25.1	30	40.7	42.3	73.3	75.9	172	172	433	429	1090	1100
DAY5	26.1	34.3	42.6	42.8	76.2	77.4	177	176	445	442	1120	1120
DAY6	26.2	34.2	42.6	43.1	75.7	77.6	178	185	449	445	1120	1120
DAY7	29.2	29.8	42.2	44.2	77.5	77.5	178	185	448	448	1130	1120
DAY8	27.5	25.8	41.04	41.6	75.9	76.2	175	179	447	444	1130	1130
DAY9	27.3	31.4	42.7	44.3	77.5	80.6	185	178	454	448	1150	1120
DAY10	28.3	34.3	43.8	45.6	80.6	82.1	189	188	463	471	1190	1150
DAY11	29.5	32.8	46.7	42	79.1	77.3	179	180	454	452	1130	1130
DAY12	27	30.2	42.3	42.9	76.2	75.6	177	178	450	451	1140	1130

pH

			TREATMENT									
	CON A	CON B	1% A	1% B	3.2% A	3.2% B	10% A	10% B	32% A	32% B	100% A	100% B
DAY1	6.36	6.55	6.59	6.6	6.61	6.6	6.74	6.72	6.75	6.96	7.19	7.31
DAY2	6.42	6.68	6.61	6.64	6.86	6.56	6.97	6.86	6.71	6.97	7.26	7.21
DAY3	6.6	7.63	6.98	6.89	6.85	6.97	7	7	7.08	7.2	7.28	7.3
DAY4	6.34	6.78	6.75	6.93	6.81	6.85	7.01	7.26	7.13	7.15	7.15	7.4
DAY5	6.32	6.73	6.73	6.71	6.6	6.7	6.76	6.86	6.84	6.89	7.09	7.26
DAY6	6.17	6.43	6.77	6.75	6.71	6.85	6.86	7.11	7.16	6.66	6.84	7.17
DAY7	6.68	6.88	6.66	6.8	6.7	6.73	6.91	6.94	6.93	6.92	7.05	7.04
DAY8	6.5	6.58	6.73	6.78	6.78	6.87	6.84	7.01	6.95	6.98	7.2	7.24
DAY9	6.36	6.85	6.77	6.77	6.72	6.73	7.15	7.08	7.05	6.97	7.23	7.29
DAY10	6.24	6.53	6.4	6.56	6.63	6.73	6.87	6.91	6.95	6.92	7.12	7.08
DAY11	6.69	6.66	6.73	6.81	6.83	6.8	6.94	6.94	7.07	7.07	7.21	7.33
DAY12	6.49	6.66	6.56	6.73	6.71	6.78	6.82	6.97	7.02	7.07	7.19	7.42

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	TREATMENT											
	CON A	CON B	1% A	1% B	3.2% A	3.2% B	10% A	10% B	32% A	32% B	100% A	100% B
DAY1	7.2	7.3	7.4	7.6	7.5	7.5	7.6	7.5	7.6	7.6	7.5	7.6
DAY2	7.7	7.3	7.4	7.5	7.6	7.6	7.6	7.2	7.3	7.5	7.3	7.1
DAY3	7.5	7.3	7.7	7.4	7.5	7.4	7.4	7.4	7.4	7.5	7.2	7.3
DAY4	7.6	7.5	7.4	7.6	7.6	7.6	7.3	7.4	7.5	7.4	7.4	7.6
DAY5	7.2	7.5	7.6	7.3	7.4	7.6	7.3	7.3	7.4	7.2	7.2	7.5
DAY6	7.2	7.3	7.2	7.5	7.6	7.4	7.5	7.5	7.5	6.9	7.1	7.4
DAY7	7.6	7.3	7.5	7.6	7.5	7.5	7.7	7.5	7.5	7.7	7.5	7.5
DAY8	7.1	7.2	7.6	7.4	7.5	7.5	7.4	7.5	7.5	7.1	7.5	7.3
DAY9	7.3	7.3	7.5	7.2	7.4	7.3	7.3	7.1	7.4	7.3	7.3	7.2
DAY10	7.6	7.4	7.3	7.5	7.4	7.3	7.3	7.5	7.3	7.4	7.3	7.5
DAY11	7.1	7.2	7.4	7.3	7.3	7.1	7.4	7.1	7.3	7.4	7.2	7.1
DAY12	7.7	7.6	7.5	7.6	7.3	7.5	7.5	7.5	7.6	7.4	7.1	7.4

## Appendix 4.a.1.

TRIAL #3 COMMENCED 13/3/91  
 ENDPOINT DEVELOPMENTAL CHARACTERISTICS PAGE 1  
 EGG NUMBERS  
 SPECIES A. carinata RAW DATA

TREATMENT : CONTROL  
 REPLICATE 1 SAMPLE A

	GASTRULA	DEVELOPMENTAL STAGE		
		HIPPO	H'LING	N'NATE
DAY1	13 11			
DAY4			13 11	
DAY5			13 11	
DAY6			13 11	
DAY7				13 11

TREATMENT : CONTROL  
 REPLICATE 1 SAMPLE B

	GASTRULA	DEVELOPMENTAL STAGE		
		HIPPO	H'LING	N'NATE
DAY1	8 7 12			
DAY4		8 7 12		
DAY5		8 7 12		
DAY6		8 7 12		
DAY7				8 7 12

TREATMENT : CONTROL  
 REPLICATE 1 SAMPLE A

	GASTRULA	DEVELOPMENTAL STAGE		
		HIPPO	H'LING	N'NATE
DAY1	9 8 9			
DAY4			9 8 9	
DAY5			9 8 9	
DAY6			9 8 9	
DAY7			9 8 9	
DAY8				9 8 9

TREATMENT : CONTROL  
 REPLICATE 2 SAMPLE B

	GASTRULA	DEVELOPMENTAL STAGE		
		HIPPO	H'LING	N'NATE
DAY1	15 9			
DAY4			15 9	
DAY5			15 9	
DAY6			15 9	
DAY7				15 9

TREATMENT : 1% RP2  
 REPLICATE 1 SAMPLE A

	GASTRULA	DEVELOPMENTAL STAGE		
		HIPPO	H'LING	N'NATE
DAY1	30			
DAY4			29	
DAY5			29	
DAY6				29
DAY7				
DAY8				

TREATMENT : 1% RP2  
 REPLICATE 1 SAMPLE B

	GASTRULA	DEVELOPMENTAL STAGE		
		HIPPO	H'LING	N'NATE
DAY1	20			
DAY4				19
DAY5				19
DAY6				19
DAY7				19
DAY8				19

TREATMENT : 1% RP2  
 REPLICATE 2 SAMPLE A

	GASTRULA	DEVELOPMENTAL STAGE		
		HIPPO	H'LING	N'NATE
DAY1	22			
DAY4				22
DAY5				22
DAY6				22
DAY7				22
DAY8				22

TREATMENT : 1% RP2  
 REPLICATE 2 SAMPLE B

	GASTRULA	DEVELOPMENTAL STAGE		
		HIPPO	H'LING	N'NATE
DAY1	27			
DAY4				26
DAY5				24
DAY6				22
DAY7				22
DAY8				22

TREATMENT : 3.2% RP2  
 REPLICATE 1 SAMPLE A

	GASTRULA	DEVELOPMENTAL STAGE		
		HIPPO	H'LING	N'NATE
DAY1	9 14 9			
DAY4			9 14 9	
DAY5			9 14 9	
DAY6			9 14 9	
DAY7				9 14 9

TREATMENT : 3.2% RP2  
 REPLICATE 1 SAMPLE B

	GASTRULA	DEVELOPMENTAL STAGE		
		HIPPO	H'LING	N'NATE
DAY1	8 7 5			
DAY4			8 7 5	
DAY5			8 7 5	
DAY6			8 7 5	
DAY7			8 7 5	
DAY8				8 7 5

TREATMENT : 3.2% RP2  
REPLICATE 2

SAMPLE A

	GASTRULA	HIPPO	H'LING	N'NATE
DAY1	9 8 11			
DAY4			9 8 11	
DAY5			9 8 11	
DAY6			9 8 11	
DAY7			9 8 11	
DAY8				9 8 11

TREATMENT : 3.2% RP2  
REPLICATE 2

SAMPLE B

	GASTRULA	HIPPO	H'LING	N'NATE
DAY1	12 12			
DAY4			12 12	
DAY5			12 12	
DAY6			12 12	
DAY7			12 12	
DAY8				12 12

TREATMENT : 10% RP2  
REPLICATE 1

SAMPLE A

	GASTRULA	HIPPO	H'LING	N'NATE
DAY1	6 9 6			
DAY4			6 9 6	
DAY5			6 9 6	
DAY6			6 9 6	
DAY7			6 9 6	
DAY8				6 9 6

TREATMENT : 10% RP2  
REPLICATE 1

SAMPLE B

	GASTRULA	HIPPO	H'LING	N'NATE
DAY1	13 16			
DAY4			13 16	
DAY5			13 16	
DAY6			13 16	
DAY7				13 16

TREATMENT : 10% RP2  
REPLICATE 2

SAMPLE A

	GASTRULA	HIPPO	H'LING	N'NATE
DAY1	9 11 9			
DAY4			9 11 9	
DAY5			9 11 9	
DAY6			9 11 9	
DAY7				9 11 9

TREATMENT : 10% RP2  
REPLICATE 2

SAMPLE B

	GASTRULA	HIPPO	H'LING	N'NATE
DAY1	24			
DAY4			22	
DAY5			22	
DAY6			22	
DAY7			22	
DAY8			22	
DAY9				22

TREATMENT : 32% RP2  
REPLICATE 1

SAMPLE A

	GASTRULA	HIPPO	H'LING	N'NATE
DAY1	29			
DAY4			29	
DAY5			29	
DAY6			29	
DAY7			29	
DAY8				29

TREATMENT : 10% RP2  
REPLICATE 1

SAMPLE B

	GASTRULA	HIPPO	H'LING	N'NATE
DAY1	11 13			
DAY4			11 13	
DAY5			11 12	
DAY6			11 2	
DAY7			11 2	
DAY8				11 2

TREATMENT : 32% RP2  
REPLICATE 2

SAMPLE A

	GASTRULA	HIPPO	H'LING	N'NATE
DAY1	14 12			
DAY4			14 12	
DAY5			14 12	
DAY6			14 12	
DAY7				14 12

TREATMENT : 32% RP2  
REPLICATE 2

SAMPLE B

	GASTRULA	HIPPO	H'LING	N'NATE
DAY1	11 13			
DAY4			11 13	
DAY5			11 13	
DAY6			11 13	
DAY7				11 13

TREATMENT : 100% RP2  
REPLICATE 1

SAMPLE A

	GASTRULA	HIPPO	H'LING	N'NATE
DAY1	9 12 11			
DAY4			9 12 11	
DAY5			9 12 11	
DAY6			9 12 11	



## Appendix 4.a.2.

DAY7	9	12	11
DAY8	9	12	11
DAY9	8	12	11
DAY10		8	12
		11	

TREATMENT :100% RP2  
REPLICATE 1

SAMPLE B

	GASTRULA	HIPPO	H'LING	N'NATE
DAY1	9	11		
DAY4		9	10	
DAY5		9	10	
DAY6		9	10	
DAY7		9	10	
DAY8		9	10	
DAY9		9	10	
DAY10			9	10

TREATMENT :100% RP2  
REPLICATE 2

SAMPLE A

	GASTRULA	HIPPO	H'LING	N'NATE
DAY1	9	9	12	
DAY4		7	9	12
DAY5		7	9	12
DAY6		7	9	12
DAY7		7	9	12
DAY8		7	9	10
DAY9				7

REPLICATE 2

SAMPLE

	GASTRULA	HIPPO	H'LING	N'NATE
DAY1	9	10	5	
DAY4		9	10	5
DAY5		9	10	5
DAY6		9	10	5
DAY7		9	10	5
DAY8		9	10	5
DAY9		9	10	5
DAY10			9	10
			5	

TRIAL #3

ENDPOINT

SPECIES

COMMENCED 13/3/91

DEVELOPMENTAL CHARACTERISTICS PAGE 1

EGG NUMBERS

A. cummingii

RAW DATA

TREATMENT :CONTROL  
REPLICATE 1

SAMPLE A

	GASTRULA	HIPPO	H'LING	N'NATE
DAY1	28			
DAY4		27		
DAY5			27	
DAY6			27	
DAY7			27	
DAY8				27

SAMPLE B

	GASTRULA	HIPPO	H'LING	N'NATE
DAY1	21			
DAY4		21		
DAY5			19	
DAY6			19	
DAY7			19	
DAY8				19

TREATMENT :CONTROL  
REPLICATE 2

SAMPLE A

	GASTRULA	HIPPO	H'LING	N'NATE
DAY1	28			
DAY4			28	
DAY5			28	
DAY6			28	
DAY7			28	
DAY8				28

SAMPLE B

	GASTRULA	HIPPO	H'LING	N'NATE
DAY1	21			
DAY4			21	
DAY5			21	
DAY6			21	
DAY7			21	
DAY8				21

TREATMENT :1% RP2  
REPLICATE 1

SAMPLE A

	GASTRULA	HIPPO	H'LING	N'NATE
DAY1	30			
DAY4			29	
DAY5			29	
DAY6			29	
DAY7				29
DAY8				

SAMPLE B

	GASTRULA	HIPPO	H'LING	N'NATE
DAY1	20			
DAY4			19	
DAY5			19	
DAY6			19	
DAY7			19	
DAY8				19

TREATMENT :1% RP2  
REPLICATE 2

SAMPLE A

	GASTRULA	DEVELOPMENTAL STAGE HIPPO H'LING N'NATE
DAY1	22	
DAY4		22
DAY5		22
DAY6		22
DAY7		22
DAY8		22

SAMPLE B

	GASTRULA	DEVELOPMENTAL STAGE HIPPO H'LING N'NATE
DAY1	27	
DAY4		22
DAY5		22
DAY6		22
DAY7		22
DAY8		22

TREATMENT :3.2% RP2  
REPLICATE 1

SAMPLE A

	GASTRULA	DEVELOPMENTAL STAGE HIPPO H'LING N'NATE
DAY1	29	
DAY4		27
DAY5		26
DAY6		25
DAY7		25

SAMPLE B

	GASTRULA	DEVELOPMENTAL STAGE HIPPO H'LING N'NATE
DAY1	20	
DAY4		19
DAY5		19
DAY6		19
DAY7		19
DAY8		19

TREATMENT :3.2% RP2  
REPLICATE 2

SAMPLE A

	GASTRULA	DEVELOPMENTAL STAGE HIPPO H'LING N'NATE
DAY1	23	
DAY4		22
DAY5		22
DAY6		22

SAMPLE B

	GASTRULA	DEVELOPMENTAL STAGE HIPPO H'LING N'NATE
DAY1	27	
DAY4		24
DAY5		24
DAY6		24
DAY7		24

TREATMENT :10% RP2  
REPLICATE 1

SAMPLE A

	GASTRULA	DEVELOPMENTAL STAGE HIPPO H'LING N'NATE
DAY1	31	
DAY4		31
DAY5		31
DAY6		31

SAMPLE B

	GASTRULA	DEVELOPMENTAL STAGE HIPPO H'LING N'NATE
DAY1	20	
DAY4		20
DAY5		20
DAY6		20
DAY7		20
DAY8		20

TREATMENT :10% RP2  
REPLICATE 2

SAMPLE A

	GASTRULA	DEVELOPMENTAL STAGE HIPPO H'LING N'NATE
DAY1	25	
DAY4		25
DAY5		25
DAY6		25
DAY7		25

SAMPLE B

	GASTRULA	DEVELOPMENTAL STAGE HIPPO H'LING N'NATE
DAY1	25	
DAY4		25
DAY5		25
DAY6		24
DAY7		24

TREATMENT :32% RP2  
REPLICATE 1

SAMPLE A

	GASTRULA	DEVELOPMENTAL STAGE HIPPO H'LING N'NATE
DAY1	20	
DAY4		19
DAY5		19
DAY6		19
DAY7		19
DAY8		19

SAMPLE B

	GASTRULA	DEVELOPMENTAL STAGE HIPPO H'LING N'NATE
DAY1	31	
DAY4		30
DAY5		30
DAY6		30
DAY7		30
DAY8		30

TREATMENT :32% RP2  
REPLICATE 2

SAMPLE A

	GASTRULA	DEVELOPMENTAL STAGE HIPPO H'LING N'NATE
DAY1	24	
DAY4		21
DAY5		21
DAY6		21
DAY7		21
DAY8		21

SAMPLE B

	GASTRULA	DEVELOPMENTAL STAGE HIPPO H'LING N'NATE
DAY1	23	
DAY4		23
DAY5		23
DAY6		23
DAY7		23
DAY8		23

## Appendix 4.a.3.

TREATMENT :100% RP2  
REPLICATE 1

SAMPLE A

	DEVELOPMENTAL STAGE			
	GASTRULA	HIPPO	H'LING	N'NATE
DAY1	29			
DAY4			29	
DAY5			28	
DAY6			28	
DAY7			28	
DAY8			28	
DAY9				28

SAMPLE B

	DEVELOPMENTAL STAGE			
	GASTRULA	HIPPO	H'LING	N'NATE
DAY1	20			
DAY4			19	
DAY5			19	
DAY6			19	
DAY7			19	
DAY8			19	
DAY9				19

TREATMENT :100% RP2  
REPLICATE 2

SAMPLE A

	DEVELOPMENTAL STAGE			
	GASTRULA	HIPPO	H'LING	N'NATE
DAY1	27			
DAY4			27	
DAY5			27	
DAY6			27	
DAY7			27	
DAY8				27

SAMPLE B

	DEVELOPMENTAL STAGE			
	GASTRULA	HIPPO	H'LING	N'NATE
DAY1	24			
DAY4			24	
DAY5			24	
DAY6			20	
DAY7				20

TRIAL #3  
ENDPOINT

SPECIES

COMMENCED 13/1/91  
DEVELOPMENTAL CHARACTERISTICS PAGE 1  
EGG NUMBERS  
GLYPTOPHYSA SP. RAW DATATREATMENT :CONTROL  
REPLICATE 1

SAMPLE A

	DEVELOPMENTAL STAGE			
	GASTRULA	HIPPO	H'LING	N'NATE
DAY1	6 6 9 10			
DAY4			6 6 9 9	
DAY5			6 6 9 9	
DAY6			6 6 9 9	
DAY7				6 6 9 9

TREATMENT :CONTROL  
REPLICATE 1

SAMPLE B

	DEVELOPMENTAL STAGE			
	GASTRULA	HIPPO	H'LING	N'NATE
DAY1	9 10 12			
DAY4			9 10 11	
DAY5			9 10 11	
DAY6			9 9 11	
DAY7			9 9 10	
DAY8			9 9 10	
DAY9				9 9 10

TREATMENT :CONTROL  
REPLICATE 2

SAMPLE A

	DEVELOPMENTAL STAGE			
	GASTRULA	HIPPO	H'LING	N'NATE
DAY1	9 10 10			
DAY4			6 10 10	
DAY5			6 7 7	
DAY6				6 7 7

TREATMENT :CONTROL  
REPLICATE 2

SAMPLE B

	DEVELOPMENTAL STAGE			
	GASTRULA	HIPPO	H'LING	N'NATE
DAY1	7 7			
DAY4			7 7	
DAY5				7 7
DAY6				6 7
DAY7				6 7
DAY8				6 7

TREATMENT :1% RP2  
REPLICATE 2

	SAMPLE A			
	GASTRULA	DEVELOPMENTAL STAGE HIPPO	H'LING	N'NATE
DAY1	10 12			
DAY4			10 8	
DAY5			10 8	
DAY6				10 8

TREATMENT :1% RP2  
REPLICATE 2

	SAMPLE B			
	GASTRULA	DEVELOPMENTAL STAGE HIPPO	H'LING	N'NATE
DAY1	11 2			
DAY4			2 2	
DAY5			2 2	
DAY6			2 2	
DAY7			2 2	
DAY8				2 2

TREATMENT :3.2% RP2  
REPLICATE 1

	SAMPLE A			
	GASTRULA	DEVELOPMENTAL STAGE HIPPO	H'LING	N'NATE
DAY1	11 2			
DAY4			9 2	
DAY5			9 2	
DAY6			9 2	
DAY7			9 2	
DAY8				9 2

TREATMENT :3.2% RP2  
REPLICATE 1

	SAMPLE B			
	GASTRULA	DEVELOPMENTAL STAGE HIPPO	H'LING	N'NATE
DAY1	10 11			
DAY4			9 9	
DAY5			9 9	
DAY6				9 9

TREATMENT :1% RP2  
REPLICATE 1

	SAMPLE A			
	GASTRULA	DEVELOPMENTAL STAGE HIPPO	H'LING	N'NATE
DAY1	12			
DAY4		12		
DAY5			5	
DAY6			5	
DAY7			5	
DAY8			5	
DAY9				5

TREATMENT :1% RP2  
REPLICATE 1

	SAMPLE B			
	GASTRULA	DEVELOPMENTAL STAGE HIPPO	H'LING	N'NATE
DAY1	14 15			
DAY4			12 15	
DAY5			12 15	
DAY6			12 15	
DAY7				12 15

TREATMENT :3.2% RP2  
REPLICATE 2

	SAMPLE A			
	GASTRULA	DEVELOPMENTAL STAGE HIPPO	H'LING	N'NATE
DAY1	11 12 8			
DAY4			9 7 4	
DAY5			4 7 3	
DAY6				4 7 3

TREATMENT :3.2% RP2  
REPLICATE 2

	SAMPLE B			
	GASTRULA	DEVELOPMENTAL STAGE HIPPO	H'LING	N'NATE
DAY1	10			
DAY4		9		
DAY5			6	
DAY6			6	
DAY7			5	
DAY8			5	
DAY9				5

TREATMENT :10% RP2  
REPLICATE 1

	SAMPLE A			
	GASTRULA	DEVELOPMENTAL STAGE HIPPO	H'LING	N'NATE
DAY1	10 12 8			
DAY4			2 4 4	
DAY5			2 4 4	
DAY6			2 4 4	
DAY7				2 4 4

TREATMENT :10% RP2  
REPLICATE 1

	SAMPLE B			
	GASTRULA	DEVELOPMENTAL STAGE HIPPO	H'LING	N'NATE
DAY1	8			
DAY4		7		
DAY5			4	
DAY6			4	
DAY7			4	
DAY8			4	
DAY9				4

TREATMENT :10% RP2  
REPLICATE 2

	SAMPLE A			
	GASTRULA	DEVELOPMENTAL STAGE HIPPO	H'LING	N'NATE
DAY1	12 12			
DAY4			12 12	
DAY5			12 12	
DAY6				12 12

TREATMENT :10% RP2  
REPLICATE 2

SAMPLE B

	GASTRULA	DEVELOPMENTAL STAGE HIPPO H'LING N'NATE
DAY1	11 11	
DAY4		11 11
DAY5		11 11
DAY6		11 11
DAY7		11 11
DAY8		11 11
DAY9		11 11

TREATMENT :32% RP2  
REPLICATE 1

SAMPLE A

	GASTRULA	DEVELOPMENTAL STAGE HIPPO H'LING N'NATE
DAY1	7 11 11 13 14 12	
DAY4		7 10 11 13 7 4
DAY5		6 2 9 4 2 1
DAY6		6 2 9 4 2 1
DAY7		6 2 4 2 1
DAY8		6 2 4 1

TREATMENT :32% RP2  
REPLICATE 1

SAMPLE B

	GASTRULA	DEVELOPMENTAL STAGE HIPPO H'LING N'NATE
DAY1	9 10	
DAY4		9 10
DAY5		7 10
DAY6		7 10

TREATMENT :32% RP2  
REPLICATE 2

SAMPLE A

	GASTRULA	DEVELOPMENTAL STAGE HIPPO H'LING N'NATE
DAY1	10 12 12	
DAY4		5 3 2
DAY5		5 3 2
DAY6		5 3 2
DAY7		5 3 2

TREATMENT :32% RP2  
REPLICATE 2

SAMPLE B

	GASTRULA	DEVELOPMENTAL STAGE HIPPO H'LING N'NATE
DAY1	7	
DAY4		7
DAY5		6
DAY6		5
DAY7		4
DAY8		4
DAY9		4

TREATMENT :100% RP2  
REPLICATE 1

SAMPLE A

	GASTRULA	DEVELOPMENTAL STAGE HIPPO H'LING N'NATE
DAY1	10 12 10	
DAY4		9 1 7
DAY5		9 1 7
DAY6		9 1 7
DAY7		9 1 7
DAY8		9 1 7
DAY9		9 1 7
DAY10		9 1 7

TREATMENT :100% RP2  
REPLICATE 1

SAMPLE B

	GASTRULA	DEVELOPMENTAL STAGE HIPPO H'LING N'NATE
DAY1	10	
DAY4		10
DAY5		10
DAY6		9
DAY7		9
DAY8		9
DAY9		9
DAY10		9

TREATMENT :100% RP2  
REPLICATE 2

SAMPLE A

	GASTRULA	DEVELOPMENTAL STAGE HIPPO H'LING N'NATE
DAY1	4 6 4 5	
DAY4		1 5 2 3
DAY5		1 3 2 3
DAY6		1 3 2 3
DAY7		1 3 2 3
DAY8		1 3 2 3
DAY9		1 3 2 3
DAY10		3 2 3

TREATMENT :100% RP2  
REPLICATE 2

# SAMPLE B

## DEVELOPMENTAL STAGE

	GASTRULA	HIPPO	H'LING	N'NATE
DAY1	10 10			
DAY4		10 10		
DAY5			10 10	
DAY6			10 10	
DAY7			10 10	
DAY8			10 10	
DAY9			10 10	
DAY10				10 10

## Appendix 4.b.1.

TRIAL #3 COMMENCED 13/3/91 PAGE 1  
ENDPOINT:DEVELOPMENTAL EMBRYONIC MORTALITY  
SPECIES A. carinata

TREATMENT:CONTROL DEVELOPMENTAL STAGE RAW DATA  
GASTRULA TO HIPPO H'LING N'NATE

REP 1A  
REP 1B  
REP 2A  
REP 2B

GASTRULA TO HIPPO H'LING PERCENTILE  
N'NATE

REP 1A  
REP 1B  
REP 2A  
REP 2B

TOTAL 0 0 0  
PERCENT. 0 0 0

TREATMENT:1% RP2 DEVELOPMENTAL STAGE RAW DATA  
GASTRULA TO HIPPO H'LING N'NATE

REP 1A  
REP 1B  
REP 2A  
REP 2B

GASTRULA TO HIPPO H'LING PERCENTILE  
N'NATE

REP 1A 1  
REP 1B 1  
REP 2A  
REP 2B 1 4  
TOTAL 3 4 0  
PERCENT. 3 4 0

TREATMENT:3.2% RP2 DEVELOPMENTAL STAGE RAW DATA  
GASTRULA TO HIPPO H'LING

REP 1A  
REP 1B  
REP 2A  
REP 2B

GASTRULA TO HIPPO H'LING PERCENTILE  
N'NATE

REP 1A  
REP 1B  
REP 2A  
REP 2B

TOTAL 0 0 0  
PERCENT. 0 0 0

TREATMENT:10% RP2 DEVELOPMENTAL STAGE RAW DATA  
GASTRULA TO HIPPO H'LING N'NATE

REP 1A  
REP 1B  
REP 2A  
REP 2B

GASTRULA TO HIPPO H'LING PERCENTILE  
N'NATE

REP 1A  
REP 1B  
REP 2A  
REP 2B 2  
TOTAL 2 0 0  
PERCENT. 1.9 0 0

## Appendix 4.b.2.

TREATMENT: 32% RP2 DEVELOPMENTAL STAGE RAW DATA  
GASTRULA TO HIPPO H'LING N'NATE

REP 1A		11	
REP 1B			
REP 2A			
REP 2B			
	GASTRULA TO HIPPO	H'LING	PERCENTILE N'NATE
REP 1A		45.8	
REP 1B			
REP 2A			
REP 2B			
TOTAL	0	11	0
PERCENT.	0	10.7	0

TREATMENT: 100% RP2 DEVELOPMENTAL STAGE RAW DATA  
GASTRULA TO HIPPO H'LING N'NATE

REP 1A	1		
REP 1B		1	
REP 2A	1		
REP 2B	2	2	
	GASTRULA TO HIPPO	H'LING	PERCENTILE N'NATE
REP 1A	5		
REP 1B		3.1	
REP 2A	4.2		
REP 2B	6.7	6.7	
TOTAL	4	3	0
PERCENT.	3.8	2.9	0

TRIAL #1 COMMENCED 11/1/91 PAGE 1  
ENDPOINT: DEVELOPMENTAL EMBRYONIC MORTALITY RAW DATA  
SPECIES A. cummingii

TREATMENT : CONTROL DEVELOPMENTAL STAGE  
GASTRULA TO HIPPO H'LING N'NATE

REP 1A	1		
REP 1B		2	
REP 2A			
REP 2B			
	GASTRULA TO HIPPO	H'LING	PERCENTILE DATA N'NATE
REP 1A	3.6		
REP 1B		9.5	
REP 2A			
REP 2B			
TOTAL	1	2	0
PERCENT.	1	2	0

TRIAL #1 COMMENCED 11/1/91 PAGE 2  
ENDPOINT: DEVELOPMENTAL EMBRYONIC MORTALITY RAW DATA  
SPECIES A. cummingii

TREATMENT : 1% RP2 DEVELOPMENTAL STAGE  
GASTRULA TO HIPPO H'LING N'NATE

REP 1A	1		
REP 1B	1		
REP 2A			
REP 2B	5		
	GASTRULA TO HIPPO	H'LING	PERCENTILE DATA N'NATE
REP 1A	3.5		
REP 1B	5		
REP 2A			
REP 2B	18.5		
TOTAL	7	0	0
PERCENT.	7.1	0	0

TREATMENT : 3.2% RP2 DEVELOPMENTAL STAGE  
GASTRULA TO HIPPO H'LING N'NATE

REP 1A	2	2	
REP 1B	1		
REP 2A	1		
REP 2B	3		
	GASTRULA TO HIPPO	H'LING	PERCENTILE DATA N'NATE
REP 1A	6.9	6.9	
REP 1B	5		
REP 2A	4.4		
REP 2B	11.1		
TOTAL	7	2	0
PERCENT.	7	2	0

TREATMENT : 10% RP2 DEVELOPMENTAL STAGE  
GASTRULA TO HIPPO H'LING N'NATE

REP 1A			
REP 1B			
REP 2A		1	
REP 2B			
	GASTRULA TO HIPPO	H'LING	PERCENTILE DATA N'NATE
REP 1A			
REP 1B			
REP 2A		4	
REP 2B			
TOTAL	0	1	0
PERCENT.	0	1	0

TRIAL #3 COMMENCED 11/3/91 PAGE 5  
 ENDPOINT: DEVELOPMENTAL EMBRYONIC MORTALITY  
 SPECIES A. cummingii RAW DATA

TREATMENT : 32% RP2 DEVELOPMENTAL STAGE  
 GASTRULA TO HIPPO H'LING N'NATE

REP 1A	1		
REP 1B	1		
REP 2A	3		
REP 2B			

PERCENTILE DATA

	GASTRULA TO HIPPO	H'LING	N'NATE
REP 1A	5		
REP 1B	4.2		
REP 2A	12.5		
REP 2B			
TOTAL	5	0	0
PERCENT.	5.4	0	0

TREATMENT : 100% RP2 DEVELOPMENTAL STAGE  
 GASTRULA TO HIPPO H'LING N'NATE

REP 1A		1	
REP 1B	1		
REP 2A			
REP 2B		4	

PERCENTILE DATA

	GASTRULA TO HIPPO	H'LING	N'NATE
REP 1A		3.4	
REP 1B	5		
REP 2A			
REP 2B		16.7	
TOTAL	1	5	0
PERCENT.	1	5	0

#### Appendix 4.b.3.

TRIAL #3 COMMENCED 11/3/91 PAGE 1  
 ENDPOINT: DEVELOPMENTAL EMBRYONIC MORTALITY  
 SPECIES GLYPTOPHYSA SP.

TREATMENT: CONTROL RAW DATA  
 BLASTULA TO HIPPO H'LING N'NATE

REP 1A	1		
REP 1B	1	2	
REP 2A	9		
REP 2B		1	

PERCENTILE DATA

	BLASTULA TO HIPPO	H'LING	N'NATE
REP 1A	3.2		
REP 1B	3.2	6.5	
REP 2A	31		
REP 2B		7.1	
TOTAL	11	3	0
PERCENT.	10.5	2.9	0

TREATMENT: 1% RP2 RAW DATA  
 BLASTULA TO HIPPO H'LING N'NATE

REP 1A			
REP 1B	2		
REP 2A	4		
REP 2B	9		

PERCENTILE DATA

	BLASTULA TO HIPPO	H'LING	N'NATE
REP 1A			
REP 1B	6.9		
REP 2A	18.2		
REP 2B	81.1		
TOTAL	15	0	0
PERCENT.	19.7	0	0

TREATMENT: 3.2% RP2 DEVELOPMENTAL STAGE  
 BLASTULA TO HIPPO H'LING N'NATE

REP 1A	2		
REP 1B	3		
REP 2A	11	6	
REP 2B	4	1	

PERCENTILE DATA

	BLASTULA TO HIPPO	H'LING	N'NATE
REP 1A	15.4		
REP 1B	14.3		
REP 2A	35.5	19.4	
REP 2B	40	10	
TOTAL	20	7	0
PERCENT.	26.7	9.3	0

TREATMENT: 10% RP2 DEVELOPMENTAL STAGE  
 BLASTULA TO HIPPO H'LING N'NATE

REP 1A	20		
REP 1B	4		
REP 2A			
REP 2B			

PERCENTILE DATA

	BLASTULA TO HIPPO	H'LING	N'NATE
REP 1A	66.7		
REP 1B	50		
REP 2A			
REP 2B			
TOTAL	24	0	0
PERCENT.	28.6	0	0

TREATMENT: 32% RP2 DEVELOPMENTAL STAGE  
 BLASTULA TO HIPPO H'LING N'NATE

REP 1A		2	
REP 1B	25		2
REP 2A	24		
REP 2B	1	2	

PERCENTILE DATA

	BLASTULA TO HIPPO	H'LING	N'NATE
REP 1A		10.5	
REP 1B	36.8		2.9
REP 2A	70.6		
REP 2B	14.3	28.6	
TOTAL	50	4	2
PERCENT.	39.7	3.2	1.6

TREATMENT: 100% RP2 DEVELOPMENTAL STAGE  
 BLASTULA TO HIPPO H'LING N'NATE

REP 1A	15		
REP 1B	1		
REP 2A	8	2	
REP 2B			

PERCENTILE DATA

	BLASTULA TO HIPPO	H'LING	N'NATE
REP 1A	46.9		
REP 1B	10		
REP 2A	42.1	10.5	
REP 2B			
TOTAL	24	2	0
PERCENT.	29.6	2.5	0



## Appendix 4.c.

TRIAL #3 COMMENCED 13/3/91 PAGE 1  
ENDPOINT: JUVENILE MORTALITY EXPECTED: FROM NUMBERS OF  
SPECIES *A. cummingii* NEONATES HATCHED

REPLICATE 1						
TREATMENT	LIVING	SAMPLE A DEAD	% MORT	LIVING	SAMPLE B DEAD	% MORT
CONTROL	16	11	40.7	17	2	10.5
1% RP2	20	9	31	17	2	10.5
3.2% RP2	15	10	40	17	2	10.5
10% RP2	22	9	29	20	0	0
32% RP2	19	0	0	21	9	30
100% RP2	2	26	92.9	2	17	89.5

REPLICATE 2						
TREATMENT	LIVING	SAMPLE A DEAD	% MORT	LIVING	SAMPLE B DEAD	% MORT
CONTROL	18	10	35.7	19	2	9.5
1% RP2	18	4	18.2	20	2	9.1
3.2% RP2	22	0	0	21	3	12.5
10% RP2	23	2	8	21	3	12.5
32% RP2	10	11	52.4	11	12	52.2
100% RP2						

TRIAL #3 COMMENCED 13/3/91 PAGE 1  
ENDPOINT: JUVENILE MORTALITY EXPECTED: FROM NUMBERS OF  
SPECIES *A. carinata* NEONATES HATCHED

REPLICATE 1						
TREATMENT	LIVING	SAMPLE A DEAD	% MORT	LIVING	SAMPLE B DEAD	% MORT
CONTROL	19	5	20.8	28	4	14.3
1% RP2	20	9	69	17	2	10.5
3.2% RP2	29	3	9.4	1	19	95
10% RP2	18	3	14.3	26	3	10.3
32% RP2	27	2	6.9	8	5	18.5
100% RP2	5	14	73.7	2	29	93.5

REPLICATE 2						
TREATMENT	LIVING	SAMPLE A DEAD	% MORT	LIVING	SAMPLE B DEAD	% MORT
CONTROL	21	5	19.2	21	3	12.5
1% RP2	18	4	18.2	20	2	9.1
3.2% RP2	17	9	34.6	15	9	37.5
10% RP2	26	3	10.3	20	2	9.1
32% RP2	16	10	38.5	10	14	58.3
100% RP2	0	24	100	0	26	100

TRIAL #3 COMMENCED 13/3/91 PAGE 1  
ENDPOINT: JUVENILE MORTALITY EXPECTED: FROM NUMBERS OF  
SPECIES *GLYPTOPHYSA* SP. NEONATES HATCHED

REPLICATE 1						
TREATMENT	LIVING	SAMPLE A DEAD	% MORT	LIVING	SAMPLE B DEAD	% MORT
CONTROL	27	3	10	17	11	39.3
1% RP2	3	2	40	24	3	11.1
3.2% RP2	10	1	9.1	11	7	38.9
10% RP2	8	2	20	3	1	25
32% RP2	8	9	52.9	16	6	27.3
100% RP2	6	11	64.7	7	2	22.2

REPLICATE 2						
TREATMENT	LIVING	SAMPLE A DEAD	% MORT	LIVING	SAMPLE B DEAD	% MORT
CONTROL	14	6	30	6	7	53.8
1% RP2	17	1	5.6	4	0	0
3.2% RP2	10	4	28.7	2	3	60
10% RP2	24	0	0	17	5	22.7
32% RP2	0	8	100	0	4	100
100% RP2	1	7	87.5	8	12	60

## Appendix 4.d.

TRIAL #3 COMMENCED 15/3/91  
PHYSICO-CHEMICAL DATA

	DISSOLVED OXYGEN mg/L											
	CON	CON	1%RP2	1%RP2	3.2%RP2	3.2%RP2	10%RP2	10%RP2	32%RP2	32%RP2	100%RP2	100%RP2
DAY1	7.5	7.5	7.5	7.5	7.3	7.6	7.3	7.5	7.5	7.2	7.3	7.5
DAY2	7.4	7.5	7.7	7.5	7.5	7.7	7.5	7.7	7.7	7.5	7.5	7.5
DAY3	7.1	7.3	7.4	6.8	7.2	7.5	7.4	7.6	7.6	7.4	7.3	7.2
DAY4	7.1	7.1	7.2	7	7.1	7.1	7.2	7.1	7.1	7.1	7.1	7
DAY5	7.1	6.8	7.2	7.4	77.4	7	6.7	7.1	7.1	7.1	7.1	7
DAY6	7.5	7.6	7.4	7.7	7.6	7.6	7.6	7.6	7.6	7.3	7.5	7.1
DAY7	7.1	7.1	7.2	7.5	7.5	7.5	7.2	7.5	7.5	7.1	7.3	7.2
DAY8	7.4	7.3	7.2	7.5	7.5	7.5	7.2	7.6	7.6	7.6	7.7	7.5
DAY9	7.3	7.3	7.5	7.5	7.5	7.6	7.3	7.3	7.3	7.4	7.3	7.4
DAY10	7.4	7.3	7.4	7.2	7.2	7.2	7.5	7.2	7.2	7.2	7.1	7
DAY11	7.4	7.1	7.1	7.3	7.5	7.3	7.5	7.5	7.1	7.2	7.2	7.2

	pH											
	CON	CON	1%RP2	1%RP2	3.2%RP2	3.2%RP2	10%RP2	10%RP2	32%RP2	32%RP2	100%RP2	100%RP2
DAY1	6.24	6.54	6.65	6.67	6.7	6.73	6.76	6.85	6.96	7.09	7.25	7.34
DAY2	6.44	6.73	6.82	6.96	6.92	7.01	7.06	7.18	7.18	7.31	7.48	7.54
DAY3	6.81	6.89	7.01	7.14	6.81	7.07	7.17	7.27	7.39	7.39	7.51	7.56
DAY4	6.57	6.61	6.84	6.89	6.86	6.95	6.9	6.98	6.86	7.04	7.13	7.31
DAY5	6.5	6.79	6.66	6.87	6.72	6.71	6.85	6.91	6.92	7.03	7.02	7.2
DAY6	6.21	6.53	6.71	6.75	6.75	6.72	6.81	6.83	6.9	7	7.11	7.33
DAY7	6.18	6.61	6.7	6.7	6.69	6.71	6.81	6.93	6.87	6.96	7.18	7.4
DAY8	6.14	6.52	7.61	7.15	6.77	6.89	6.93	6.99	7.04	7.17	7.43	
DAY9	6.48	6.67	6.78	6.78	6.66	6.74	6.7	6.84	6.83	6.97	7.05	7.2
DAY10	6.31	6.7	6.69	6.71	6.61	8.41	7.7	6.72	7.15	7.18	7.31	7.4
DAY11	6.66	6.79	6.83	6.76	6.73	7.21	7.18	7.15	7	7.08	7.11	7.4

	CONDUCTIVITY uS/cm											
	CON	CON	1%RP2	1%RP2	3.2%RP2	3.2%RP2	10%RP2	10%RP2	32%RP2	32%RP2	100%RP2	100%RP2
DAY1	19.7	21.2	35	34	68.9	71.8	179	177	462	461	1170	1170
DAY2	19	23.4	39	37.3	73.4	75.2	188	183	469	476	1190	1200
DAY3	21.9	23.4	34.6	35.2	73.8	70.4	174	174	445	449	1150	1180
DAY4	22	23	35.8	38.4	76.1	73.7	179	177	454	463	1190	1200
DAY5	29	24.9	35.4	37.9	75	72.2	178	181	450	456	1170	1200
DAY6	19.8	26.6	36.7	38.1	73	72.6	181	179	461	462	1190	1190
DAY7	22.5	23.5	354.2	37.3	71.3	73.4	182	171	462	460	1190	1190
DAY8	23.1	25.6	106.5	39.5	71.8	72.3	182	176	465	458	1170	1180
DAY9	24.3	35.2	38.9	42.5	77.1	76.4	180	180	460	463	1180	1180
DAY10	24	25	38	39.6	74.6	255	189	183	468	467	1190	1200
DAY11	22.9	24.3	36.6	37.7	72.6	112	179	178	450	453	1160	1170

## Appendix 5.b.1.

TRIAL #4  
ENDPOINTCOMMENCED 17/3/91 CONDUCTED AT 25deg.C  
DEVELOPMENTAL CHARACTERISTICS PAGE 1  
EGG NUMBERS RAW DATA

## Appendix 5.a.

TRIAL #4  
ENDPOINT  
SPECIES  
COMMENCED 17/3/91 CONDUCTED AT 25deg.C  
DEVELOPMENTAL CHARACTERISTICS PAGE 1  
DAYS TO EGG CAPSULE RUPTURE  
RAW DATA  
A. carinata

TREATMENT	CONTROL	32% RP2	42% RP2	56% RP2	75% RP2	100% RP2
EGG MASS						
A	10	10	9	9	9	11
B	9	9	10	10	10	13
C	10	10	9	10	9	14
D	10	10	9	10	10	9
E	10	9	11	14	14	10
F	10	10	9	9	14	14
G	9	9	9	10	10	
H	10	12	10			

TRIAL #4  
ENDPOINT  
SPECIES  
COMMENCED 17/3/91 CONDUCTED AT 25deg.C  
DEVELOPMENTAL CHARACTERISTICS PAGE 1  
DAYS TO EGG CAPSULE RUPTURE  
RAW DATA  
A. cummingsii

TREATMENT	CONTROL	32% RP2	42% RP2	56% RP2	75% RP2	100% RP2
EGG MASS						
A	9	12	10	9	14	12
B	9	9	11	10	12	12
C	10	8	9	10	12	11
D	10	10	9	10	11	12
E	9	10	11	12	11	11
F	10	12	11	14	13	11
G	9	10	10	10	14	11
H	10	12	11	10	11	13
I	8	10	9	10	11	14
J	9	11	10	11	11	13
K	10	11	11	11	11	
L		11		12		
H		11				

TRIAL #4  
ENDPOINT  
SPECIES  
COMMENCED 17/3/91 CONDUCTED AT 25deg.C  
DEVELOPMENTAL CHARACTERISTICS PAGE 1  
DAYS TO EGG CAPSULE RUPTURE  
RAW DATA  
GLYPTOPHYSA SP.

TREATMENT	CONTROL	32% RP2	42% RP2	56% RP2	75% RP2	100% RP2
EGG MASS						
A	12	10	14	10	13	14
B	10	11	10	10	14	14
C			10	11	14	
D				12	14	
E				9		
F				11		

SPECIES

A. carinata

TREATMENT:CONTROL

REPLICATE 1

	GASTRULA	DEVELOPMENTAL STAGE				H'LING	N'NATE
		TROCH	VEL	HIPPO			
DAY1	9						
DAY2		9					
DAY3			9				
DAY4				9			
DAY5						9	
DAY6						9	
DAY7						9	
DAY8						9	
DAY9						9	
DAY10							9

TREATMENT:CONTROL

REPLICATE 2

	GASTRULA	DEVELOPMENTAL STAGE				H'LING	N'NATE
		TROCH	VEL	HIPPO			
DAY1	7 5						
DAY2		7 5					
DAY3			5		7		
DAY4				7 5			
DAY5				5		7	
DAY6						7 5	
DAY7						7 5	
DAY8						7 5	
DAY9							7
DAY10						5	
							5

TREATMENT:CONTROL

REPLICATE 3

	GASTRULA	DEVELOPMENTAL STAGE				H'LING	N'NATE
		TROCH	VEL	HIPPO			
DAY1	11 10 8						
DAY2			11 10				
DAY3		8					
DAY4				11 10			
DAY5				11 10 8			
DAY6						11 10 8	
DAY7						11 10 8	
DAY8						11 10 8	
DAY9						11 10 8	
DAY10							11 10
							8

TREATMENT: CONTROL		REPLICATE 4				
		DEVELOPMENTAL STAGE			H'LING	N'NATE
GASTRULA		TROCH	VEL	HIPPO		
DAY1	24 15					
DAY2		24	13			
DAY3			24 13			
DAY4				24 13		
DAY5					24 13	
DAY6					24 13	
DAY7					24 13	
DAY8					24 13	
DAY9					24	13
DAY10						24

TREATMENT: CONTROL		REPLICATE 5				
		DEVELOPMENTAL STAGE			H'LING	N'NATE
GASTRULA		TROCH	VEL	HIPPO		
DAY1	4					
DAY2		2				
DAY3			2			
DAY4				1		
DAY5					1	
DAY6					1	
DAY7					0	

TREATMENT: 32% RP2		REPLICATE 1				
		DEVELOPMENTAL STAGE			H'LING	N'NATE
GASTRULA		TROCH	VEL	HIPPO		
DAY1	8					
DAY2		7				
DAY3			7			
DAY4				7		
DAY5					6	
DAY6					6	
DAY7					6	
DAY8					6	
DAY9					6	
DAY10						6

TREATMENT: 32% RP2		REPLICATE 2				
		DEVELOPMENTAL STAGE			H'LING	N'NATE
GASTRULA		TROCH	VEL	HIPPO		
DAY1	16 9 7					
DAY2		8	16 9			
DAY3			16 9 8			
DAY4				16 9 8		
DAY5					14 9 8	
DAY6					13 9 8	
DAY7					13 9 8	
DAY8					13 9 8	
DAY9					9 8	13
DAY10						9 8

TREATMENT: 32% RP2		REPLICATE 3				
		DEVELOPMENTAL STAGE			H'LING	N'NATE
GASTRULA		TROCH	VEL	HIPPO		
DAY1	8 9					
DAY2		8	8			
DAY3			8 8			
DAY4				8 8		
DAY5					8 8	
DAY6					8 8	
DAY7					8 8	
DAY8					8 8	
DAY9					8	8
DAY10						8

TREATMENT: 32% RP2		REPLICATE 4				
		DEVELOPMENTAL STAGE			H'LING	N'NATE
GASTRULA		TROCH	VEL	HIPPO		
DAY1	10 17					
DAY2		16	10			
DAY3		16	10			
DAY4			16		10	
DAY5				16	10	
DAY6				16	10	
DAY7					10 16	
DAY8					10 16	
DAY9					16	10
DAY10					16	
DAY11					16	
DAY12						16

TREATMENT: 42% RP2		REPLICATE 1				
		DEVELOPMENTAL STAGE			H'LING	N'NATE
GASTRULA		TROCH	VEL	HIPPO		
DAY1	9 10					
DAY2		9	10			
DAY3			9 10			
DAY4				9 10		
DAY5					9 10	
DAY6					9 9	
DAY7					9 9	
DAY8					9 9	
DAY9					9	9
DAY10						9

TREATMENT: 42% RP2		REPLICATE 2				
		DEVELOPMENTAL STAGE			H'LING	N'NATE
GASTRULA		TROCH	VEL	HIPPO		
DAY1	7					
DAY2			7			
DAY3			7			
DAY4				7		
DAY5					7	
DAY6					7	
DAY7					7	
DAY8					7	
DAY9						7

TREATMENT: 42% RP2

## REPLICATE 3

	GASTRULA	TROCH	DEVELOPMENTAL STAGE VEL HIPPO	H'LING	N'NATE
DAY1	9				
DAY2			9		
DAY3			9		
DAY4				9	
DAY5				9	
DAY6				9	
DAY7				9	
DAY8				9	
DAY9					9

TREATMENT: 42% RP2

## REPLICATE 4

	GASTRULA	TROCH	DEVELOPMENTAL STAGE VEL HIPPO	H'LING	N'NATE
DAY1	20				
DAY2		20			
DAY3			20		
DAY4			20		
DAY5				20	
DAY6				20	
DAY7				20	
DAY8				20	
DAY9				20	
DAY10				20	
DAY11					20

TREATMENT: 42% RP2

## REPLICATE 5

	GASTRULA	TROCH	DEVELOPMENTAL STAGE VEL HIPPO	H'LING	N'NATE
DAY1	6 5 9				
DAY2		6 5	9		
DAY3			4 5 9		
DAY4				4 5 9	
DAY5				4 5 9	
DAY6				4 5 8	
DAY7				4 5 8	
DAY8				4 5	8
DAY9				4	5
DAY10					4

TREATMENT: 56% RP2

## REPLICATE 1

	GASTRULA	TROCH	DEVELOPMENTAL STAGE VEL HIPPO	H'LING	N'NATE
DAY1	1 5				
DAY2		1 5			
DAY3			1 5		
DAY4				1 5	
DAY5				1 5	
DAY6				1 5	
DAY7				1 5	
DAY8				1 5	
DAY9				1	5
DAY10					1

TREATMENT: 56% RP2

## REPLICATE 2

	GASTRULA	TROCH	DEVELOPMENTAL STAGE VEL HIPPO	H'LING	N'NATE
DAY1	15 10				
DAY2		15 10			
DAY3			15 10		
DAY4				15 10	
DAY5					15 10
DAY6					15 10
DAY7					15 10
DAY8					15 8
DAY9					15
DAY10					15

TREATMENT: 56% RP2

## REPLICATE 3

	GASTRULA	TROCH	DEVELOPMENTAL STAGE VEL HIPPO	H'LING	N'NATE
DAY1	7				
DAY2		7			
DAY3			7		
DAY4			1	6	
DAY5			1	6	
DAY6				4	3
DAY7				4	3
DAY8					3
DAY9					2
DAY10					2
DAY11					2
DAY12					2
DAY13					2
DAY14					2
					TERMINATION

TREATMENT: 56% RP2

## REPLICATE 4

	GASTRULA	TROCH	DEVELOPMENTAL STAGE VEL HIPPO	H'LING	N'NATE
DAY1	14 14				
DAY2		14	14		
DAY3			13	14	
DAY4				13 14	
DAY5					13 14
DAY6					13 14
DAY7					13 14
DAY8					13 14
DAY9					13
DAY10					14

TREATMENT: 75% RP2		REPLICATE 1				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE VEL HIPPO	H'LING	N'NATE	
DAY1	11 8					
DAY2		11 8				
DAY3			11 8			
DAY4				11 8		
DAY5				11 8		
DAY6				11 8		
DAY7				11 8		
DAY8				11 8		
DAY9				11		
DAY10					8 11	

TREATMENT: 75% RP2		REPLICATE 2				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE VEL HIPPO	H'LING	N'NATE	
DAY1	8 11					
DAY2		8	11			
DAY3			8	11		
DAY4			8	11		
DAY5				8		
DAY6				11		
DAY7				8 11		
DAY8				8 11		
DAY9				8 11		
DAY10				8 11		
DAY11				8 11		
DAY12				8 11		
DAY13				8 11		
DAY14				8 11		
TERMINATION						

TREATMENT: 75% RP2		REPLICATE 3				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE VEL HIPPO	H'LING	N'NATE	
DAY1	10 2					
DAY2		10 2				
DAY3			10 2			
DAY4				10		
DAY5				2		
DAY6				10 2		
DAY7				10 2		
DAY8				8 2		
DAY9				8 2		
DAY10				2	8 2	

TREATMENT: 75% RP2		REPLICATE 4				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE VEL HIPPO	H'LING	N'NATE	
DAY1	11					
DAY2			11			
DAY3			11			
DAY4				11		
DAY5				11		
DAY6				11		

DAY7						11
DAY8						11
DAY9						11
DAY10						11

TREATMENT: 100% RP2		REPLICATE 1				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE VEL HIPPO	H'LING	N'NATE	
DAY1	15					
DAY2		15				
DAY3			15			
DAY4				15		
DAY5				15		
DAY6					15	
DAY7					15	
DAY8					15	
DAY9					15	
DAY10					15	
DAY11						15

TREATMENT: 100% RP2		REPLICATE 2				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE VEL HIPPO	H'LING	N'NATE	
DAY1	7 4 7					
DAY2		7 4 7				
DAY3			7 4 7			
DAY4				7 4		
DAY5					3 4 7	2
DAY6						2 4 7
DAY7						2 4 7
DAY8						2 4 7
DAY9						2 4 7
DAY10						2 7
DAY11						2 7
DAY12						2 7
DAY13						7
DAY14						7
TERMINATION						

TREATMENT: 100% RP2		REPLICATE 3				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE VEL HIPPO	H'LING	N'NATE	
DAY1	13					
DAY2			13			
DAY3			13			
DAY4				13		
DAY5					13	
DAY6					13	
DAY7					13	
DAY8					13	
DAY9						13

TREATMENT: 100% RP2		REPLICATE				
	GASTRULA	DEVELOPMENTAL STAGE				
		TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	12 3					
DAY2		12 3				
DAY3			12 3			
DAY4				12 3		
DAY5					12 3	
DAY6					12 3	
DAY7					12 3	
DAY8					12 3	
DAY9					12 3	
DAY10						12
DAY11					3	
DAY12					3	
DAY13					3	
DAY14					3	
TERMINATION						

TREATMENT: CONTROL		REPLICATE 3				
	BLAST	DEVELOPMENTAL STAGE				
		TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	23 28					
DAY2		23 27				
DAY3			23 27			
DAY4			3	23 24		
DAY5			1	26	23	
DAY6					23 26	
DAY7					23 26	
DAY8					23 26	
DAY9					23 26	
DAY10						23 26

TREATMENT: CONTROL		REPLICATE 4				
	BLAST	DEVELOPMENTAL STAGE				
		TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	21 26					
DAY2		21 25				
DAY3			21 25			
DAY4			1	21 24		
DAY5				25	21	
DAY6					21 25	
DAY7					21 25	
DAY8					21 25	
DAY9					25	21
DAY10						25

## Appendix 5.b.2.

TRIAL #4 COMMENCED 17/1/91 CONDUCTED AT 25deg.C  
 ENDPOINT DEVELOPMENTAL CHARACTERISTICS PAGE 1  
 EGG NUMBERS  
 SPECIES A. cummingii RAW DATA

TREATMENT: CONTROL		REPLICATE 1				
	BLAST	DEVELOPMENTAL STAGE				
		TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	20 13 25					
DAY2		13 25	19			
DAY3			19 13 25			
DAY4				18 13 23		
DAY5				2	15 12 24	
DAY6					13 11 24	
DAY7					13 11 24	
DAY8					13 11 24	
DAY9						13 11
DAY10					24	24

TREATMENT: CONTROL		REPLICATE 2				
	BLAST	DEVELOPMENTAL STAGE				
		TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	25					
DAY2		24				
DAY3			24			
DAY4				24		
DAY5					24	
DAY6					24	
DAY7					24	
DAY8					24	
DAY9					24	
DAY10						24

TREATMENT: CONTROL		REPLICATE 5				
	BLAST	DEVELOPMENTAL STAGE				
		TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	27 23 29					
DAY2		26 23 29				
DAY3			23 29	26		
DAY4			23	29	26	
DAY5				23	26 23 29	
DAY6					26 23 29	
DAY7					26 23 29	
DAY8					23 29	26
DAY9					23	29
DAY10						23

TREATMENT: 32% RP2		REPLICATE 1				
	BLAST	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NAT
			VEL	HIPPO		
DAY1	29 24					
DAY2		26 23				
DAY3			25 21			
DAY4			5 21	18		
DAY5			1	4	19 20	
DAY6					23 21	
DAY7					23 21	
DAY8					22 21	
DAY9					22 21	
DAY10					22	21
DAY11					22	
DAY12						22

TREATMENT: 32% RP2		REPLICATE 2				
	BLAST	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NAT
			VEL	HIPPO		
DAY1	16 20					
DAY2		16 20				
DAY3			16 20			
DAY4			16			
DAY5				20		
DAY6				16	20	
DAY7				1	15 20	
DAY8					16 20	
DAY9					16	
DAY10					16	20
DAY11					16	
DAY12						16

TREATMENT: 32% RP2		REPLICATE 3				
	BLAST	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NAT
			VEL	HIPPO		
DAY1	31 20 34					
DAY2		31 20				
DAY3			34			
DAY4			31 20 34			
DAY5			1 20	29		
DAY6				33		
DAY7				2	28	
DAY8				20	33	
DAY9				1	29	
DAY10				1	19 33	
DAY11					29 20 33	
DAY12					29 20 33	
DAY13					29 20 33	
DAY14					29 20 33	
DAY15					29 20 33	
DAY16					29 20 33	
DAY17					29 20 33	
DAY18					29 20 33	
DAY19					29 20 33	
DAY20					29 20 33	
DAY21					29 20 33	
DAY22					29 20 33	
DAY23					29 20 33	
DAY24					29 20 33	
DAY25					29 20 33	
DAY26					29 20 33	
DAY27					29 20 33	
DAY28					29 20 33	
DAY29					29 20 33	
DAY30					29 20 33	
DAY31					29 20 33	
DAY32					29 20 33	
DAY33					29 20 33	
DAY34					29 20 33	
DAY35					29 20 33	
DAY36					29 20 33	
DAY37					29 20 33	
DAY38					29 20 33	
DAY39					29 20 33	
DAY40					29 20 33	
DAY41					29 20 33	
DAY42					29 20 33	
DAY43					29 20 33	
DAY44					29 20 33	
DAY45					29 20 33	
DAY46					29 20 33	
DAY47					29 20 33	
DAY48					29 20 33	
DAY49					29 20 33	
DAY50					29 20 33	
DAY51					29 20 33	
DAY52					29 20 33	
DAY53					29 20 33	
DAY54					29 20 33	
DAY55					29 20 33	
DAY56					29 20 33	
DAY57					29 20 33	
DAY58					29 20 33	
DAY59					29 20 33	
DAY60					29 20 33	
DAY61					29 20 33	
DAY62					29 20 33	
DAY63					29 20 33	
DAY64					29 20 33	
DAY65					29 20 33	
DAY66					29 20 33	
DAY67					29 20 33	
DAY68					29 20 33	
DAY69					29 20 33	
DAY70					29 20 33	
DAY71					29 20 33	
DAY72					29 20 33	
DAY73					29 20 33	
DAY74					29 20 33	
DAY75					29 20 33	
DAY76					29 20 33	
DAY77					29 20 33	
DAY78					29 20 33	
DAY79					29 20 33	
DAY80					29 20 33	
DAY81					29 20 33	
DAY82					29 20 33	
DAY83					29 20 33	
DAY84					29 20 33	
DAY85					29 20 33	
DAY86					29 20 33	
DAY87					29 20 33	
DAY88					29 20 33	
DAY89					29 20 33	
DAY90					29 20 33	
DAY91					29 20 33	
DAY92					29 20 33	
DAY93					29 20 33	
DAY94					29 20 33	
DAY95					29 20 33	
DAY96					29 20 33	
DAY97					29 20 33	
DAY98					29 20 33	
DAY99					29 20 33	
DAY100					29 20 33	

TREATMENT: 32% RP2		REPLICATE 4				
	BLAST	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NAT
			VEL	HIPPO		
DAY1	28 11					
DAY2		28 11				
DAY3			28 11			
DAY4				28 11		
DAY5					28 11	
DAY6					28 11	
DAY7					27 11	
DAY8					27 11	
DAY9					27 9	
DAY10					9	27
DAY11						9

TREATMENT: 32% RP2		REPLICATE 5				
	BLAST	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NAT
			VEL	HIPPO		
DAY1	10 24					
DAY2		8 24				
DAY3			8 24			
DAY4			2	4 24		
DAY5			1	5 24		
DAY6				1	4 24	
DAY7					5 24	
DAY8					5 24	
DAY9					5	24
DAY10					5	
DAY11					5	
DAY12						5

TREATMENT: 32% RP2		REPLICATE 6				
	BLAST	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NAT
			VEL	HIPPO		
DAY1	28					
DAY2		28				
DAY3			28			
DAY4				28		
DAY5					28	
DAY6					28	
DAY7					28	
DAY8						28

TREATMENT: 42% RP2		REPLICATE 1				
	BLAST	DEVELOPMENTAL STAGE			H'LING	N'NAT
		TROCH	VEL	HIPPO		
DAY1	22 29					
DAY2		22 29				
DAY3			21 28			
DAY4				21 28		
DAY5					21 28	
DAY6					20 28	
DAY7					20 28	
DAY8					20 28	
DAY9					20 28	
DAY10					20	28
DAY11						20

TREATMENT: 42% RP2		REPLICATE 2				
	BLAST	DEVELOPMENTAL STAGE			H'LING	N'NAT
		TROCH	VEL	HIPPO		
DAY1	12 34 16 30					
DAY2		12 34 16 30				
DAY3			12 34 16 30			
DAY4				12 34 16 30		
DAY5					12 34 16 30	
DAY6					12 34 16 30	
DAY7					12 34 16 30	
DAY8					12 34 16 30	
DAY9					12	34 16
DAY10					30	12 30

TREATMENT: 42% RP2		REPLICATE 3				
	BLAST	DEVELOPMENTAL STAGE			H'LING	N'NAT
		TROCH	VEL	HIPPO		
DAY1	12 26					
DAY2		12	26			
DAY3			12 26			
DAY4				12 26		
DAY5					12 26	
DAY6					12 26	
DAY7					12 26	
DAY8					11 26	
DAY9					11 25	
DAY10					20	11
DAY11						20

TREATMENT: 42% RP2		REPLICATE 4				
	BLAST	DEVELOPMENTAL STAGE			H'LING	
		TROCH	VEL	HIPPO		
DAY1	24 17 22					
DAY2		24 16 22				
DAY3			24 15 22			
DAY4				24 15 22		
DAY5					24 15 22	
DAY6					24 15 22	
DAY7					24 15 22	
DAY8					24 15 22	
DAY9					15 22	
DAY10					15 22	
DAY11					15 22	

TREATMENT: 56% RP2		REPLICATE 1				
	BLAST	DEVELOPMENTAL STAGE			H'LING	N'
		TROCH	VEL	HIPPO		
DAY1	34 25 29					
DAY2		34 25 29				
DAY3			34 25 28			
DAY4			1	33 25 10		
DAY5				15	2 8 11	32 17 8
DAY6					8 8	34 17 11
DAY7					6 8	33 19 11
DAY8					6 6	33 19 13
DAY9						31 25 16
DAY10						

TREATMENT: 56% RP2		REPLICATE 2				
	BLAST	DEVELOPMENTAL STAGE			H'LING	N'
		TROCH	VEL	HIPPO		
DAY1	19 27					
DAY2		19 27				
DAY3			19 25			
DAY4			7 5	12 17		
DAY5				8 18		11
DAY6			4	6 22		11
DAY7				2 18		11 4
DAY8					18	11 4
DAY9						11 20
DAY10						11 20
DAY11						11 20
DAY12						20
DAY13						20



TREATMENT: 56% RP2		REPLICATE 1				
	BLAST	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NAT
			VEL	HIPPO		
DAY1	14 17					
DAY2			14 27			
DAY3			14 27			
DAY4				14 27		
DAY5					14 27	
DAY6					14 27	
DAY7					14 27	
DAY8					14 27	
DAY9					14 27	
DAY10						14 27

TREATMENT: 56% RP2		REPLICATE 4				
	BLAST	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NAT
			VEL	HIPPO		
DAY1	9 35 27					
DAY2		9 34 27				
DAY3			9 14 27			
DAY4			9 8 15	26 7		
DAY5			5	4 7 15	26 7	
DAY6				2 7 4	7 26 16	
DAY7					8 25 18	
DAY8					8 25 18	
DAY9					8 25 18	
DAY10					7	25
DAY11						7 18

TREATMENT: 75% RP2		REPLICATE 1				
	BLAST	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NAT
			VEL	HIPPO		
DAY1	12 23 25					
DAY2		12 23 25				
DAY3			12 23 25			
DAY4				12 23 25		
DAY5				12 23 25		
DAY6					12 23 25	
DAY7					11 22 24	
DAY8					11 22 24	
DAY9					11 22 24	
DAY10					11 22 24	
DAY11					11 22 24	
DAY12					11	22
DAY13					11	24

TREATMENT: 75% RP2		REPLICATE 2				
	BLAST	TROCH	DEVELOPMENTAL STAGE		H'LING	N'
			VEL	HIPPO		
DAY1	23					
DAY2		23				
DAY3			23			
DAY4			8	15		
DAY5			8	15		
DAY6			8		15	
DAY7				7	15	
DAY8					16	
DAY9					16	
DAY10					16	
DAY11					16	
DAY12					16	
DAY13					16	
DAY14					16	
						TERMINATION

TREATMENT: 75% RP2		REPLICATE 3				
	BLAST	TROCH	DEVELOPMENTAL STAGE		H'LING	N'
			VEL	HIPPO		
DAY1	27 21					
DAY2		27 20				
DAY3			27 20			
DAY4			4	23 20		
DAY5			1	5 19	22	
DAY6				5	22 20	
DAY7					22 20	
DAY8					22 20	
DAY9					22 20	
DAY10					22 20	
DAY11						22 20

TREATMENT: 75% RP2		REPLICATE 4				
	BLAST	TROCH	DEVELOPMENTAL STAGE		H'LING	N'
			VEL	HIPPO		
DAY1	20 22					
DAY2		20 22				
DAY3			20 22			
DAY4				20 21		
DAY5				2	18 10	
DAY6				2	18 10	
DAY7				6	18 15	
DAY8					17 17	
DAY9					17 17	
DAY10					17 17	
DAY11						

TREATMENT:100% RP2		REPLICATE 1				
	BLAST	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NAT
			VEL	HIPPO		
DAY1	11 24					
DAY2		10 24				
DAY3			29 24			
DAY4			13 11	16 13		
DAY5			13 10	16 13		
DAY6				18 13	9 10	
DAY7				15	12 9	
DAY8					18 9	
DAY9					18 9	
DAY10					18 9	
DAY11					18 9	
DAY12					18 9	
DAY13					18 9	
DAY14					18 9	

TREATMENT:100% RP2		REPLICATE 2				
	BLAST	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NAT
			VEL	HIPPO		
DAY1	7 13 15					
DAY2		7 13 15				
DAY3			7 12 15			
DAY4				7		
DAY5			12 15			
DAY6			4	7 7 15		
DAY7				2	2 7 9 15	
DAY8					7 9 15	
DAY9					7 9 15	
DAY10					7 9 15	
DAY11					7 9	15
DAY12						7 9

TREATMENT:100% RP2		REPLICATE 3				
	BLAST	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NAT
			VEL	HIPPO		
DAY1	18 26					
DAY2		18 26				
DAY3			18 26			
DAY4			1 5	17 21		
DAY5			2	1 24	17	
DAY6				1 1	17 23	
DAY7				2	17 23	
DAY8					17 25	
DAY9					16 25	
DAY10					16 25	

TREATMENT:100% RP2		REPLICATE 4				
	BLAST	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NAT
			VEL	HIPPO		
DAY1	21 23 21					
DAY2		21 23 21				
DAY3			21 23 21			
DAY4			8 2 3	13 21 18		
DAY5			7	9 16 5	5 7 16	
DAY6				13 8 2	8 15 19	
DAY7				10 8 2	5 12 19	
DAY8					5 12 19	
DAY9					5 12 16	
DAY10					4 12 16	
DAY11					4	12 16
DAY12					4	
DAY13					4	
DAY14					4	

TREATMENT:100% RP2		REPLICATE 5				
	BLAST	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NAT
			VEL	HIPPO		
DAY1	12 16					
DAY2		11 16		1		
DAY3		2	9 16			
DAY4		2	9 16			
DAY5			4 8	7 7		
DAY6					7 13	4 2
DAY7				4 4	6 3	
DAY8					7 2	
DAY9					5 2	
DAY10					5 2	
DAY11						2
DAY12						2
DAY13						2
DAY14						2

TERMINATION

TREATMENT:100% RP2		REPLICATE 1				
	BLAST	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NAT
			VEL	HIPPO		
DAY1	31 24					
DAY2		30 24				
DAY3			29 24			
DAY4			13 11	16 13		
DAY5			13 10	16 13		
DAY6				18 13	9 10	
DAY7				15	12 9	
DAY8					18 9	
DAY9					18 9	
DAY10					18 9	
DAY11					18 9	
DAY12					18 9	
DAY13					18 9	
DAY14					18 9	

TREATMENT:100% RP2

REPLICATE 2

	BLAST	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NAT
			VEL	HIPPO		
DAY1	7 13 15					
DAY2		7 13 15				
DAY3			7 12 15			
DAY4				7		
DAY5			12 15	7 7 15		
DAY6			4	2	7 9 15	
DAY7			2	2	7 9 15	
DAY8			1	1	7 9 15	
DAY9					7 9 15	
DAY10					7 9 15	
DAY11					7 9	15

TREATMENT:100% RP2

REPLICATE 3

	BLAST	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NAT
			VEL	HIPPO		
DAY1	18 26					
DAY2		18 26				
DAY3			18 26			
DAY4			1 5	17 21		
DAY5			2	1 24	17	
DAY6			2	1 1	17 23	
DAY7			2	1	17 23	
DAY8					17 25	
DAY9					16 25	
DAY10					16 25	

TREATMENT:100% RP2

REPLICATE 4

	BLAST	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NAT
			VEL	HIPPO		
DAY1	21 23 21					
DAY2		21 23 21				
DAY3			21 23 21			
DAY4			8 2 3	13 21 18		
DAY5			7	9 16 5	5 7 16	
DAY6				13 8 2	8 15 19	
DAY7				10 8 2	5 12 19	
DAY8					5 12 19	
DAY9					5 12 16	
DAY10					4 12 16	
DAY11					4	12 16
DAY12					4	
DAY13					4	
DAY14					4	

TERMINATION

TREATMENT:100% RP2

REPLICATE 5

	BLAST	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NAT
			VEL	HIPPO		
DAY1	12 16					
DAY2		11 16		1		
DAY3		2		9 16		
DAY4		2		9 16		
DAY5				4 8	7 7	
DAY6					7 13	4 2
DAY7				4 4	6 3	
DAY8					7 2	
DAY9					5 2	
DAY10					5 2	
DAY11					2	
DAY12					2	
DAY13					2	
DAY14					2	

TERMINATION

# Appendix 5.b.3.

TRIAL #4 COMMENCED 17/3/91 CONDUCTED AT 25deg.C  
ENDPOINT DEVELOPMENTAL CHARACTERISTICS PAGE 1  
EGG NUMBERS RAW DATA  
SPECIES GLYPTOPHYSA SP.

TREATMENT: CONTROL		REPLICATE 1				
	GASTRULA	DEVELOPMENTAL STAGE				N'NATE
		TROCH	VEL	HIPPO	H'LING	
DAY1	11 8 8					
DAY2		11 1 7				
DAY3		9 1 7				
DAY4		2	2			
DAY5		5		2		
DAY6		2		3		
DAY7				1	1 3	
DAY8					1 3	
DAY9					1 3	
DAY10					1 3	
DAY11					1 3	
DAY12					1	3
DAY13					1	
DAY14					1	
TERMINATION						

TREATMENT: CONTROL		REPLICATE 2				
	GASTRULA	DEVELOPMENTAL STAGE				N'NATE
		TROCH	VEL	HIPPO	H'LING	
DAY1	8					
DAY2		8				
DAY3		7				
DAY4		5	1			
DAY5			1	4		
DAY6					4	
DAY7					4	
DAY8					4	
DAY9						4

TREATMENT: 32% RP2		REPLICATE 1				
	GASTRULA	DEVELOPMENTAL STAGE				N'NATE
		TROCH	VEL	HIPPO	H'LING	
DAY1	13 8					
DAY2		13 8				
DAY3				13		
DAY4				8	13	
DAY5					13 8	
DAY6					13 8	
DAY7					13 8	
DAY8					8	13
DAY9						8

TREATMENT: 32% RP2		REPLICATE 2				
	GASTRULA	DEVELOPMENTAL STAGE				N'NATE
		TROCH	VEL	HIPPO	H'LING	
DAY1	14					
DAY2		10				
DAY3			4			
DAY4				3		
DAY5					3	
DAY6					3	
DAY7					3	
DAY8					3	
DAY9					3	
DAY10						3

TREATMENT: 42% RP2		REPLICATE 1				
	GASTRULA	DEVELOPMENTAL STAGE				N'NATE
		TROCH	VEL	HIPPO	H'LING	
DAY1	10 10 8					
DAY2		9 7 7				
DAY3				9 7 6		
DAY4				9 7 6		
DAY5					9 7 3	
DAY6						9 7 2
DAY7						9 7 2
DAY8						9 7 2
DAY9						
DAY10						9 7
DAY11						2
DAY12						2
DAY13						2
DAY14						2

TREATMENT: 42% RP2		REPLICATE 2				
	GASTRULA	DEVELOPMENTAL STAGE				N'NATE
		TROCH	VEL	HIPPO	H'LING	
DAY1	6					
DAY2		6				
DAY3		6				
DAY4			5			
DAY5			4	1		
DAY6			DEAD	DEAD		

TREATMENT: 56% RP2		REPLICATE 1				
	GASTRULA	DEVELOPMENTAL STAGE				N'NATE
		TROCH	VEL	HIPPO	H'LING	
DAY1	14 12 7 11					
DAY2		12 11	12 7			
DAY3				1 6	10	
DAY4				1 11 3	10 6 8	
DAY5				1 11 3	10 6 7	
DAY6					10 11 6 6	
DAY7					10 11 6 7	
DAY8					10 11 5 7	
DAY9					11	10
DAY10					7	5
DAY11					11 7	11 7

TREATMENT: 56% RP2

REPLICATE 2

	GASTRULA	DEVELOPMENTAL STAGE			H'LING	N'NATE
		TROCH	VEL	HIPPO		
DAY1	8 9					
DAY2	8	9				
DAY3		8 1	8			
DAY4		1	7	8		
DAY5					7 8	
DAY6					7 8	
DAY7					7 8	
DAY8					7	8
DAY9					7	
DAY10						7

TREATMENT: 75% RP2

REPLICATE 1

	GASTRULA	DEVELOPMENTAL STAGE			H'LING	N'NATE
		TROCH	VEL	HIPPO		
DAY1	8 4 14					
DAY2		6 4 14				
DAY3		6 4 13				
DAY4			1 4 1	9 12		
DAY5				3 4 11	2	
DAY6					5 4 10	
DAY7					5 4 10	
DAY8					5 4 10	
DAY9					5 4 10	
DAY10					5 4 10	
DAY11					5 4 10	
DAY12						5
DAY13					4 10	10
DAY14					4	

TERMINATION

TREATMENT: 75% RP2

REPLICATE 2

	GASTRULA	DEVELOPMENTAL STAGE			H'LING	N'NATE
		TROCH	VEL	HIPPO		
DAY1	8					
DAY2		8				
DAY3		8				
DAY4			7			
DAY5				7		
DAY6					6	
DAY7					6	
DAY8					6	
DAY9					5	
DAY10					5	
DAY11					5	
DAY12					5	
DAY13					5	
DAY14					5	

TERMINATION

TREATMENT: 100% RP2

REPLICATE 1

	GASTRULA	DEVELOPMENTAL STAGE			H'LING	N'NATE
		TROCH	VEL	HIPPO		
DAY1	9 7					
DAY2		8 7				
DAY3		8 6				
DAY4			7			
DAY5				6 7 6		
DAY6						4 5
DAY7						4 5
DAY8						4 6
DAY9						4 6
DAY10						4 5
DAY11						4 5
DAY12						4 6
DAY13						4 6
DAY14						4

TERMINATION

TREATMENT: 100% RP2

REPLICATE 2

	GASTRULA	DEVELOPMENTAL STAGE			H'LING	N'NATE
		TROCH	VEL	HIPPO		
DAY1	13					
DAY2		7				
DAY3		7				
DAY4			4			
DAY5				4		
DAY6						1
DAY7						1
DAY8						1
DAY9						1
DAY10						1
DAY11						1
DAY12						1
DAY13						1
DAY14						DEAD

## Appendix 5.c.1.

TRIAL #4 COMMENCED 13/3/91 PAGE 1  
ENDPOINT: DEVELOPMENTAL EMBRYONIC MORTALITY  
SPECIES A. carinata

TREATMENT: CONTROL	DEVELOPMENTAL STAGE				RAW DATA
	GASTRULA	TROCH	VEL	HIPPO	H'LING
REP 1					
REP 2					
REP 3					
REP 4		2			
REP 5	2		1		1

	GASTRULA	TROCH	VEL	HIPPO	PERCENTILE
					H'LING
REP 1					
REP 2					
REP 3					
REP 4		5.1			
REP 5	50		25	25	
TOTAL	2	2	1	0	1
PERCENT.	2.1	2.1	1.1	0	1.1

TREATMENT: 32% RP2	DEVELOPMENTAL STAGE				RAW DATA
	GASTRULA	TROCH	VEL	HIPPO	H'LING
REP 1	1			1	
REP 2				2	1
REP 3	1				
REP 4		1			
	GASTRULA	TROCH	VEL	HIPPO	PERCENTILE
					H'LING
REP 1	12.5			12.5	
REP 2				6.1	3
REP 3	5.9				
REP 4		3.7			
TOTAL	2	1	0	3	1
PERCENT.	2.4	1.2	0	3.5	1.2

TREATMENT: 42% RP2	DEVELOPMENTAL STAGE				RAW DATA
	GASTRULA	TROCH	VEL	HIPPO	H'LING
REP 1					1
REP 2					
REP 3					
REP 4					
REP 5			1		1
	GASTRULA	TROCH	VEL	HIPPO	PERCENTILE
					H'LING
REP 1					5.3
REP 2					
REP 3					
REP 4					
REP 5			5		5
TOTAL	0	0	1	0	2
PERCENT.	0	0	1.3	0	2.7

TREATMENT: 56% RP2	DEVELOPMENTAL STAGE				RAW DATA
	GASTRULA	TROCH	VEL	HIPPO	H'LING
REP 1					
REP 2					2
REP 3				4	1
REP 4		1			
	GASTRULA	TROCH	VEL	HIPPO	PERCENTILE
					H'LING
REP 1					
REP 2					
REP 3				57.1	14.3
REP 4		3.6			
TOTAL	0	1	0	4	3
PERCENT.	0	1.5	0	6.1	4.5

TREATMENT: 75% RP2	DEVELOPMENTAL STAGE				RAW DATA
	GASTRULA	TROCH	VEL	HIPPO	H'LING
REP 1					
REP 2					8
REP 3					2
REP 4					

	GASTRULA	TROCH	VEL	HIPPO	PERCENTILE
					H'LING
REP 1					
REP 2					42.1
REP 3					16.7
REP 4					
TOTAL	0	0	0	0	10
PERCENT.	0	0	0	0	16.4

TREATMENT: 100% RP2	DEVELOPMENTAL STAGE				RAW DATA
	GASTRULA	TROCH	VEL	HIPPO	H'LING
REP 1					
REP 2				5	9
REP 3					
REP 4					3
	GASTRULA	TROCH	VEL	HIPPO	PERCENTILE
					H'LING
REP 1					
REP 2				27.8	50
REP 3					
REP 4					20
TOTAL	0	0	0	5	12
PERCENT.	0	0	0	8.2	19.7

## Appendix 5.c.2.

TRIAL #4 COMMENCED 11/3/91 PAGE 1  
ENDPOINT: DEVELOPMENTAL EMBRYONIC MORTALITY  
SPECIES A. cummingii

TREATMENT: CONTROL	GASTRULA	DEVELOPMENTAL STAGE				RAW DATA
		TROCH	VEL	HIPPO	H'LING	
REP 1	2	1	2	4	3	
REP 2		1				
REP 3		1			1	
REP 4		1				
REP 5	1					
PERCENTILE DATA						
	GASTRULA	TROCH	VEL	HIPPO	H'LING	
REP 1	3.3	1.7	3.3	6.7	5	
REP 2		4				
REP 3		2			2	
REP 4		2.1				
REP 5	1.3					
TOTAL	3	4	2	4	4	
PERCENT.	1.1	1.5	0.8	1.5	1.5	

TREATMENT: 32% RP2	GASTRULA	DEVELOPMENTAL STAGE				RAW DATA
		TROCH	VEL	HIPPO	H'LING	
REP 1	11	3	2		1	
REP 2	4					
REP 3			1	1	1	
REP 4	2				3	
REP 5	1	2	1			
REP 6						
PERCENTILE DATA						
	GASTRULA	TROCH	VEL	HIPPO	H'LING	
REP 1	11.7	6.7	3.3		1.7	
REP 2	10					
REP 3			1.2	1.2	1.2	
REP 4	4.9				7.3	
REP 5	2.9	5.7	2.9			
REP 6						
TOTAL	18	5	4	1	5	
PERCENT.	6.2	1.7	1.4	0.3	1.7	

TREATMENT: 42% RP2	GASTRULA	DEVELOPMENTAL STAGE				RAW DATA
		TROCH	VEL	HIPPO	H'LING	
REP 1	1	2			1	
REP 2						
REP 3					7	
REP 4	2	1				
PERCENTILE DATA						
	GASTRULA	TROCH	VEL	HIPPO	H'LING	
REP 1	1.9	3.8			1.9	
REP 2						
REP 3					15.8	
REP 4	1.6	3.1				
TOTAL	3	3	0	0	8	
PERCENT.	1.2	1.2	0	0	3.3	

TREATMENT: 56% RP2	GASTRULA	DEVELOPMENTAL STAGE				RAW DATA
		TROCH	VEL	HIPPO	H'LING	
REP 1		1	3	6	6	
REP 2	1	2		10		
REP 3						
REP 4	1		6	16	2	
PERCENTILE DATA						
	GASTRULA	TROCH	VEL	HIPPO	H'LING	
REP 1		1.1	3.4	6.8	6.8	
REP 2	1.6	3.2		15.6		
REP 3						
REP 4	1.4		8.4	22.5	2.8	
TOTAL	2	3	9	32	8	
PERCENT.	0.9	1.3	3.9	13.6	3.4	

TREATMENT: 75% RP2	GASTRULA	DEVELOPMENTAL STAGE				RAW DATA
		TROCH	VEL	HIPPO	H'LING	
REP 1	2			1	2	
REP 2			1	6		
REP 3	4			5		
REP 4	1			4	1	

PERCENTILE DATA						
	GASTRULA	TROCH	VEL	HIPPO	H'LING	
REP 1	3.2			1.6	3.2	
REP 2			4.3	26.1		
REP 3	7.8			9.8		
REP 4	2.3			9.3	2.3	
TOTAL	7	0	1	16	3	
PERCENT.	3.9	0	0.6	8.9	1.7	

TREATMENT: 100% RP2	GASTRULA	DEVELOPMENTAL STAGE				RAW DATA
		TROCH	VEL	HIPPO	H'LING	
REP 1	1	1	3	22	1	
REP 2	6	1	3	2	6	
REP 3			1	1	1	
REP 4				19	14	
REP 5		1	1	17	2	

PERCENTILE DATA						
	GASTRULA	TROCH	VEL	HIPPO	H'LING	
REP 1	1.8	1.8	5.5	0.4	1.8	
REP 2	14.6	2.4	7.3	4.9	14.6	
REP 3			2.4	2.4	2.4	
REP 4				29.2	21.5	
REP 5		3.6	3.6	60.7	7.1	
TOTAL	7	3	8	61	24	
PERCENT.	3	1.3	3.4	26.3	10.3	

# Appendix 5.c.3.

TRIAL #4 COMMENCED 11/3/91 PAGE 1  
ENDPOINT: DEVELOPMENTAL EMBRYONIC MORTALITY  
SPECIES GLYPTOPHYSA SP.

TREATMENT: CONTROL		DEVELOPMENTAL STAGE				RAW DATA
GASTRULA		TROCH	VEL	HIPPO	H'LING	
REP 1	8	15	1	1		
REP 2		2	2			
REP 3	2		1		1	
GASTRULA		TROCH	VEL	HIPPO	H'LING	PERCENTILE
REP 1	29.6	55.6	3.7	3.7		
REP 2		25	25			
REP 3	50		25		25	
TOTAL	10	17	4	1	1	
PERCENT.	25.6	43.6	10.3	2.6	2.6	

TREATMENT: 32% RP2		DEVELOPMENTAL STAGE				RAW DATA
GASTRULA		TROCH	VEL	HIPPO	H'LING	
REP 1						
REP 2	4	6	1			
GASTRULA		TROCH	VEL	HIPPO	H'LING	PERCENTILE
REP 1						
REP 2	28.6	42.9	7.1			
TOTAL	4	6	1	0	0	
PERCENT.	11.4	17.1	2.9	0	0	

TREATMENT: 42% RP2		DEVELOPMENTAL STAGE				RAW DATA
GASTRULA		TROCH	VEL	HIPPO	H'LING	
REP 1	5	1	1			
REP 2		1	4	1		
GASTRULA		TROCH	VEL	HIPPO	H'LING	PERCENTILE
REP 1	17.9	3.6	3.6			
REP 2		16.7	66.7	16.7		
TOTAL	5	2	5	1	0	
PERCENT.	14.7	5.9	14.7	2.9	0	

TREATMENT: 56% RP2		DEVELOPMENTAL STAGE				RAW DATA
GASTRULA		TROCH	VEL	HIPPO	H'LING	
REP 1	2	3	3	2	1	
REP 2		2				
GASTRULA		TROCH	VEL	HIPPO	H'LING	PERCENTILE
REP 1	4.5	6.8	6.8	4.5	2.3	
REP 2		11.8				
TOTAL	2	5	3	2	1	
PERCENT.	3.3	8.2	4.9	3.3	1.6	

TREATMENT: 75% RP2		DEVELOPMENTAL STAGE				RAW DATA
GASTRULA		TROCH	VEL	HIPPO	H'LING	
REP 1	2	1	2	2	4	
REP 2		1		1	6	
GASTRULA		TROCH	VEL	HIPPO	H'LING	PERCENTILE
REP 1	7.7	3.8	7.7	7.7	15.4	
REP 2		12.5		12.5	75	
TOTAL	2	2	2	3	10	
PERCENT.	5.9	5.9	5.9	8.8	29.4	

TREATMENT: 100% RP2		DEVELOPMENTAL STAGE				RAW DATA
GASTRULA		TROCH	VEL	HIPPO	H'LING	
REP 1	1	2		3	6	
REP 2	6	3		3	1	
GASTRULA		TROCH	VEL	HIPPO	H'LING	PERCENTILE
REP 1	6.3	12.5		18.8	37.5	
REP 2	46.2	23.1		23.1	7.7	
TOTAL	7	5	0	6	7	
PERCENT.	26.1	17.2	0	20.7	24.1	



## Appendix 6.a.1.

TRIAL #5 COMMENCED 31/3/91 PAGE 1  
ENDPOINT: DAILY EGGMASS NUMBER  
SPECIES: A. cacinaca

RAW DATA

TREATMENT :CONTROL

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	1	3	2	1	0	0
DAY2	2	2	0	1	1	4
DAY3	3	4	3	2	2	4
DAY4	1	0	3	1	1	2
DAY5	4	3	2	4	2	2
DAY6	2	2	2	1	2	1
DAY7	2	1	4	2	3	3

TREATMENT :1% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	0	0	0	1
DAY2	1	2	1	0	1	0
DAY3	4	0	2	1	1	1
DAY4	0	0	1	0	1	1
DAY5	6	2	3	0	1	1
DAY6	3	2	1	1	1	0
DAY7	1	3	2	0	0	3

TREATMENT :3.2% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	2	2	1	1
DAY2	1	1	0	0	0	0
DAY3	2	1	3	3	3	3
DAY4	0	1	1	0	0	0
DAY5	0	1	0	4	2	4
DAY6	1	3	2	1	2	1
DAY7	1	2	2	1	1	1

TREATMENT :10% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	1	3	1	0
DAY2	0	2	0	1	0	0
DAY3	3	1	3	4	0	3
DAY4	0	0	1	0	1	1
DAY5	2	1	0	2	2	2
DAY6	1	1	1	0	0	1
DAY7	0	0	0	1	1	2

TREATMENT :32% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	0	0	0	0
DAY2	0	0	0	0	0	1
DAY3	0	0	0	0	0	0
DAY4	0	0	0	0	0	1
DAY5	0	0	0	1	1	0
DAY6	0	0	0	0	0	1
DAY7	0	1	0	0	1	0

TREATMENT :100% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	0	0	0	0
DAY2	0	0	0	0	1	0
DAY3	0	0	0	2	2	1
DAY4	0	0	0	0	1	1
DAY5	0	0	0	0	0	0
DAY6	0	1	0	0	0	0
DAY7	0	0	0	0	0	0

TRIAL #5 COMMENCED 31/3/91 PAGE 1  
ENDPOINT: DAILY TOTAL EGG PRODUCTION  
SPECIES: A. carinata

RAW DATA

TREATMENT :CONTROL

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	12	25	45	13	0	0
DAY2	17	18	13	13	3	53
DAY3	0	28	36	32	20	40
DAY4	0	0	22	20	8	28
DAY5	0	10	23	58	14	24
DAY6	0	16	18	17	13	16
DAY7	0	0	36	27	24	35

TREATMENT :1% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	0	0	0	8
DAY2	11	5	11	0	7	14
DAY3	44	0	22	5	7	6
DAY4	0	0	15	0	11	7
DAY5	63	8	37	0	3	1
DAY6	31	18	20	4	10	0
DAY7	9	23	24	0	0	15

TREATMENT :3.2% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	13	9	13	0
DAY2	10	5	0	0	0	9
DAY3	21	0	22	25	23	31
DAY4	11	9	4	0	0	0
DAY5	0	8	0	42	15	32
DAY6	13	14	18	13	24	9
DAY7	5	5	13	11	11	9

TREATMENT :10% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	9	31	15	0
DAY2	0	17	0	10	0	0
DAY3	15	12	19	41	0	27
DAY4	0	0	8	0	15	11
DAY5	13	0	0	21	33	23
DAY6	3	12	5	0	16	10
DAY7	0	0	8	12	13	18

TREATMENT :32% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	0	0	0	0
DAY2	0	0	0	0	0	2
DAY3	0	0	0	0	0	0
DAY4	0	0	0	0	0	7
DAY5	0	0	0	13	7	0
DAY6	0	0	0	0	0	5
DAY7	0	4	0	0	8	0

TREATMENT :100% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	0	0	0	0
DAY2	0	0	0	0	0	0
DAY3	0	0	0	0	0	0
DAY4	0	0	0	0	0	0
DAY5	0	0	0	0	0	0
DAY6	0	0	0	0	0	0
DAY7	0	0	0	0	0	0

## Appendix 6.a.2.

TRIAL #5 COMMENCED 31/3/91 PAGE 1  
ENDPOINT: DAILY EGGMASS NUMBER  
SPECIES: A. cumingii

RAW DATA

TREATMENT :CONTROL

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	1	3	2	1	1	2
DAY2	2	2	4	1	1	1
DAY3	1	3	4	3	3	3
DAY4	0	1	1	0	1	2
DAY5	0	1	2	3	1	2
DAY6	1	2	2	1	1	2
DAY7	1	3	3	3	2	2

TREATMENT :1% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	1	1	2	1	2	1
DAY2	2	1	2	2	3	3
DAY3	1	3	4	1	1	1
DAY4	2	1	2	2	2	0
DAY5	4	2	2	2	4	2
DAY6	1	0	2	2	2	3
DAY7	2	2	0	1	2	1

TREATMENT :3.2% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	1	0	2	1	0
DAY2	2	2	0	3	5	3
DAY3	2	1	4	3	1	2
DAY4	2	3	0	0	0	0
DAY5	3	1	3	3	0	1
DAY6	2	2	2	2	1	2
DAY7	1	2	2	2	2	2

TREATMENT :10% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	0	1	0	0
DAY2	1	1	2	1	0	1
DAY3	1	3	0	2	2	1
DAY4	1	1	0	0	0	0
DAY5	0	1	0	2	0	1
DAY6	3	1	0	2	2	1
DAY7	2	2	0	2	2	2

TREATMENT :32% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	1	0	0	0	2	0
DAY2	0	0	0	0	0	0
DAY3	0	1	1	0	0	0
DAY4	0	1	0	0	0	0
DAY5	0	0	0	0	0	0
DAY6	0	0	1	1	1	0
DAY7	1	1	0	1	0	0

TREATMENT :100% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	0	0	1	0
DAY2	0	0	1	0	0	0
DAY3	1	1	0	0	0	0
DAY4	0	0	0	0	0	0
DAY5	0	0	0	0	0	0
DAY6	0	0	0	0	0	0
DAY7	0	0	0	0	0	0

TRIAL #5 COMMENCED 11/3/91 PAGE 1  
ENDPOINT: DAILY TOTAL EGG PRODUCTION  
SPECIES: A. cummingii

# RAW DATA

## TREATMENT :CONTROL

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	16	54	40	29	25	34
DAY2	28	56	75	20	15	17
DAY3	28	66	61	51	71	49
DAY4	0	26	22	0	16	29
DAY5	0	20	47	83	61	49
DAY6	34	56	30	28	13	27
DAY7	24	10	26	37	36	16

## TREATMENT :1% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	18	17	39	22	62	15
DAY2	40	15	45	46	51	36
DAY3	34	41	79	24	65	16
DAY4	38	18	35	35	54	0
DAY5	63	46	41	49	81	36
DAY6	2	0	21	30	39	48
DAY7	34	35	27	14	50	15

## TREATMENT :3.2% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	27	0	53	19	0
DAY2	60	44	0	38	77	53
DAY3	83	19	59	63	24	36
DAY4	52	44	0	0	0	0
DAY5	78	18	63	68	24	18
DAY6	53	35	31	10	7	28
DAY7	18	27	35	22	7	38

## TREATMENT :10% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	0	24	0	0
DAY2	18	16	25	22	0	31
DAY3	15	54	0	46	18	30
DAY4	25	25	0	0	0	0
DAY5	0	14	0	48	0	33
DAY6	40	14	0	29	33	14
DAY7	35	24	0	24	42	25

## TREATMENT :32% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	16	0	0	0	62	0
DAY2	0	0	0	0	0	0
DAY3	0	21	14	0	0	0
DAY4	0	21	0	0	0	0
DAY5	0	0	0	0	0	0
DAY6	0	0	10	10	19	0
DAY7	14	13	0	13	0	0

## TREATMENT :100% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	0	0	22	0
DAY2	0	0	22	0	0	0
DAY3	28	18	0	0	0	0
DAY4	0	0	0	0	0	0
DAY5	0	0	0	0	0	0
DAY6	0	0	0	0	0	0
DAY7	0	0	0	0	0	0

## Appendix 6.a.3.

TRIAL #5 COMMENCED 11/3/91 PAGE 1  
ENDPOINT: DAILY EGG MASS NUMBER  
SPECIES: GLYPTOPHYSA SP.

# RAW DATA

## TREATMENT:CONTROL

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	5	4	0	3	4
DAY2	0	1	2	0	0	3
DAY3	0	2	6	0	0	3
DAY4	0	1	0	1	0	1
DAY5	0	1	5	4	3	4
DAY6	0	0	1	3	1	2
DAY7	0	0	2	2	2	1

## TREATMENT:1% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	3	2	2	3	0	4
DAY2	1	1	3	3	0	2
DAY3	4	2	2	3	0	1
DAY4	2	2	3	1	0	2
DAY5	4	2	2	1	0	4
DAY6	2	2	0	1	0	1
DAY7	2	1	2	2	0	2

## TREATMENT:3.2% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	3	0	1	0
DAY2	3	3	3	0	0	3
DAY3	1	2	3	0	6	1
DAY4	1	0	0	1	1	1
DAY5	1	1	0	0	3	2
DAY6	0	1	2	0	1	2
DAY7	0	1	0	0	2	3

## TREATMENT:10% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	1	0	0	0
DAY2	1	3	2	2	0	0
DAY3	1	2	3	7	0	0
DAY4	1	1	1	0	0	0
DAY5	0	1	5	0	0	2
DAY6	1	1	2	0	2	0
DAY7	2	4	2	3	2	1

## TREATMENT:32% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	2	1	1	0	0
DAY2	1	0	0	0	0	0
DAY3	0	0	0	0	0	0
DAY4	0	0	0	0	0	1
DAY5	0	2	0	0	0	0
DAY6	1	1	0	1	0	0
DAY7	1	0	0	1	0	1

## TREATMENT:100% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	0	0	0	0
DAY2	0	0	0	0	0	0
DAY3	0	0	0	0	0	0
DAY4	0	0	0	0	0	0
DAY5	0	0	0	0	0	0
DAY6	0	0	0	0	0	0
DAY7	0	0	0	0	0	0

TRIAL #5 COMMENCED 11/3/91 PAGE 1  
ENDPOINT: DAILY TOTAL EGG PRODUCTION  
SPECIES: GLYPTOPHYSA SP.

# RAW DATA

## TREATMENT :CONTROL

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	49	0	29	54
DAY2	0	26	20	0	0	30
DAY3	0	16	55	0	0	30
DAY4	0	0	0	8	0	7
DAY5	0	5	39	25	27	39
DAY6	0	0	10	26	10	16
DAY7	0	0	15	5	12	9

## TREATMENT :1% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	24	23	19	24	0	18
DAY2	3	12	36	24	0	16
DAY3	35	26	3	16	0	18
DAY4	8	24	25	6	0	15
DAY5	32	24	14	4	0	39
DAY6	5	19	0	5	0	6
DAY7	22	10	15	13	0	16

## TREATMENT :3.2% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	29	0	4	0
DAY2	14	19	27	0	0	15
DAY3	4	16	31	0	45	7
DAY4	8	0	0	3	0	9
DAY5	9	4	0	0	27	19
DAY6	0	7	15	0	12	14
DAY7	0	7	0	0	15	27

## TREATMENT :10% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	13	0	0	0
DAY2	6	10	17	18	0	0
DAY3	5	12	33	69	0	0
DAY4	7	6	9	0	0	0
DAY5	0	0	51	0	0	7
DAY6	6	6	28	0	12	0
DAY7	11	17	16	21	11	4

## TREATMENT :32% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	21	7	13	0	0
DAY2	11	0	0	0	0	0
DAY3	0	0	0	0	0	0
DAY4	0	0	0	0	0	6
DAY5	0	13	0	0	0	0
DAY6	10	3	0	10	0	0
DAY7	12	0	0	9	0	9

## TREATMENT :100% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	0	0	0	0
DAY2	0	0	0	0	0	0
DAY3	0	0	0	0	0	0
DAY4	0	0	0	0	0	0
DAY5	0	0	0	0	0	0
DAY6	0	0	0	0	0	0
DAY7	0	0	0	0	0	0

## Appendix 6.b.

TRIAL #5 COMMENCE 31/3/91  
PHYSICO-CHEMICAL DATA

DISSOLVED OXYGEN mg/L

	TREATMENT											
	CON	CON	1XRP2	1XRP2	3.2XRP2	3.2XRP2	10XRP2	10XRP2	32XRP2	32XRP2	100XRP2	100XRP2
DAY1	7.5	7.2	7.5	7.4	7.4	7.5	7.4	7.2	7.5	7.3	7.3	7.4
DAY2	7.7	7.5	7.7	7.4	7.5	7.5	7.5	7.5	7.5	7.6	7.6	7.4
DAY3	7.4	7.4	7.5	7.4	7.3	7.3	7.4	7.4	7.4	7.4	7.3	7.4
DAY4	7.5	7.5	7.4	7.3	7.3	7.6	7.3	7.3	7.5	7.4	7.1	7.4
DAY5	7.2	7.3	7.4	7.3	7.2	7.4	7.4	7.5	7.5	7.3	7.4	7.6
DAY6	7.9	7.2	7.8	7.5	7.8	7.9	7.7	7.9	7.8	7.8	7.5	7.9
pH	6.3	6.35	6.45	6.75	6.65	6.78	6.74	6.84	6.76	6.81	6.9	7.4
DAY1	6.28	6.75	6.66	6.88	6.75	6.81	6.71	6.92	6.81	7.05	7.13	7.43
DAY2	6.35	6.72	6.66	6.63	6.48	6.65	6.67	6.72	6.79	6.99	6.97	6.96
DAY3	6.48	6.84	6.64	6.64	6.57	6.68	6.7	6.96	6.9	7.03	7.04	7.3
DAY4	6.51	6.61	6.59	6.7	6.6	6.71	6.74	6.94	6.85	7.11	7.12	7.34
DAY5	6.25	6.48	6.49	6.76	6.64	6.86	6.57	7.09	6.93	7.15	7.28	7.48

CONDUCTIVITY uS/cm

	TREATMENT											
	CON	CON	1XRP2	1XRP2	3.2XRP2	3.2XRP2	10XRP2	10XRP2	32XRP2	32XRP2	100XRP2	100XRP2
DAY1	29.2	22.1	44.7	42.1	75.3	74.8	181	187	464	465	1180	1180
DAY2	29.3	27.3	50.4	43	77.2	75.9	186	184	464	466	1190	1190
DAY3	32.6	26.6	39.7	40.1	77.4	77.3	189	183	460	472	1180	1180
DAY4	46.4	27.8	42	42	85	79.9	191	186	469	474	1190	1190
DAY5	38.5	26.2	44.1	44.1	82.8	80.6	183	180	458	462	1170	1170
DAY6	33.5	31.7	46.5	46.5	94.2	88.4	191	191	474	480	1190	1190

## Appendix 7.a.1.

TRIAL #6 COMMENCED 12/4/91 PAGE 1  
ENDPOINT: DAILY EGG MASS PRODUCTION  
SPECIES: A. carinata

RAW DATA

TREATMENT :CONTROL						
SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	3	0	2	4	1
DAY2	0	1	0	0	0	2
DAY3	0	0	5	1	2	2
DAY4	1	1	0	2	0	1
DAY5	1	1	1	2	2	2
TREATMENT :1% RP2						
SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	1	0	1	0	0	0
DAY2	0	0	1	1	0	0
DAY3	1	0	0	0	1	1
DAY4	1	0	1	0	1	1
DAY5	2	0	2	0	0	3
TREATMENT :3.2% RP2						
SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	1	2	0	5	0	1
DAY2	0	1	0	0	0	0
DAY3	1	0	0	0	0	0
DAY4	2	2	1	2	0	3
DAY5	2	1	0	0	0	0
TREATMENT :10% RP2						
SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	2	0	1	1	0
DAY2	0	1	0	0	0	0
DAY3	0	0	1	0	0	0
DAY4	0	1	0	0	1	1
DAY5	0	0	1	0	1	0
TREATMENT :32% RP2						
SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	0	0	0	0
DAY2	0	0	0	0	0	0
DAY3	0	0	0	0	0	0
DAY4	0	0	0	1	0	0
DAY5	0	0	0	0	0	0
TREATMENT :100% RP2						
SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	0	0	0	0
DAY2	0	0	0	0	0	0
DAY3	0	0	0	0	0	0
DAY4	0	0	0	0	0	2
DAY5	0	0	0	0	0	0

TRIAL #6 COMMENCED 12/4/91 PAGE 1  
ENDPOINT: DAILY TOTAL EGG PRODUCTION  
SPECIES: A. carinata

RAW DATA

TREATMENT :CONTROL						
SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	46	0	12	41	7
DAY2	0	17	0	0	0	16
DAY3	0	0	47	6	24	13
DAY4	0	8	0	12	0	9
DAY5	2	17	11	11	21	16
TREATMENT :1% RP2						
SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	2	0	11	0	0	0
DAY2	0	0	10	6	0	0
DAY3	7	0	0	0	12	8
DAY4	9	0	9	0	15	7
DAY5	17	0	20	0	0	15
TREATMENT :3.2% RP2						
SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	11	14	0	7	0	10
DAY2	0	7	0	0	0	0
DAY3	22	9	0	0	0	0
DAY4	8	18	23	10	0	21
DAY5	18	9	0	0	0	0
TREATMENT :10% RP2						
SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	18	0	14	2	0
DAY2	0	10	0	0	0	0
DAY3	0	0	10	0	0	0
DAY4	0	13	0	0	15	3
DAY5	0	0	13	0	0	0
TREATMENT :32% RP2						
SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	0	0	0	0
DAY2	0	0	0	0	0	0
DAY3	0	0	0	0	0	0
DAY4	0	0	0	0	0	0
DAY5	0	0	0	0	0	0
TREATMENT :100% RP2						
SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	0	0	0	0
DAY2	0	0	0	0	0	0
DAY3	0	0	0	0	0	4
DAY4	0	0	0	0	0	4
DAY5	0	0	0	0	0	0

## Appendix 7.a.2.

TRIAL #6 COMMENCED 12/4/91 PAGE 1  
ENDPOINT: DAILY EGG MASS PRODUCTION  
SPECIES: A. cummingii

RAW DATA

TREATMENT :CONTROL						
SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	1	2	0	0	1	1
DAY2	0	0	0	2	1	0
DAY3	0	1	0	1	3	0
DAY4	2	2	2	2	1	2
DAY5	0	0	0	2	1	2
TREATMENT :1% RP2						
SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	0	0	0	0
DAY2	2	0	2	2	1	1
DAY3	2	2	0	1	2	2
DAY4	0	1	1	2	2	2
DAY5	0	1	1	2	1	2
TREATMENT :3.2% RP2						
SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	0	0	0	0
DAY2	0	1	0	1	1	1
DAY3	0	0	2	0	0	0
DAY4	1	2	0	2	2	1
DAY5	0	1	2	1	1	1
TREATMENT :10% RP2						
SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	0	0	0	0
DAY2	0	2	0	0	0	0
DAY3	1	0	0	0	0	0
DAY4	0	1	0	1	0	0
DAY5	1	0	1	0	1	0
TREATMENT :32% RP2						
SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	0	0	0	0
DAY2	0	0	0	0	0	0
DAY3	1	0	0	0	0	0
DAY4	0	0	1	0	0	0
DAY5	0	1	0	0	0	1
TREATMENT :100% RP2						
SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	0	0	0	0
DAY2	0	2	0	0	0	0
DAY3	0	0	0	1	0	0
DAY4	0	0	0	0	0	0
DAY5	0	1	0	0	0	0

TRIAL #6 COMMENCED 12/4/91 PAGE 1  
ENDPOINT: DAILY TOTAL EGG PRODUCTION  
SPECIES: A. cummingii

RAW DATA

TREATMENT :CONTROL						
SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	17	37	0	0	25	12
DAY2	0	0	0	45	29	0
DAY3	0	21	0	24	58	0
DAY4	30	46	29	38	18	25
DAY5	0	0	0	38	17	28
TREATMENT :1% RP2						
SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	0	0	0	0
DAY2	48	0	52	40	25	17
DAY3	48	55	0	17	51	34
DAY4	0	28	25	35	36	21
DAY5	0	28	15	37	22	34
TREATMENT :3.2% RP2						
SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	0	0	0	0
DAY2	0	19	0	23	21	17
DAY3	0	0	38	0	0	0
DAY4	11	44	0	35	37	6
DAY5	13	26	29	19	19	18
TREATMENT :10% RP2						
SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	0	0	0	0
DAY2	0	40	0	0	0	0
DAY3	14	0	0	0	0	0
DAY4	0	15	0	14	0	0
DAY5	7	0	21	0	14	0
TREATMENT :32% RP2						
SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	0	0	0	0
DAY2	0	0	0	0	0	0
DAY3	0	0	0	0	0	0
DAY4	13	0	3	0	0	0
DAY5	0	18	0	0	0	3
TREATMENT :100% RP2						
SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	0	0	0	0
DAY2	0	0	0	0	0	0
DAY3	0	0	0	15	0	0
DAY4	0	0	0	0	0	0
DAY5	0	14	0	0	0	0

## Appendix 7.a.3.

TRIAL #6 COMMENCED 12/4/91 PAGE 1  
ENDPOINT: DAILY EGG MASS PRODUCTION  
SPECIES: GLYPTOPHYSA SP.

## RAW DATA

TREATMENT : CONTROL						
SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	4	5	5	3	4	2
DAY2	5	1	0	4	4	2
DAY3	1	2	0	3	2	4
DAY4	0	1	1	1	0	3
DAY5	0	2	0	1	0	0

TREATMENT : 1% RP2						
SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	3	1	4	3	0	2
DAY2	1	2	3	2	0	3
DAY3	1	2	0	3	0	1
DAY4	2	2	0	2	0	1
DAY5	1	0	0	2	0	1

TREATMENT : 3.2% RP2						
SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	4	5	1	2	6	0
DAY2	4	4	1	1	4	1
DAY3	2	2	0	1	1	1
DAY4	0	0	0	0	0	0
DAY5	0	0	0	1	0	2

TREATMENT : 10% RP2						
SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	1	2	2	0	0	0
DAY2	3	4	3	4	2	3
DAY3	0	1	1	0	0	0
DAY4	1	2	2	3	3	1
DAY5	0	0	0	2	1	1

TREATMENT : 32% RP2						
SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	0	0	0	0
DAY2	0	0	2	0	0	0
DAY3	0	0	0	0	0	0
DAY4	0	0	1	0	0	0
DAY5	0	0	1	0	0	0

TREATMENT : 100% RP2						
SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	0	0	0	0
DAY2	0	0	0	0	0	0
DAY3	0	0	0	0	3	0
DAY4	1	0	0	0	2	0
DAY5	1	0	0	0	1	0

TRIAL #6 COMMENCED 12/4/91 PAGE 2  
ENDPOINT: DAILY TOTAL EGG PRODUCTION  
SPECIES: GLYPTOPHYSA SP.

## RAW DATA

TREATMENT: 10% RP2						
SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	14	31	9	0	0	0
DAY2	32	39	15	51	15	34
DAY3	0	7	6	0	0	0
DAY4	5	14	9	39	27	10
DAY5	0	0	0	22	11	10

TREATMENT: 32% RP2						
SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	0	0	0	0
DAY2	0	0	27	0	0	0
DAY3	0	0	0	0	0	0
DAY4	0	0	11	0	0	0
DAY5	0	0	13	0	0	0

TREATMENT: 100% RP2						
SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	0	0	0	0
DAY2	0	0	0	0	0	0
DAY3	0	0	0	0	34	0
DAY4	7	0	0	0	11	0
DAY5	3	0	0	0	8	0

TREATMENT: CONTROL						
SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	34	50	44	41	0	11
DAY2	46	6	0	41	36	13
DAY3	6	16	0	33	13	26
DAY4	0	0	4	10	0	22
DAY5	15	18	0	12	0	0

TREATMENT: 1% RP2						
SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	38	12	45	26	0	15
DAY2	14	22	28	21	0	27
DAY3	12	19	0	27	0	10
DAY4	26	16	0	12	0	7
DAY5	0	0	0	24	0	10

TREATMENT: 3.2% RP2						
SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	40	57	11	22	66	0
DAY2	35	42	9	8	37	5
DAY3	16	21	0	8	11	7
DAY4	0	0	0	0	0	0
DAY5	0	0	0	0	6	12

## Appendix 7.b.1.

TRIAL #6 VALIDATION COMMENCED 12/4/91  
ENDPOINT: DEVELOPMENTAL CHARACTERISTICS  
EGG NUMBERS  
SPECIES: A. carinata

TREATMENT : CONTROL						
GASTRULA	TROCH	REPLICATE 1A DEVELOPMENTAL STAGE				
		VEL	HIPPO	H'LING	N'NATE	
DAY1	18 16 12					

DAY2	16 12	18				
DAY3		18 16 12				

DAY4			18 16 12			
DAY5				18 16 12		

DAY6				18 16 12		
DAY7				18 16		12
DAY8					18 16	

TREATMENT : CONTROL						
GASTRULA	TROCH	REPLICATE 1B DEVELOPMENTAL STAGE				
		VEL	HIPPO	H'LING	N'NATE	
DAY1	11 13 9 8					

DAY2	11	13 9				
DAY3	8		11		13	

DAY4			9 8		11 13	
DAY5				9 8	11 13 9 8	

DAY6					11 13 9 8	
DAY7						11 13 9 8

TREATMENT : CONTROL						
GASTRULA	TROCH	REPLICATE 2A DEVELOPMENTAL STAGE				
		VEL	HIPPO	H'LING	N'NATE	
DAY1	11 10					

DAY2	11 10					
DAY3		11 10				

DAY4			11 10			
DAY5				11 10		
DAY6					11 10	

TREATMENT : CONTROL						
GASTRULA	TROCH	REPLICATE 2B DEVELOPMENTAL STAGE				
		VEL	HIPPO	H'LING	N'NATE	
DAY1	7 9					

DAY2	9	7				
DAY3			9		7	

DAY4				9	7	
DAY5					7 ?	
DAY6					7 ?	
DAY7						7 ?

TREATMENT : CONTROL		REPLICATE 2C				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	11 12 10 8					
DAY2		11 12 10 3				
DAY3				11 12 10 8		
DAY4					11 12 10 8	
DAY5					11 12 10 8	
DAY6					11 12 10 8	
DAY7						11 12 10 8

TREATMENT : 1% RP2		REPLICATE 1A				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	10 7					
DAY2	10 7					
DAY3			10 7			
DAY4				10 7		
DAY5					10 7	
DAY6					10 7	
DAY7					10 7	
DAY8						10 7

TREATMENT : 1% RP2		REPLICATE 1B				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	11					
DAY2	11					
DAY3		11				
DAY4				11		
DAY5					11	
DAY6					11	
DAY7					11	
DAY8						11

TREATMENT : 1% RP2		REPLICATE 2A				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	6					
DAY2		6				
DAY3				6		
DAY4					6	
DAY5					6	
DAY6					6	
DAY7					6	
DAY8						6

TREATMENT : 1% RP2		REPLICATE 2B				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	12					
DAY2	12					
DAY3			12			
DAY4				12		
DAY5					12	
DAY6					12	
DAY7					12	
DAY8					10	
DAY9						10

TREATMENT : 1% RP2		REPLICATE 2C				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	9					
DAY2		8				
DAY3				8		
DAY4					8	
DAY5						8
DAY6						8
DAY7						8
DAY8						8
DAY9						8

TREATMENT : 3.2% RP2		REPLICATE 1A				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	11 11					
DAY2	11	11				
DAY3		11		11		
DAY4					11 11	
DAY5						11 11
DAY6						11 11
DAY7						11 11
DAY8						11 11
DAY9						11 11

TREATMENT : 3.2% RP2		REPLICATE 1B				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	8 10					
DAY2	8 9					
DAY3				8 9		
DAY4					8 9	
DAY5						8 9
DAY6						8 9
DAY7						8 9
DAY8						8 9

TREATMENT : 3.2% RP2		REPLICATE 2A				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	7					
DAY2	7					
DAY3		7				
DAY4					7	
DAY5						7
DAY6						7
DAY7						7
DAY8						7
DAY9						7

TREATMENT : 3.2% RP2		REPLICATE 2B				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE		H'LING	N'NATE
			VEL	HIPPO		
DAY1	10					
DAY2		10				
DAY3		10				
DAY4					10	
DAY5						10
DAY6						10
DAY7						10
DAY8						10

## Appendix 7.b.2.

TREATMENT : 10% RP2		REPLICATE 1A				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE			N'NATE
			VEL	HIPPO	H'LING	
DAY1	8 10					
DAY2		8 10				
DAY3		8 10				
DAY4				8 10		
DAY5					8 10	
DAY6					8 10	
DAY7						8 10

TREATMENT : 10% RP2		REPLICATE 1B				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE			N'NATE
			VEL	HIPPO	H'LING	
DAY1	13					
DAY2		13				
DAY3				13		
DAY4					13	
DAY5					13	
DAY6					13	
DAY7					13	
DAY8						13

TREATMENT : 10% RP2		REPLICATE A				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE			N'NATE
			VEL	HIPPO	H'LING	
DAY1	15					
DAY2	15					
DAY3		15				
DAY4			15			
DAY5					14	
DAY6					14	
DAY7					14	
DAY8					14	
DAY9						14

TREATMENT : 100% RP2		REPLICATE 1A				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE			N'NATE
			VEL	HIPPO	H'LING	
DAY1	4					
DAY2	4					
DAY3		4				
DAY4				4		
DAY5					3	
DAY6					3	
DAY7						DEAD

TRIAL #6 VALIDATION COMMENCED 12/4/91  
 ENDPOINT DEVELOPMENTAL CHARACTERISTICS PAGE 1  
 EGG NUMBERS RAW DATA  
 SPECIES A. cummingii

TREATMENT: CONTROL		REPLICATE 1A				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE			N'NATE
			VEL	HIPPO	H'LING	
DAY1	17					
DAY2	17					
DAY3		17				
DAY4					17	
DAY5						17
DAY6						17
DAY7						17
DAY8						17

TREATMENT: CONTROL		REPLICATE 1B				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE			N'NATE
			VEL	HIPPO	H'LING	
DAY1	15 22					
DAY2	14 22					
DAY3		14 22				
DAY4				13	22	
DAY5						13 22
DAY6						13 22
DAY7						13 22
DAY8						13 22

TREATMENT: CONTROL		REPLICATE 1C				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE			N'NATE
			VEL	HIPPO	H'LING	
DAY1	17					
DAY2	17					
DAY3		17				
DAY4				17		
DAY5					17	
DAY6						17
DAY7						17
DAY8						17
DAY9						17

TREATMENT: CONTROL		REPLICATE 2A				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE			N'NATE
			VEL	HIPPO	H'LING	
DAY1	24 21					
DAY2	24					
DAY3		21				
DAY4				20 21		
DAY5					17 19	
DAY6						17 19
DAY7						17 19

TREATMENT: CONTROL		REPLICATE 2B				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE			N'NATE
			VEL	HIPPO	H'LING	
DAY1	25					
DAY2	25					
DAY3		25				
DAY4				25		
DAY5					25	
DAY6					25	
DAY7					25	
DAY8						25

TREATMENT: CONTROL		REPLICATE 2C				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE VEL HIPPO	H'LING	N'NATE	
DAY1	12 16					
DAY2	12 15					
DAY3		12 16				
DAY4			12 16			
DAY5				12 16		
DAY6					12 16	
DAY7					12 16	
DAY8					12 16	
DAY9						12 15

TREATMENT: 1% RP2		REPLICATE 1A				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE VEL HIPPO	H'LING	N'NATE	
DAY1	23 24					
DAY2	23 24					
DAY3		23 24				
DAY4			23 24			
DAY5				23 23		
DAY6				23 23		
DAY7					23 23	

TREATMENT: 1% RP2		REPLICATE 1B				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE VEL HIPPO	H'LING	N'NATE	
DAY1	30 25					
DAY2	30 25					
DAY3		29 25				
DAY4				25	29	
DAY5					29 23	
DAY6					29 23	
DAY7						29 21

TREATMENT: 1% RP2		REPLICATE 1C				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE VEL HIPPO	H'LING	N'NATE	
DAY1	28 23					
DAY2	28 23					
DAY3		27 22				
DAY4			22	27		
DAY5					27 22	
DAY6					27 22	
DAY7					26 22	
DAY8						26 22

TREATMENT: 1% RP2		REPLICATE 2A				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE VEL HIPPO	H'LING	N'NATE	
DAY1	20 20					
DAY2	20 20					
DAY3			20 19			
DAY4				20 18		
DAY5					20 18	
DAY6					20 18	
DAY7					20 18	
DAY8						20 18

TREATMENT: 1% RP2		REPLICATE 1B				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE VEL HIPPO	H'LING	N'NATE	
DAY1	27 24					
DAY2		27 24				
DAY3				24 24		
DAY4					21 23	
DAY5					21 23	
DAY6					21 23	
DAY7					21 23	
DAY8						21 23

TREATMENT: 1% RP2		REPLICATE 2C				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE VEL HIPPO	H'LING	N'NATE	
DAY1	19 15					
DAY2		19				
DAY3		19 15				
DAY4				19 15		
DAY5					19 15	
DAY6					19 15	
DAY7					19 15	
DAY8					19 15	
DAY9					19 15	
DAY10					FORCED HATCH	19 15

TREATMENT: 3.2% RP2		REPLICATE 1A				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE VEL HIPPO	H'LING	N'NATE	
DAY1	13					
DAY2	13					
DAY3		13				
DAY4			12			
DAY5				12		
DAY6					12	
DAY7					12	
DAY8						12

TREATMENT: 3.2% RP2		REPLICATE 1B				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE VEL HIPPO	H'LING	N'NATE	
DAY1	25 19					
DAY2	25 19					
DAY3			24 19			
DAY4				19	24	
DAY5					24 19	
DAY6					24 19	
DAY7					24 19	
DAY8					24 19	
DAY9						24 19



TREATMENT: 3.2% RP2		REPLICATE 1C				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE VEL HIPPO	H'LING	N'HATE	
DAY1	27 26					
DAY2		27 26				
DAY3			24 24			
DAY4				21 23		
DAY5				21 23		
DAY6				21 23		
DAY7				21 23		
DAY8					21 23	

TREATMENT: 3.2% RP2		REPLICATE 2A				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE VEL HIPPO	H'LING	N'HATE	
DAY1	23					
DAY2	21					
DAY3		21				
DAY4			28			
DAY5				18		
DAY6				18		
DAY7				18		
DAY8				18		
DAY9					18	
DAY10						

TREATMENT: 3.2% RP2		REPLICATE 2B				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE VEL HIPPO	H'LING	N'HATE	
DAY1	21					
DAY2	20					
DAY3		19				
DAY4			19			
DAY5				19		
DAY6				19		
DAY7				19		
DAY8					19	

TREATMENT: 3.2% RP2		REPLICATE 2C				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE VEL HIPPO	H'LING	N'HATE	
DAY1	17					
DAY2	17					
DAY3			16			
DAY4				15		
DAY5				15		
DAY6				15		
DAY7				15		
DAY8				15		
DAY9					15	

TREATMENT: 10% RP2		REPLICATE 1A				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE VEL HIPPO	H'LING	N'HATE	
DAY1	14					
DAY2	14					
DAY3		13				
DAY4			11			
DAY5				11		
DAY6				11		
DAY7				11		
DAY8				11		
DAY9				11		
DAY10					11	

TREATMENT: 10% RP2		REPLICATE 1B				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE VEL HIPPO	H'LING	N'HATE	
DAY1	23 17					
DAY2	23 17					
DAY3		23 17				
DAY4			23 16			
DAY5				23 16		
DAY6					23 16	
DAY7					23 16	
DAY8					23 16	
DAY9					23 16	
DAY10						23 16

TREATMENT: 10% RP2		REPLICATE 1C				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE VEL HIPPO	H'LING	N'HATE	
DAY1	21					
DAY2	20					
DAY3		20				
DAY4				20		
DAY5					20	
DAY6					20	
DAY7					20	
DAY8						20

TREATMENT: 10% RP2		REPLICATE 2A				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE VEL HIPPO	H'LING	N'HATE	
DAY1	14					
DAY2	14					
DAY3		14				
DAY4			14			
DAY5				13		
DAY6					13	
DAY7					13	
DAY8					13	
DAY9						13

TREATMENT: 10% RP2		REPLICATE 2B				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE VEL HIPPO	H'LING	N'HATE	
DAY1	14					
DAY2	14					
DAY3		14				
DAY4			14			
DAY5				13		
DAY6					13	
DAY7					13	
DAY8					13	
DAY9					13	
DAY10						13

TREATMENT: 32% RP2		REPLICATE 1A				
	GASTRULA	TROCH	DEVELOPMENTAL STAGE VEL HIPPO	H'LING	N'HATE	
DAY1	13					
DAY2	13					
DAY3			11			
DAY4				11		
DAY5					11	
DAY6					11	
DAY7					11	
DAY8					11	
DAY9					11	
DAY10						11

TREATMENT: 32% RP2

REPLICATE 2B

	GASTRULA	DEVELOPMENTAL STAGE				H'LING	N'NATE
		TROCH	VEL	HIPPO			
DAY1	18						
DAY2	18						
DAY3		18					
DAY4			17				
DAY5				17			
DAY6				17			
DAY7				17			
DAY8				17			
DAY9					17		

TREATMENT: 32% RP2

REPLICATE 2A

	GASTRULA	DEVELOPMENTAL STAGE				H'LING	N'NATE
		TROCH	VEL	HIPPO			
DAY1	9						
DAY2	9						
DAY3		9					
DAY4				9			
DAY5				9			
DAY6					9		
DAY7					9		
DAY8					9		
DAY9					9		
DAY10						9	

TREATMENT: 100% RP2

REPLICATE 1A

	GASTRULA	DEVELOPMENTAL STAGE				H'LING	N'NATE
		TROCH	VEL	HIPPO			
DAY1	15						
DAY2	15						
DAY3		15					
DAY4			15				
DAY5					15		
DAY6					15		
DAY7					15		
DAY8					15		
DAY9					15		
DAY10						15	

## Appendix 7.b.3.

TRIAL #6 VALIDATION COMMENCED 12/4/91  
 ENDPOINT DEVELOPMENTAL CHARACTERISTICS PAGE 1  
 EGG NUMBERS RAW DATA  
 SPECIES GLYPTOPHYSA SP.

TREATMENT: CONTROL

REPLICATE 1A

	GASTRULA	DEVELOPMENTAL STAGE				H'LING	N'NATE
		TROCH	VEL	HIPPO			
DAY1	6 7						
DAY2	6 5						
DAY3		4 3					
DAY4				2 3			
DAY5					2 3		
DAY6						2 3	

TREATMENT: CONTROL

REPLICATE 1B

	GASTRULA	DEVELOPMENTAL STAGE				H'LING	N'NATE
		TROCH	VEL	HIPPO			
DAY1	10 8						
DAY2	8 8						
DAY3			8 7				
DAY4					8 7		
DAY5					7 7		
DAY6						7 7	

TREATMENT: CONTROL

REPLICATE 1C

	GASTRULA	DEVELOPMENTAL STAGE				H'LING	N'NATE
		TROCH	VEL	HIPPO			
DAY1	9 11 10						
DAY2	8 7 6						
DAY3		4 4 6					
DAY4				3 4 6			
DAY5					2 3 6		
DAY6					2 2 6		
DAY7						2 2 5	

TREATMENT: CONTROL

REPLICATE 2A

	GASTRULA	DEVELOPMENTAL STAGE				H'LING	N'NATE
		TROCH	VEL	HIPPO			
DAY1	13 14 14						
DAY2		12 14 11					
DAY3				10 12 11			
DAY4					10 11 8		
DAY5					10 11 8		
DAY6					11 8	10	
DAY7						11 8	

TREATMENT: CONTROL.		REPLICATE 2B				
	GASTRULA	DEVELOPMENTAL STAGE				N'NATE
		TROCH	VEL	HIPPO	H'LING	
DAY1	11 11					
DAY2	11 11					
DAY3			11 9			
DAY4				11 9		
DAY5					11 9	
DAY6					11 8	
DAY7						11 8

TREATMENT: CONTROL.		REPLICATE 2C				
	GASTRULA	DEVELOPMENTAL STAGE				N'NATE
		TROCH	VEL	HIPPO	H'LING	
DAY1	8 5 9					
DAY2	8 5 9					
DAY3		3 4 9				
DAY4			5 4			
DAY5				4		
DAY6					5 4 4	
DAY7					5 3 2	
DAY8						5 3 2

TREATMENT: 1% RP2		REPLICATE 1A				
	GASTRULA	DEVELOPMENTAL STAGE				N'NATE
		TROCH	VEL	HIPPO	H'LING	
DAY1	13 13 12					
DAY2	7 13 12					
DAY3		5 11 9				
DAY4				4 10 7		
DAY5					2 10 5	
DAY6					2 10 5	
DAY7						2 10 5

TREATMENT: 1% RP2		REPLICATE 1B				
	GASTRULA	DEVELOPMENTAL STAGE				N'NATE
		TROCH	VEL	HIPPO	H'LING	
DAY1	10 11 13					
DAY2	10 11 13					
DAY3		3 11 11				
DAY4				3 11 11		
DAY5					3 10 11	
DAY6					3 10 11	
DAY7						3 10 11

TREATMENT: 1% RP2		REPLICATE 2A				
	GASTRULA	DEVELOPMENTAL STAGE				N'NATE
		TROCH	VEL	HIPPO	H'LING	
DAY1	10 13					
DAY2	10 13					
DAY3		10				
DAY4				10 13		
DAY5					10 13	
DAY6					9 13	
DAY7					9 13	
DAY8						9 10

TREATMENT: 1% RP2		REPLICATE 2B				
	GASTRULA	DEVELOPMENTAL STAGE				N'NATE
		TROCH	VEL	HIPPO	H'LING	
DAY1	8 9 10					
DAY2	8 9 10					
DAY3		3 9				
DAY4				3 9 10		
DAY5					3 9 10	
DAY6					3 9 10	
DAY7						3 9 10

TREATMENT: 3.2 % RP2		REPLICATE 1A				
	GASTRULA	DEVELOPMENTAL STAGE				N'NATE
		TROCH	VEL	HIPPO	H'LING	
DAY1	10 10					
DAY2	10 10					
DAY3		10 3				
DAY4				10 3		
DAY5					10 2	
DAY6						10 2

TREATMENT: 3.2 % RP2		REPLICATE 1B				
	GASTRULA	DEVELOPMENTAL STAGE				N'NATE
		TROCH	VEL	HIPPO	H'LING	
DAY1	11					
DAY2	11					
DAY3		10				
DAY4				10		
DAY5					10	
DAY6					10	
DAY7						10

TREATMENT: 3.2 % RP2		REPLICATE 1C				
	GASTRULA	DEVELOPMENTAL STAGE				N'NATE
		TROCH	VEL	HIPPO	H'LING	
DAY1	11					
DAY2		2				
DAY3		DEAD				

TREATMENT: 3.2 % RP2		REPLICATE 2A				
	GASTRULA	DEVELOPMENTAL STAGE				N'NATE
		TROCH	VEL	HIPPO	H'LING	
DAY1	8					
DAY2	8					
DAY3		8				
DAY4				8		
DAY5					8	
DAY6					8	
DAY7						8

TREATMENT: 3.2 % RP2

REPLICATE 2B

	GASTRULA	DEVELOPMENTAL STAGE		H'LING	N'NATE
		TROCH	VEL HIPPO		
DAY1	14 14 13				
DAY2	14 14 13				
DAY3		13 14 8			
DAY4			13 14 8		
DAY5				13 14 8	
DAY6				13 14 8	
DAY7					13 14 8

TREATMENT: 3.2 % RP2

REPLICATE 2C

	GASTRULA	DEVELOPMENTAL STAGE		H'LING	N'NATE
		TROCH	VEL HIPPO		
DAY1	8 7				
DAY2			8 7		
DAY3				8 7	
DAY4				8 7	
DAY5				8 7	
DAY6					8 7

TREATMENT: 10% RP2

REPLICATE 1A

	GASTRULA	DEVELOPMENTAL STAGE		H'LING	N'NATE
		TROCH	VEL HIPPO		
DAY1	14				
DAY2		14			
DAY3			14		
DAY4				14	
DAY5				14	
DAY6				14	
DAY7					14

TREATMENT: 10% RP2

REPLICATE 1B

	GASTRULA	DEVELOPMENTAL STAGE		H'LING	N'NATE
		TROCH	VEL HIPPO		
DAY1	3 6				
DAY2	3 6				
DAY3	3	6			
DAY4			3		
DAY5				2	
DAY6				2 6	
DAY7					2 6

TREATMENT: 10% RP2

REPLICATE 2A

	GASTRULA	DEVELOPMENTAL STAGE		H'LING	N'NATE
		TROCH	VEL HIPPO		
DAY1	12 5 10				
DAY2	10 5 10				
DAY3	10	3	10		
DAY4			8 3		
DAY5				8 2 8	
DAY6				8 2 8	
DAY7					8 2 3

TREATMENT: 10% RP2

REPLICATE 2B

	GASTRULA	DEVELOPMENTAL STAGE		H'LING	N'NATE
		TROCH	VEL HIPPO		
DAY1	11 12 11				
DAY2	11 12 11				
DAY3			11 12 11		
DAY4				11 12 10	
DAY5					11 12 10
DAY6					11 12 10
DAY7					11 12 10

TREATMENT: 32% RP2

REPLICATE 1A

	GASTRULA	DEVELOPMENTAL STAGE		H'LING	N'NATE
		TROCH	VEL HIPPO		
DAY1	13 14				
DAY2	12 13				
DAY3		12 13			
DAY4			12 13		
DAY5				12 13	
DAY6				11 13	
DAY7				11 13	
DAY8					11 13

TREATMENT: 100% RP2

REPLICATE 1A

	GASTRULA	DEVELOPMENTAL STAGE		H'LING	N'NATE
		TROCH	VEL HIPPO		
DAY1	7				
DAY2		7			
DAY3			7		
DAY4				7	
DAY5				7	
DAY6				7	
DAY7				7	
DAY8					7

TREATMENT: 100% RP2

REPLICATE 1B

	GASTRULA	DEVELOPMENTAL STAGE		H'LING	N'NATE
		TROCH	VEL HIPPO		
DAY1	13 13				
DAY2	13 13				
DAY3			12 13		
DAY4				11 13	
DAY5				11 13	
DAY6				11 13	
DAY7				11 13	
DAY8					11 13

## Appendix 7.c.1.

TRIAL #6 VALIDATION COMMENCED 12/4/91 PAGE 1  
ENDPOINT: WEIGHT LOSS IN EXPOSED ADULTS IN MILLIGRAMS  
SPECIES *A. cummingsi*

## TREATMENT: CONTROL

	BEFORE	MARKED		BEFORE	UNMARKED	
		AFTER	%LOSS		AFTER	%LOSS
REP 1A	256	250	2.3	111	116	11.5
REP 1B	231	213	7.8	144	162	12.5
REP 1C	125	128	-2.4	111	112	-0.9
REP 2A	237	243	-2.5	121	121	0
REP 2B	191	139	27.2	127	141	-11
REP 2C	100	114	-14	113	114	-0.8

## TREATMENT: 1% RP2

	BEFORE	MARKED		BEFORE	UNMARKED	
		AFTER	%LOSS		AFTER	%LOSS
REP 1A	141	145	-2.8	172	117	4.1
REP 1B	118	197	-66.9	139	162	-16.5
REP 1C	106	101	4.7	101	98	3
REP 2A	243	259	-6.5	112	114	-1.8
REP 2B	151	151	0	125	123	1.6
REP 2C	110	102	7.3	91	84	7.7

## TREATMENT: 3.2% RP2

	BEFORE	MARKED		BEFORE	UNMARKED	
		AFTER	%LOSS		AFTER	%LOSS
REP 1A	725	221	2.2	109	103	5.5
REP 1B	148	149	-0.7	130	109	16.2
REP 1C	204	191	6.4	116	109	6
REP 2A	191	188	1.6	118	115	2.5
REP 2B	132	131	0.8	122	136	-11.5
REP 2C	114	97	14.9	93	88	5.4

## TREATMENT: 10% RP2

	BEFORE	MARKED		BEFORE	UNMARKED	
		AFTER	%LOSS		AFTER	%LOSS
REP 1A	187	167	10.7	125	130	4
REP 1B	139	116	16.5	153	201	-31.4
REP 1C	106	149	-40.6	101	158	-56.4
REP 2A	144	148	-2.8	96	95	1
REP 2B	123	131	-6.5	127	131	3.1
REP 2C	108	122	-13	97	102	-5.2

## TREATMENT: 32% RP2

	BEFORE	MARKED		BEFORE	UNMARKED	
		AFTER	%LOSS		AFTER	%LOSS
REP 1A	179	203	-13.4	99	171	-77.2
REP 1B	175	190	-8.6	121	116	4.1
REP 1C	124	119	4	122	126	-3.3
REP 2A	258	286	-10.9	115	111	3.5
REP 2B	166	164	1.2	152	145	4.6
REP 2C	130	129	0.8	117	DEAD	

## TREATMENT: 100% RP2

	BEFORE	MARKED		BEFORE	UNMARKED	
		AFTER	%LOSS		AFTER	%LOSS
REP 1A	182	183	-0.5	124	130	4.8
REP 1B	168	167	0.6	139	155	-11.5
REP 1C	90	106	-17.8	141	149	-5.7
REP 2A	198	205	-3.5	101	114	-12.9
REP 2B	104	126	-21.2	124	135	-8.9
REP 2C	113	DEAD		93	109	-17.2

## Appendix 7.c.3.

TRIAL #6 VALIDATION COMMENCED 12/4/91 PAGE 1  
ENDPOINT: WEIGHT LOSS IN EXPOSED ADULTS IN MILLIGRAMS  
SPECIES *GLYPTOPHYSA* SP.

## TREATMENT: CONTROL

	BEFORE	MARKED		BEFORE	UNMARKED	
		AFTER	%LOSS		AFTER	%LOSS
REP 1A	353	314	11	375	331	11.3
REP 1B	316	297	6	364	329	9.6
REP 1C	333	305	8.4	357	315	11.8
REP 2A	383	350	8.6	421	402	4.5
REP 2B	294	284	3.4	342	310	9.4
REP 2C	316	298	5.7	254	253	3.9

## TREATMENT: 1% RP2

	BEFORE	MARKED		BEFORE	UNMARKED	
		AFTER	%LOSS		AFTER	%LOSS
REP 1A	386	377	2.3	337	314	6.8
REP 1B	340	321	5.6	368	341	7.3
REP 1C	330	319	3.3	318	296	6.9
REP 2A	424	397	6.4	273	297	-8.8
REP 2B	313	308	1.6	392	388	1
REP 2C	296	297	-0.3	286	258	9.8

## TREATMENT: 3.2% RP2

	BEFORE	MARKED		BEFORE	UNMARKED	
		AFTER	%LOSS		AFTER	%LOSS
REP 1A	327	302	7.6	320	308	3.8
REP 1B	410	403	1.7	356	349	2
REP 1C	319	292	8.5	307	309	0.7
REP 2A	344	321	6.7	383	369	3.7
REP 2B	367	367	0	426	421	1.2
REP 2C	298	315	-5.7	283	283	0

## TREATMENT: 10% RP2

	BEFORE	MARKED		BEFORE	UNMARKED	
		AFTER	%LOSS		AFTER	%LOSS
REP 1A	388	392	-1	334	320	4.2
REP 1B	385	373	3.1	378	376	0.5
REP 1C	305	295	3.3	296	286	0.7
REP 2A	391	382	2.3	350	368	-5.1
REP 2B	408	402	1.2	417	400	4.3
REP 2C	346	343	0.9	296	276	6.8

## TREATMENT: 32% RP2

	BEFORE	MARKED		BEFORE	UNMARKED	
		AFTER	%LOSS		AFTER	%LOSS
REP 1A	307	303	1.3	334	317	5
REP 1B	344	346	-0.6	379	373	1.6
REP 1C	337	329	2.4	350	367	0.9
REP 2A	401	408	-1.7	368	343	6.8
REP 2B	343	320	6.7	374	370	1.1
REP 2C	278	283	-1.8	279	269	3.7

## TREATMENT: 100% RP2

	BEFORE	MARKED		BEFORE	UNMARKED	
		AFTER	%LOSS		AFTER	%LOSS
REP 1A	363	347	4.4	390	387	0.8
REP 1B	383	381	0.5	354	358	-1.1
REP 1C	240	267	-11.3	308	326	-3.8
REP 2A	368	378	-2.7	292	311	-6.5
REP 2B	378	367	2.9	349	380	-8.9
REP 2C	240	250	-4.2	210	248	-11.2

## Appendix 7.c.2.

TRIAL #6 VALIDATION COMMENCED 12/4/91 PAGE 1  
ENDPOINT: WEIGHT LOSS IN EXPOSED ADULTS IN MILLIGRAMS  
SPECIES *A. cummingsi*

## TREATMENT: CONTROL

	BEFORE	MARKED		BEFORE	UNMARKED	
		AFTER	%LOSS		AFTER	%LOSS
REP 1A	292	261	10.6	303	271	7.3
REP 1B	221	230	-4.1	264	231	12.5
REP 1C	253	240	13	233	204	12.6
REP 2A	252	228	24	218	194	12.4
REP 2B	255	245	10	285	275	3.5
REP 2C	177	191	-14	200	200	0

## TREATMENT: 1% RP2

	BEFORE	MARKED		BEFORE	UNMARKED	
		AFTER	%LOSS		AFTER	%LOSS
REP 1A	263	247	6.1	284	292	-2.3
REP 1B	284	277	2.5	247	248	-0.4
REP 1C	162	186	-14.8	259	230	11.2
REP 2A	239	237	0.8	214	221	-3.2
REP 2B	238	234	1.7	265	263	0.8
REP 2C	201	189	6	223	214	4

## TREATMENT: 3.2% RP2

	BEFORE	MARKED		BEFORE	UNMARKED	
		AFTER	%LOSS		AFTER	%LOSS
REP 1A	198	186	6.1	242	230	5
REP 1B	229	200	12.7	258	265	-2.7
REP 1C	232	195	15.9	188	212	-12.8
REP 2A	240	253	-5.4	208	268	-28.8
REP 2B	241	221	8.3	248	247	0.4
REP 2C	199	210	-5.5	222	240	-8.1

## TREATMENT: 10% RP2

	BEFORE	MARKED		BEFORE	UNMARKED	
		AFTER	%LOSS		AFTER	%LOSS
REP 1A	251	258	-2.8	212	221	-4.2
REP 1B	274	256	6.6	256	268	-4.7
REP 1C	229	241	-5.2	203	203	0
REP 2A	277	278	-0.4	234	236	-0.9
REP 2B	234	237	-1.3	227	220	3.1
REP 2C	188	185	1.6	186	199	-7

## TREATMENT: 32% RP2

	BEFORE	MARKED		BEFORE	UNMARKED	
		AFTER	%LOSS		AFTER	%LOSS
REP 1A	224	219	2.2	239	217	9.2
REP 1B	254	235	7.5	217	174	19.8
REP 1C	197	163	17.3	214	198	7.5
REP 2A	221	207	6.3	215	208	3.2
REP 2B	259	251	3.1	263	241	9.4
REP 2C	208	212	-1.9	237	223	5.9

## TREATMENT: 100% RP2

	BEFORE	MARKED		BEFORE	UNMARKED	
		AFTER	%LOSS		AFTER	%LOSS
REP 1A	252	245	2.3	273	255	6.6
REP 1B	264	245	7.2	272	235	13.6
REP 1C	192	180	6.3	205	193	6.3
REP 2A	233	225	3.4	222	212	4.5
REP 2B	217	208	4.1	264	258	2.3
REP 2C	188	195	-3.7	207	192	7.2

# Appendix 7.d.1.

TRIAL #6 VALIDATION COMMENCED 12/4/91 PAGE 1  
ENDPOINT: DEVELOPMENTAL EMBRYONIC MORTALITY  
SPECIES A. gacinaea

TREATMENT :CONTROL DEVELOPMENTAL STAGE RAW DATA  
GASTRULA TROCH VEL HIPPO H'LING N'NATE

REP 1A  
REP 1B

## PERCENTILE DATA

REP 1A  
REP 1B

TOTAL 0 0 0 0 0 0  
PERCENT. 0 0 0 0 0 0

TREATMENT :CONTROL DEVELOPMENTAL STAGE RAW DATA  
GASTRULA TROCH VEL HIPPO H'LING N'NATE

REP 2A  
REP 2B  
REP 2C

## PERCENTILE DATA

REP 2A  
REP 2B  
REP 2C

TOTAL 0 0 0 0 0 0  
PERCENT. 0 0 0 0 0 3.9

TREATMENT :1% RP2 DEVELOPMENTAL STAGE RAW DATA  
GASTRULA TROCH VEL HIPPO H'LING N'NATE

REP 1A  
REP 1B

## PERCENTILE DATA

REP 1A  
REP 1B

TOTAL 0 0 0 0 0 0  
PERCENT. 0 0 0 0 0 0

TREATMENT :1% RP2 DEVELOPMENTAL STAGE RAW DATA  
GASTRULA TROCH VEL HIPPO H'LING N'NATE

REP 2A  
REP 2B  
REP 2C

## PERCENTILE DATA

REP 2A  
REP 2B  
REP 2C

TOTAL 0 0 0 0 0 0  
PERCENT. 0 0 0 0 0 0

TREATMENT :3.2% RP2 DEVELOPMENTAL STAGE RAW DATA  
GASTRULA TROCH VEL HIPPO H'LING N'NATE

REP 1A  
REP 1B

## PERCENTILE DATA

REP 1A  
REP 1B

TOTAL 1 0 0 0 0 0  
PERCENT. 3.6 0 0 0 0 0

TREATMENT :3.2% RP2 DEVELOPMENTAL STAGE RAW DATA  
GASTRULA TROCH VEL HIPPO H'LING N'NATE

REP 2A  
REP 2B

## PERCENTILE DATA

REP 2A  
REP 2B

TOTAL 0 0 0 0 0 0  
PERCENT. 0 0 0 0 0 0

TREATMENT :10% RP2 DEVELOPMENTAL STAGE RAW DATA  
GASTRULA TROCH VEL HIPPO H'LING N'NATE

REP 1A  
REP 1B

## PERCENTILE DATA

REP 1A  
REP 1B

TOTAL 0 0 0 0 0 0  
PERCENT. 0 0 0 0 0 0

TREATMENT :10% RP2 DEVELOPMENTAL STAGE RAW DATA  
GASTRULA TROCH VEL HIPPO H'LING N'NATE

REP 2A  
REP 2A

## PERCENTILE DATA

TOTAL 0 0 0 0 0 0  
PERCENT. 0 0 0 0 0 0

TREATMENT :100% RP2 DEVELOPMENTAL STAGE RAW DATA  
GASTRULA TROCH VEL HIPPO H'LING N'NATE

REP 1A

## PERCENTILE DATA

REP 1A

TOTAL 4

PERCENT. 100

# Appendix 7.d.2.

TRIAL #6 VALIDATION COMMENCED 12/4/91 PAGE 1  
ENDPOINT: DEVELOPMENTAL EMBRYONIC MORTALITY  
SPECIES A. cummingii

TREATMENT :CONTROL DEVELOPMENTAL STAGE RAW DATA  
GASTRULA TROCH VEL HIPPO H'LING N'NATE

REP 1A  
REP 1B  
REP 1C

## PERCENTILE DATA

REP 1A 5.9 5.9

REP 1B

REP 1C

TOTAL 1 0 1 0 0 0  
PERCENT. 1.4 0 1.4 0 0 0

TREATMENT :CONTROL DEVELOPMENTAL STAGE RAW DATA  
GASTRULA TROCH VEL HIPPO H'LING N'NATE

REP 2A  
REP 2B  
REP 2C

## PERCENTILE DATA

REP 2A  
REP 2B  
REP 2C

TOTAL 0 0 4 5 0 0  
PERCENT. 0 0 4.1 5.1 0 0

TREATMENT :1% RP2 DEVELOPMENTAL STAGE RAW DATA  
GASTRULA TROCH VEL HIPPO H'LING N'NATE

REP 1A  
REP 1B  
REP 1C

## PERCENTILE DATA

REP 1A  
REP 1B  
REP 1C

TOTAL 1 2 0 0 4  
PERCENT. 0.7 1.3 0 0 2.6

TREATMENT: 1% RP2 GASTRULA	DEVELOPMENTAL STAGE			RAW DATA	
	TROCH	VEL	HIPPO	H'LING	N'NATE
REP 1A		1	1		
REP 1B		3	2	1	
REP 1C					
PERCENTILE DATA					
REP 2A		2.5	2.5		
REP 2B		5.9	3.9	2	
REP 2C					
TOTAL	0	0	4	3	1
PERCENT.	0	0	3.2	2.4	0.8

TREATMENT: 3.2% RP2 GASTRULA	DEVELOPMENTAL STAGE			RAW DATA	
	TROCH	VEL	HIPPO	H'LING	N'NATE
REP 1A		1			
REP 1B		1			
REP 1C	1		2		
PERCENTILE DATA					
REP 1A		7.7			
REP 1B		2.3			
REP 1C	2.6		5.3		
TOTAL	0	1	2	2	0
PERCENT.	0	1.1	2.1	2.1	0

TREATMENT: 3.2% RP2 GASTRULA	DEVELOPMENTAL STAGE			RAW DATA	
	TROCH	VEL	HIPPO	H'LING	N'NATE
REP 2A	2	3			
REP 2B	1	1			
REP 2C		1	1		
PERCENTILE DATA					
REP 2A	8.7	1.3			
REP 2B	4.8	4.8			
REP 2C		5.9	5.9		
TOTAL	3	1	4	1	0
PERCENT.	4.9	1.6	6.6	1.6	0

TREATMENT: 10% RP2 GASTRULA	DEVELOPMENTAL STAGE			RAW DATA	
	TROCH	VEL	HIPPO	H'LING	N'NATE
REP 1A	1	2			
REP 1B		1	1		
REP 1C	1				
PERCENTILE DATA					
REP 1A	7.1	1.4			
REP 1B		2.5	2.5		
REP 1C	4.8				
TOTAL	1	1	3	1	0
PERCENT.	1.3	1.3	4	1.3	0

TREATMENT: 10% RP2 GASTRULA	DEVELOPMENTAL STAGE			RAW DATA	
	TROCH	VEL	HIPPO	H'LING	N'NATE
REP 2A			1		
REP 2B			1		
PERCENTILE DATA					
REP 2A			7.1		
REP 2B			7.1		
TOTAL	0	0	0	2	0
PERCENT.	0	0	0	7.1	0

TREATMENT: 32% RP2 GASTRULA	DEVELOPMENTAL STAGE			RAW DATA	
	TROCH	VEL	HIPPO	H'LING	N'NATE
REP 1A	1	1			
REP 1B			1		
PERCENTILE DATA					
REP 1A	7.7	7.7			
REP 1B			5.6		
TOTAL	1	0	1	1	0
PERCENT.	3.2	0	3.2	3.2	0

TREATMENT: 32% RP2 GASTRULA	DEVELOPMENTAL STAGE			RAW DATA	
	TROCH	VEL	HIPPO	H'LING	N'NATE
REP 2A					
PERCENTILE DATA					
REP 2A					
TOTAL	0	0	0	0	0
PERCENT.	0	0	0	0	0

TREATMENT: 100% RP2 GASTRULA	DEVELOPMENTAL STAGE			RAW DATA	
	TROCH	VEL	HIPPO	H'LING	N'NATE
REP 1A					
PERCENTILE DATA					
REP 1A					
TOTAL	0	0	0	0	0
PERCENT.	0	0	0	0	0

## Appendix 7.d.3.

TRIAL #6 VALIDATION COMMENCED 12/4/91 PAGE 1  
ENDPOINT: DEVELOPMENTAL EMBRYONIC MORTALITY  
SPECIES: GLYPTOPHYSA SP.

TREATMENT: CONTROL	CASTRULA	DEVELOPMENTAL STAGE			RAW DATA	
		TROCH	VEL	HIPPO	H'LING	N'NATE
REP 1A	2	4		2		
REP 1B	2		1	1		
REP 1C	9	7		1	3	

## PERCENTILE DATA

REP 1A	15.3	30.3		15.3		
REP 1B	22.2		11.1	11.1		
REP 1C	30	23.3		3.3	10	
TOTAL	13	11	1	4	3	0
PERCENT.	21.3	18	1.6	6.6	4.9	0

TREATMENT: CONTROL	CASTRULA	DEVELOPMENTAL STAGE			RAW DATA	
		TROCH	VEL	HIPPO	H'LING	N'NATE
REP 2A		4		4	3	
REP 2B			2	1		
REP 2C		1	8		3	

## PERCENTILE DATA

REP 2A		9.8		9.8	7.3	
REP 2B			9.1	4.5		
REP 2C		4.5	16.4		13.6	
TOTAL	0	5	10	4	6	0
PERCENT.	0	5.3	10.5	4.2	6.3	0

TREATMENT: 1% RP2	CASTRULA	DEVELOPMENTAL STAGE			RAW DATA	
		TROCH	VEL	HIPPO	H'LING	N'NATE
REP 1A	6	7	4	4		
REP 1B		9			1	

## PERCENTILE DATA

REP 1A	15.8	18.4	10.5			
REP 1B		26.5			2.9	
TOTAL	6	16	4	4	1	0
PERCENT.	8.3	22.2	5.6	5.6	1.4	0

TREATMENT: 1% RP2	CASTRULA	DEVELOPMENTAL STAGE			RAW DATA	
		TROCH	VEL	HIPPO	H'LING	N'NATE
REP 2A					1	
REP 2B		5				

## PERCENTILE DATA

REP 2A					4.3	
REP 2B		17.9				
TOTAL	0	5	0	0	1	0
PERCENT.	0	9.8	0	0	2	0

TREATMENT: 3.2% RP2	CASTRULA	DEVELOPMENTAL STAGE			RAW DATA	
		TROCH	VEL	HIPPO	H'LING	N'NATE
REP 1A		7			1	
REP 1B		1				3
REP 1C		11				

## PERCENTILE DATA

REP 1A		35			5	
REP 1B		9.1				27.3
REP 1C		100				
TOTAL	0	19	0	0	1	3
PERCENT.	0	45.2	0	0	2.4	7.1

TREATMENT: 3.2% RP2	CASTRULA	DEVELOPMENTAL STAGE			RAW DATA	
		TROCH	VEL	HIPPO	H'LING	N'NATE
REP 2A						
REP 2B		6				
REP 2C						

## PERCENTILE DATA

REP 2A						
REP 2B		14.6				
REP 2C						
TOTAL	0	6	0	0	0	0
PERCENT.	0	9.4	0	0	0	0

TREATMENT: 10% RP2	CASTRULA	DEVELOPMENTAL STAGE			RAW DATA	
		TROCH	VEL	HIPPO	H'LING	N'NATE

REP 1A

REP 1B

1

## PERCENTILE DATA

REF 1A 11.1

REP 1B

TOTAL

PERCENT.

0	0	0	1	0	0
---	---	---	---	---	---

TREATMENT: 10% RP2

CASTRULA

REP 2A

REP 2B

## PERCENTILE DATA

REP 2A 7.4 7.4 7.4

REP 2B

2.9

TOTAL

PERCENT.

0	2	2	3	0	0
---	---	---	---	---	---

TREATMENT: 32% RP2

CASTRULA

REP 1A

TOTAL

PERCENT.

1	0	0	0	1	0
---	---	---	---	---	---

REP 1A

REP 1A

TOTAL

PERCENT.

1	0	0	0	1	0
---	---	---	---	---	---

TREATMENT: 100% RP2

CASTRULA

REP 2A

REP 2A

## PERCENTILE DATA

TOTAL

PERCENT.

TOTAL

PERCENT.

0	0	1	1	0	0
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0	0	14.3	14.3	0	0
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0	0	0	0	0	0
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0	0	0	0	0	0
---	---	---	---	---	---



## Appendix 7.e.1.

TRIAL #6 VALIDATION COMMENCED 12/4/91 PAGE 1  
 ENDPOINT: JUVENILE MORTALITY EXPECTED: FROM NUMBERS OF  
 SPECIES A. carinaca NEONATES HATCHED

## TREATMENT: CONTROL

	LIVING	DEAD	%MORT.
REP 1A NOT SUIT.			
REP 1B	29	5	14.7
REP 1C	31	10	24.4
REP 2A	21	0	0
REP 2B	25	14	35.9
REP 2C	15	1	6.3

## TREATMENT: 1% RP2

	LIVING	DEAD	%MORT.
REP 1A	11	5	35.3
REP 1B NONE LAID			
REP 1C	8	3	27.3
REP 2A	6	0	0
REP 2B	10	0	0
REP 2C	3	0	0

## TREATMENT: 3.2% RP2

	LIVING	DEAD	%MORT.
REP 1A	19	3	13.6
REP 1B	13	4	23.5
REP 1C	20	3	13
REP 2A	7	0	0
REP 2B NOT SUIT.			
REP 2C	8	2	20

## TREATMENT: 10% RP2

	LIVING	DEAD	%MORT.
REP 1A NONE LAID			
REP 1B	14	4	22.2
REP 1C	9	4	30.8
REP 2A NOT SUIT.			
REP 2B	13	1	7.1
REP 2C NOT SUIT.			

## TREATMENT: 32% RP2

	LIVING	DEAD	%MORT.
REP 1A NONE LAID			
REP 1B NONE LAID			
REP 1C NONE LAID			
REP 2A NOT SUIT.			
REP 2B NONE LAID			
REP 2C NONE LAID			

## TREATMENT: 100% RP2

	LIVING	DEAD	%MORT.
REP 1A NONE LAID			
REP 1B NONE LAID			
REP 1C NONE LAID			
REP 2A NONE LAID			
REP 2B NONE LAID			
REP 2C NOT SUIT.			

## Appendix 7.e.3.

TRIAL #6 VALIDATION COMMENCED 12/4/91 PAGE 1  
 ENDPOINT: JUVENILE MORTALITY EXPECTED: FROM NUMBERS OF  
 SPECIES GLYPTOPHYSA SP. NEONATES HATCHED

## TREATMENT: CONTROL

	LIVING	DEAD	%MORT.
REP 1A	5	0	0
REP 1B	6	4	40
REP 1C	13	1	7.1
REP 2A	23	6	20.7
REP 2B	19	0	0
REP 2C	8	2	20

## TREATMENT: 1% RP2

	LIVING	DEAD	%MORT.
REP 1A	18	0	0
REP 1B	18	3	14.3
REP 1C	22	2	8.3
REP 2A	22	1	4.3
REP 2B NONE LAID			
REP 2C	19	3	13.6

## TREATMENT: 3.2% RP2

	LIVING	DEAD	%MORT.
REP 1A	11	0	0
REP 1B	6	1	14.3
REP 1C NOT SUIT.			
REP 2A	10	0	0
REP 2B	12	3	20
REP 2C	35	4	10.3

## TREATMENT: 10% RP2

	LIVING	DEAD	%MORT.
REP 1A	14	0	0
REP 1B NOT SUIT.			
REP 1C	8	0	0
REP 2A NOT SUIT.			
REP 2B	17	1	5.6
REP 2C	26	7	21.2

## TREATMENT: 32% RP2

	LIVING	DEAD	%MORT.
REP 1A NONE LAID			
REP 1B NONE LAID			
REP 1C	21	3	12.5
REP 2A NONE LAID			
REP 2B NONE LAID			
REP 2C NONE LAID			

## TREATMENT: 100% RP2

	LIVING	DEAD	%MORT.
REP 1A	10	3	30
REP 1B NONE LAID			
REP 1C NONE LAID			
REP 2A NONE LAID			
REP 2B	17	8	32
REP 2C NONE LAID			

## Appendix 7.e.2.

TRIAL #6 VALIDATION COMMENCED 12/4/91 PAGE 1  
 ENDPOINT: JUVENILE MORTALITY EXPECTED: FROM NUMBERS OF  
 SPECIES A. cummingii NEONATES HATCHED

## TREATMENT: CONTROL

	LIVING	DEAD	%MORT.
REP 1A	13	4	23.5
REP 1B	30	5	14.3
REP 1C	17	0	0
REP 2A	35	1	2.8
REP 2B	25	0	0
REP 2C	27	1	3.6

## TREATMENT: 1% RP2

	LIVING	DEAD	%MORT.
REP 1A	42	4	8.7
REP 1B	52	0	0
REP 1C	42	6	12.5
REP 2A	32	6	15.8
REP 2B	36	8	18.2
REP 2C	33	1	2.9

## TREATMENT: 3.2% RP2

	LIVING	DEAD	%MORT.
REP 1A	10	2	16.7
REP 1B	41	2	4.7
REP 1C	33	2	5.7
REP 2A	16	2	11.1
REP 2B	16	3	15.8
REP 2C	13	2	13.3

## TREATMENT: 10% RP2

	LIVING	DEAD	%MORT.
REP 1A	11	0	0
REP 1B	36	3	7.7
REP 1C	21	0	0
REP 2A	13	0	0
REP 2B	12	1	7.7
REP 2C NONE LAID			

## TREATMENT: 32% RP2

	LIVING	DEAD	%MORT.
REP 1A	11	0	0
REP 1B	14	3	17.6
REP 1C NONE LAID			
REP 2A NONE LAID			
REP 2B NONE LAID			
REP 2C	8	1	11.1

## TREATMENT: 100% RP2

	LIVING	DEAD	%MORT.
REP 1A NONE LAID			
REP 1B	12	3	20
REP 1C NONE LAID			
REP 2A	9	6	40
REP 2B NONE LAID			
REP 2C NONE LAID			

## Appendix 7.f.1.

TRIAL #6 VALIDATION COMMENCED 12/4/91  
ENDPOINT: MORTALITY OF CONTROL REARED JUVENILES  
SPECIES *A. cumingii*

PAGE 1

## TREATMENT: CONTROL

	LIVING	DEAD	SHORT.
REP 1A	6	4	40
REP 1B	9	1	10
REP 1C	10	0	0
REP 2A	5	5	50
REP 2B	10	0	0
REP 2C	8	2	2

## TREATMENT: 1% RP2

	LIVING	DEAD	SHORT.
REP 1A	9	1	10
REP 1B	9	1	10
REP 1C	9	1	10
REP 2A	6	4	40
REP 2B	9	1	10
REP 2C	3	7	70

## TREATMENT: 3.2% RP2

	LIVING	DEAD	SHORT.
REP 1A	8	2	20
REP 1B	8	2	20
REP 1C	10	0	0
REP 2A	4	6	60
REP 2B	9	1	10
REP 2C	9	1	10

## TREATMENT: 10% RP2

	LIVING	DEAD	SHORT.
REP 1A	10	0	0
REP 1B	7	3	30
REP 1C	8	2	20
REP 2A	9	1	10
REP 2B	7	3	30
REP 2C	8	2	20

## TREATMENT: 32% RP2

	LIVING	DEAD	SHORT.
REP 1A	10	0	0
REP 1B	9	1	10
REP 1C	8	2	20
REP 2A	5	5	50
REP 2B	6	4	40
REP 2C	8	2	20

## TREATMENT: 100% RP2

	LIVING	DEAD	SHORT.
REP 1A	9	1	10
REP 1B	10	0	0
REP 1C	10	0	0
REP 2A	10	0	0
REP 2B	10	0	0
REP 2C	8	2	2

## Appendix 7.g.

TRIAL #6 VALIDATION COMMENCED 12/4/91  
PHYSICO-CHEMICAL DATA

## DISSOLVED OXYGEN - mg/L

	CON	CON	1%RP2	1%RP2	3.2%RP2	3.2%RP2	10%RP2	10%RP2	32%RP2	32%RP2	100%RP2	100%
DAY1	7.6	7.6	7.8	7.6	7.8	7.8	7.7	7.7	7.7	7.9	8	8
DAY2	7.5	7.5	7.5	7.2	7.5	7.6	7.5	7.5	7.5	7.4	7.5	7
DAY3	7.6	7.5	7.5	7.5	7.4	7.5	7.4	7.3	7.2	7.4	7.5	7
DAY4	INSTRUMENT FAILURE											
DAY5												
DAY6												
DAY7												
DAY8	7.8	7.8	7.8	7.8	7.9	7.8	7.9	7.9	7.9	7.9	7.7	7
DAY9	7.8	7.9	7.8	7.8	7.8	7.7	7.8	7.7	7.8	7.7	7.7	7
DAY10	7.7	7.8	7.8	7.8	7.8	7.5	7.8	7.5	7.7	7.7	7.7	7
DAY11	7.8	7.7	7.7	7.8	7.8	7.7	7.7	7.7	7.7	7.7	7.4	7
DAY12	7.8	7.6	7.6	7.6	7.8	7.8	7.6	7.7	7.7	7.7	7.5	7
DAY13	7.7	7.7	7.8	7.8	7.8	7.8	7.8	7.8	7.8	7.8	7.9	7
DAY14	7.7	7.8	7.8	7.8	7.8	7.8	7.8	7.8	7.8	7.8	7.8	7
DAY15	7.7	7.8	7.8	7.7	7.7	7.8	7.8	7.8	7.8	7.8	7.8	7
DAY16	7.8	7.6	7.9	7.7	7.7	7.7	7.7	7.7	7.7	7.7	7.8	7
DAY17	7.7	7.7	7.8	7.8	7.8	7.7	7.7	7.7	7.7	7.8	7.8	7
DAY18	7.8	7.7	7.7	7.7	7.7	7.7	7.7	7.7	7.8	7.8	7.8	7

## ON

DAY1	8	6.18	6.24	5.29	6.3	6.4	6.43	6.63	6.79	6.99	6.9	7.2
DAY2	6.41	6.34	6.46	5.71	6.33	6.26	6.38	6.67	6.73	6.8	6.88	7.2
DAY3	6.32	6.54	6.52	6.81	6.72	6.78	6.69	6.88	7.9	7.03	7.2	7.2
DAY4	INSTRUMENT FAILURE											
DAY5												
DAY6												
DAY7												
DAY8	6	5.87	5.95	5.91	6.02	5.96	6.14	6.25	6.45	6.25	6.71	6.27
DAY9	6.41	6.14	6.44	6.62	6.85	6.73	6.93	7.17	7.2	7.26	7.57	7.41
DAY10	6.34	6.61	6.72	5.92	7.04	7.12	7.14	7.32	7.33	7.46	7.8	7.41
DAY11	6.26	6.63	6.85	6.91	6.94	7.02	7.1	7.34	7.42	7.5	7.55	7.27
DAY12	5.89	6.45	5.7	5.81	7.02	7.15	7.07	7.47	7.64	7.67	7.78	7.2
DAY13	6.22	6.2	6.16	6.3	6.84	7.2	7.21	7.15	7.45	7.5	7.67	7.27
DAY14	6.38	6.45	6.51	6.74	6.65	6.78	6.85	7.07	7.1	7.43	7.35	7.5
DAY15	6.32	6.45	6.62	6.78	6.7	6.8	6.77	6.9	6.97	7.12	7.43	7.5
DAY16	6.51	6.4	6.29	6.42	6.43	6.54	6.64	6.72	6.87	7.03	7.33	7.5
DAY17	6.71	6.54	6.62	6.68	6.64	6.78	6.96	7.01	6.95	7.1	7.28	7.35
DAY18	6.48	6.72	6.82	6.79	6.76	6.82	6.82	6.99	7.13	7.13	7.33	7.4

## CONDUCTIVITY - uS/cm/cm

DAY1	25.7	26	38.6	39.3	71.9	72.2	157	166	420	423	1080	1090
DAY2	36.4	29.6	46.3	44.1	76.5	75.6	177	174	421	433	1100	1100
DAY3	33.8	24.2	43.6	41.2	70.8	74.7	171	168	421	420	1070	1077
DAY4	INSTRUMENT FAILURE											
DAY5												
DAY6												
DAY7												
DAY8	52.2	43.6	56.1	53.2	82.2	79.4	176	170	457	434	1110	1112
DAY9	21.9	20.1	34.9	36.2	73.7	71.2	171	172	461	438	1120	1120
DAY10	20.4	21.6	38.1	40.8	72.7	72.6	177	171	446	441	1140	1140
DAY11	21.7	24.7	37	37.8	70.2	75.7	170	175	448	442	1140	1140
DAY12	22.4	28.3	35.8	37.1	70.4	79.1	174	178	449	441	1140	1140
DAY13	25.8	31	39.9	41.5	70.7	83.7	170	174	430	423	1080	1080
DAY14	24.5	25.1	38.1	37.3	70.3	76.4	172	173	430	426	1080	1080
DAY15	22.7	22.3	35.6	25.7	70.7	69.9	172	173	441	434	1120	1120
DAY16	20.7	21.9	35.6	35.3	68.4	68	166	167	438	429	1110	1110
DAY17	21.1	24	34.7	34.1	71.7	68.2	177	175	440	447	1090	1090
DAY18	19.7	21.9	35.1	34.4	67.2	67.2	162	171	430	428	1090	1100

## Appendix 8.a.1.

TRIAL #7 COMMENCED 2/5/91 RP2-URANIUM COMPARISON  
ENDPOINT: DAILY EGG MASS PRODUCTION RAW DATA  
SPECIES: A. carinata

## TREATMENT :1% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	2	2	1	1	1	2
DAY2	0	2	0	2	0	1
DAY3	0	1	0	0	1	0
DAY4	1	1	1	2	1	2
DAY5	0	0	1	1	0	1

## TREATMENT :1% U

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	4	2	2	3	3	4
DAY2	1	1	1	0	0	0
DAY3	1	0	0	0	2	0
DAY4	0	2	1	2	2	1
DAY5	2	0	0	1	1	0

## TREATMENT :10% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	0	0	0	2
DAY2	0	0	0	0	0	0
DAY3	0	0	0	0	0	0
DAY4	0	0	0	0	0	0
DAY5	0	0	0	0	0	0

## TREATMENT :10% U

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	3	1	1	2	3	2
DAY2	0	0	0	0	0	0
DAY3	0	0	0	0	1	0
DAY4	0	1	1	0	0	0
DAY5	0	0	1	0	0	0

## TREATMENT :32% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	1	1	0	1	0
DAY2	0	0	0	0	0	0
DAY3	0	0	0	0	0	0
DAY4	1	1	0	0	0	0
DAY5	0	1	0	0	0	0

## TREATMENT :32% U

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	0	1	1	2
DAY2	0	0	0	0	0	0
DAY3	0	0	0	0	0	0
DAY4	0	0	0	0	0	0
DAY5	0	0	0	0	0	0

TRIAL #7 COMMENCED 2/5/91 RP2-URANIUM COMPARISON  
ENDPOINT: DAILY TOTAL EGG PRODUCTION RAW DATA  
SPECIES: A. carinata

## TREATMENT :1% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	24	8	11	21	11	17
DAY2	0	0	9	12	0	8
DAY3	0	10	0	0	10	0
DAY4	18	5	12	27	11	22
DAY5	0	0	9	13	0	10

## TREATMENT :1% U

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	29	10	0	22	13	14
DAY2	12	0	0	0	0	0
DAY3	26	0	0	0	39	0
DAY4	0	13	0	8	3	11
DAY5	14	0	0	7	0	0

## TREATMENT :10% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	0	0	0	28
DAY2	0	0	0	0	0	0
DAY3	0	0	0	0	0	0
DAY4	0	0	0	0	0	0
DAY5	0	0	0	0	0	0

## TREATMENT :10% U

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	12	12	16	0	12
DAY2	0	0	0	0	0	0
DAY3	0	0	0	0	0	0
DAY4	0	12	10	0	0	0
DAY5	0	0	12	0	0	0

## TREATMENT :32% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	0	0	0	0
DAY2	0	11	0	0	0	0
DAY3	0	0	0	0	0	0
DAY4	5	11	0	0	0	0
DAY5	0	0	0	0	0	0

## TREATMENT :32% U

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	0	19	16	29
DAY2	0	0	0	0	0	0
DAY3	0	0	0	0	0	0
DAY4	0	0	0	0	0	0
DAY5	0	0	0	0	0	0

## Appendix 8.a.2.

TRIAL #7 COMMENCED 2/5/91 RP2-URANIUM COMPARISON  
ENDPOINT: DAILY EGG MASS PRODUCTION RAW DATA  
SPECIES: A. cummingii

## TREATMENT :1% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	2	3	3	1	1	2
DAY2	2	2	2	1	1	2
DAY3	0	2	3	1	2	2
DAY4	3	1	2	4	2	1
DAY5	3	2	1	2	2	2

## TREATMENT :1% U

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	2	2	3	2	1	0
DAY2	0	0	1	2	2	2
DAY3	2	0	2	0	0	1
DAY4	1	2	1	2	1	2
DAY5	2	3	2	3	3	3

## TREATMENT :10% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	2	1	0	2	1	1
DAY2	1	0	0	2	0	0
DAY3	0	0	0	0	0	0
DAY4	0	1	1	0	1	1
DAY5	0	0	0	2	2	0

## TREATMENT :10% U

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	1	2	4	2	2	2
DAY2	1	2	1	3	0	0
DAY3	0	0	1	0	1	0
DAY4	2	1	0	0	0	1
DAY5	1	1	0	1	0	1

## TREATMENT :32% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	2	0	1	0	0	0
DAY2	0	0	0	0	0	0
DAY3	0	0	0	0	0	0
DAY4	0	0	0	0	0	0
DAY5	1	0	0	0	0	0

## TREATMENT :32% U

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	0	0	0	0
DAY2	0	0	0	0	0	0
DAY3	1	1	1	0	0	0
DAY4	1	0	0	0	0	0
DAY5	0	0	0	0	3	0

TRIAL #7 COMMENCED 2/5/91 RP2-URANIUM COMPARISON  
ENDPOINT: DAILY TOTAL EGG PRODUCTION RAW DATA  
SPECIES: A. cummingii

## TREATMENT :1% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	31	53	36	20	16	31
DAY2	33	38	23	22	18	28
DAY3	0	44	38	23	31	25
DAY4	59	22	28	56	26	14
DAY5	25	30	11	31	29	31

## TREATMENT :1% U

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	29	35	53	28	20	0
DAY2	0	0	15	28	19	35
DAY3	35	0	30	0	0	12
DAY4	30	33	20	34	77	25
DAY5	20	36	38	33	57	50

## TREATMENT :10% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	22	17	0	32	15	10
DAY2	13	0	0	35	0	0
DAY3	0	0	0	0	0	0
DAY4	0	18	6	0	15	4
DAY5	0	0	0	23	35	0

## TREATMENT :10% U

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	22	32	48	36	33	33
DAY2	24	26	0	33	0	0
DAY3	0	0	9	0	18	0
DAY4	37	13	0	0	0	18
DAY5	13	9	0	16	0	18

## TREATMENT :32% RP2

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	19	0	14	0	0	0
DAY2	0	0	0	0	0	0
DAY3	0	0	0	0	0	0
DAY4	0	0	0	0	0	0
DAY5	6	0	0	0	0	0

## TREATMENT :32% U

SAMPLE	A	REP 1 B	C	A	REP 2 B	C
DAY1	0	0	0	0	0	0
DAY2	0	0	0	0	0	0
DAY3	19	16	29	0	0	0
DAY4	0	0	0	0	0	0
DAY5	0	0	0	0	2	0

## Appendix 8.b.1.

TRIAL #7 COMMENCED 1/5/91 RP2-URANIUM COMPARISON  
 ENDPOINT: DEVELOPMENTAL CHARACTERISTICS PAGE 1  
 SPECIES: *A. carinata*

TREATMENT: 1% RP2 REPLICATE 1A  
 GASTRULA TROCH VEL HIPPO H'LING N'MATE

DAY1 6  
 15  
 DAYS 6  
 14  
 DAYS 5  
 14  
 DAY7 5  
 14

TREATMENT: 1% RP2 REPLICATE 1B  
 GASTRULA TROCH VEL HIPPO H'LING N'MATE

DAY1 10  
 DAY4 10  
 DAYS 10  
 DAY6 10  
 DAY7 10  
 DAY8 10

TREATMENT: 1% RP2 REPLICATE 1C  
 GASTRULA TROCH VEL HIPPO H'LING N'MATE

DAY1 11  
 DAYS 11  
 DAY6 11  
 DAY7 11  
 DAY8 11

TREATMENT: 1% RP2 REPLICATE 2A  
 GASTRULA TROCH VEL HIPPO H'LING N'MATE

DAY1 17  
 DAY4 15  
 DAYS 15  
 DAY6 15  
 DAY7 15  
 DAY8 15  
 DAY9 15

TREATMENT: 1% RP2 REPLICATE 2B  
 GASTRULA TROCH VEL HIPPO H'LING N'MATE

DAY1 11  
 DAYS 11  
 DAY6 INFECTED  
 DAY7 DEAD

TREATMENT: 1% RP2 REPLICATE 2C  
 GASTRULA TROCH VEL HIPPO H'LING N'MATE

DAY1 13  
 14  
 DAYS 13  
 14  
 DAY6 13  
 14  
 DAY7 13  
 14  
 DAY8 13  
 14

TREATMENT: 1% U REPLICATE 1A  
 GASTRULA TROCH VEL HIPPO H'LING N'MATE

DAY1 12  
 DAYS 11  
 DAY6 11  
 DAY7 11

TREATMENT: 1% U REPLICATE 1B  
 GASTRULA TROCH VEL HIPPO H'LING N'MATE

DAY1 13  
 DAY3 13  
 DAY4 13  
 DAYS 13  
 DAY6 13  
 DAY7 13  
 DAY8 13  
 DAY9 13  
 DAY10 13

TREATMENT: 1% U REPLICATE 2A  
 GASTRULA TROCH VEL HIPPO H'LING N'MATE

DAY1 8  
 8  
 5  
 DAYS 8  
 8  
 4  
 DAY6 INFECTED  
 INFECTED  
 INFECTED  
 DAY7 DEAD  
 DEAD  
 DEAD

TREATMENT: 1% U REPLICATE 2C  
 GASTRULA TROCH VEL HIPPO H'LING N'MATE

DAY1 13  
 DAYS 13  
 DAY6 10  
 DAY7 7

TREATMENT: 10% RP2 REPLICATE 1 NO EGGS LAID  
 TREATMENT: 10% RP2 REPLICATE 2C  
 GASTRULA TROCH VEL HIPPO H'LING N'MATE

DAY1 10  
 18  
 DAYS 10  
 18  
 DAY6 10  
 18  
 DAY7 10  
 18  
 DAY8 10  
 18

TREATMENT: 10% U REPLICATE 1B  
 GASTRULA TROCH VEL HIPPO H'LING N'MATE

DAY1 12  
 DAYS 12  
 DAY6 12  
 DAY7 12  
 DAY8 12

TREATMENT: 10% U REPLICATE 1C  
 GASTRULA TROCH VEL HIPPO H'LING N'MATE

DAY1 12  
 DAYS 12  
 DAY7 12

TREATMENT: 10% U REPLICATE 2C  
 GASTRULA TROCH VEL HIPPO H'LING N'MATE

DAY1 12  
 DAYS 12  
 DAY7 12  
 DAY8 12

TREATMENT: 32% RP2 REPLICATE 1A  
 GASTRULA TROCH VEL HIPPO H'LING N'MATE

DAY1 13  
 8  
 DAY4 13  
 8  
 DAYS 13  
 8  
 DAY6 13  
 8  
 DAY7 13  
 8

TREATMENT: 32% RP2 REPLICATE 1B  
 GASTRULA TROCH VEL HIPPO H'LING N'MATE

DAY1 14  
 DAY3 14  
 DAY4 14  
 DAYS 14  
 DAY6 14  
 DAY7 14

TREATMENT: 32% RP2 REPLICATE 1C  
 GASTRULA TROCH VEL HIPPO H'LING N'MATE

DAY1 19  
 26  
 10  
 DAY4 19  
 24  
 10  
 DAYS 18  
 24  
 9  
 DAY6 18  
 24  
 9  
 DAY7 18  
 24  
 9

TREATMENT: 32% RP2 REPLICATE 2A  
GASTRULA TROCH VEL HIPPO H'LING N'NATE

DAY1 13  
9  
DAY4  
DAY5 13  
9  
DAY6 13  
9  
DAY7 13  
9

TREATMENT: 32% RP2 REPLICATE 2B  
GASTRULA TROCH VEL HIPPO H'LING N'NATE

DAY1 7  
20  
7  
DAY5 7  
20  
7  
DAY6 7  
20  
7  
DAY7 7  
20

TREATMENT: 32% RP2 REPLICATE 2C  
GASTRULA TROCH VEL HIPPO H'LING N'NATE

DAY1 15  
18  
DAY3 15  
18  
DAY4 14  
18  
DAY5 14  
18  
DAY6 12  
18  
DAY7 12  
18

TREATMENT: 32% U REPLICATE 1A  
GASTRULA TROCH VEL HIPPO H'LING N'NATE

DAY1 9  
10  
DAY4 9  
10  
DAY5 9  
10  
DAY6 9  
10  
DAY7 9  
10  
DAY8 9  
10

TREATMENT: 32% U REPLICATE 1B  
GASTRULA TROCH VEL HIPPO H'LING N'NATE

DAY1 12  
DAY3 12  
DAY4 12  
DAY5 12  
DAY6 12  
DAY7 12  
DAY8 12

TREATMENT: 32% U REPLICATE 1C  
GASTRULA TROCH VEL HIPPO H'LING N'NATE

DAY1 11  
12  
DAY3 11  
12  
DAY4 11  
12  
DAY5 11  
12  
DAY6 11  
12  
DAY7 11  
12  
DAY8 11  
12

TREATMENT: 32% U REPLICATE 2A  
GASTRULA TROCH VEL HIPPO H'LING N'NATE

DAY1 7  
11  
DAY4 7  
11  
DAY5 7  
11  
DAY6 7  
11  
DAY7 7  
11

TREATMENT: 32% U REPLICATE 2B  
GASTRULA TROCH VEL HIPPO H'LING N'NATE

DAY1 11  
11  
DAY3 11  
10  
DAY4 11  
10  
DAY5 11  
10  
DAY6 11  
10  
DAY7 11  
10  
DAY8 10  
10  
DAY9 10  
10

TREATMENT: 32% U REPLICATE 2C  
GASTRULA TROCH VEL HIPPO H'LING N'NATE

DAY1 12  
DAY3 12  
DAY4 12  
DAY5 12  
DAY6 12  
DAY7 12  
DAY8 12

12  
12  
12  
12

## Appendix 8.b.2.

TRIAL #7 COMMENCED 1/5/91 RP2-URANIUM COMPARISON  
ENDPOINT: DEVELOPMENTAL CHARACTERISTICS PAGE 1  
SPECIES: *A. cummingii*

TREATMENT: 1% RP2 REPLICATE 1A  
GASTRULA TROCH VEL HIPPO H'LING N'NATE

DAY1 13  
20

DAY5

12  
20

DAY6

11  
19

DAY7

11  
19

DAY8

11  
19

DAY9

11  
19

TREATMENT: 1% RP2 REPLICATE 1B  
GASTRULA TROCH VEL HIPPO H'LING N'NATE

DAY1 16  
22

DAY5

12  
18

DAY6

11  
18

DAY7

11  
18

DAY8

11  
18

DAY9

11  
18

TREATMENT: 1% RP2 REPLICATE 1C  
GASTRULA TROCH VEL HIPPO H'LING N'NATE

DAY1 8  
15

DAY5

7

15

DAY6

7

15

DAY7

7  
15

DAY8

7  
15

DAY9

7  
15

TREATMENT: 1% RP2 REPLICATE 2A  
GASTRULA TROCH VEL HIPPO H'LING N'NATE

DAY1 12  
22

DAY4

12

22

DAY5

12

22

DAY6

12  
22

DAY7

12  
22

DAY8

12  
22

TREATMENT: 1% RP2 REPLICATE 2B  
GASTRULA TROCH VEL HIPPO H'LING N'NATE

DAY1 18

DAY5

18

DAY6

18

DAY7

18

DAY8

18

DAY9

18

TREATMENT: 1% RP2 REPLICATE 2C  
GASTRULA TROCH VEL HIPPO H'LING N'NATE

DAY1 16

DAY5

15

DAY6

15

DAY7

15

DAY8

15

TREATMENT: 1% U REPLICATE 1A  
GASTRULA TROCH VEL HIPPO H'LING N'NATE

DAY1 14

DAY5

10

DAY6

10

DAY7

10

DAY8

10

TREATMENT: 1% RP2 REPLICATE 1B  
GASTRULA TROCH VEL HIPPO H'LING N'NATE

DAY1 19

DAY4

18

DAY5

18

DAY6

18

DAY7

18

DAY8

18

TREATMENT: 1% RP2 REPLICATE 1C  
GASTRULA TROCH VEL HIPPO H'LING N'NATE

DAY1 16  
15

DAY5

16  
14

DAY6

13  
14

DAY7

13  
14

DAY8

13  
14

TREATMENT: 1% U REPLICATE 2A  
GASTRULA TROCH VEL HIPPO H'LING N'NATE

DAY1 13  
15

DAY4

11  
14

DAY5

11  
14

DAY6

11  
14

DAY7

11  
14

DAY8

11  
14

TREATMENT: 1% U REPLICATE 2B  
GASTRULA TROCH VEL HIPPO H'LING N'NATE

DAY1 20

DAY6

14

DAY5

INFECTED

DAY6

DEAD

TREATMENT: 1% U REPLICATE 2C  
GASTRULA TROCH VEL HIPPO H'LING N'NATE

DAY1 19  
16

DAY4

19

16

DAY5

INFECTED  
INFECTED

DAY6

DEAD  
DEAD

TREATMENT: 10% RP2 REPLICATE 1A  
GASTRULA TROCH VEL HIPPO H'LING N'NATE

DAY1 13

DAY5

13

DAY6

13

DAY7

13

DAY8

13

DAY9

13

TREATMENT: 10% RP2 REPLICATE 1B  
GASTRULA TROCH VEL HIPPO H'LING N'NATE

DAY1 17

DAY6

16

DAY7

16

DAY8

16

TREATMENT: 10% RP2 REPLICATE 1C	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	6					
DAY5		6				
DAY6			6			
DAY7				6		
DAY8					6	
DAY9					5	
DAY10					5	
DAY11					5	
DAY12						5

TREATMENT: 10% RP2 REPLICATE 1D	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	24					
DAY2	22					
DAY3		20				
DAY4			20			
DAY5				19		
DAY6					19	
DAY7					18	
DAY8						18

TREATMENT: 10% RP2 REPLICATE 1E	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	22					
DAY3		19				
DAY4		19				
DAY5				19		
DAY6				19		
DAY7					17	
DAY8						17

TREATMENT: 10% RP2 REPLICATE 1F	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	18					
DAY3		16				
DAY4				16		
DAY5				2	14	
DAY6					16	
DAY7					16	
DAY8						16

TREATMENT: 10% RP2 REPLICATE 2A	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	16					
DAY6					14	
DAY7					14	
DAY8						14

TREATMENT: 10% RP2 REPLICATE 2B	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	15					
DAY6					12	
DAY7					12	
DAY8						12

TREATMENT: 10% RP2 REPLICATE 2D	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	30					
DAY3		30				
DAY4		29				
DAY5			25			
DAY6			11			
DAY7			INFECTED			
DAY8			DEAD			

TREATMENT: 10% RP2 REPLICATE 2E	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	16					
DAY3		14				
DAY4		12				
DAY5			12			
DAY6			INFECTED			
DAY7			DEAD			

TREATMENT: 10% RP2 REPLICATE 2F	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	16					
DAY3		13				
DAY4		12				
DAY5				10		
DAY6				INFECTED		
DAY7				DEAD		

TREATMENT: 10% U REPLICATE 1A	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	22					
DAY6					22	
DAY7					22	
DAY8						22

TREATMENT: 10% U REPLICATE 1B	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	13					
	16					
DAY6					13	
					16	
DAY7					13	
					16	
DAY8						13
						16

TREATMENT: 10% U REPLICATE 1C	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	15					
	16					
DAY6					15	
					16	
DAY7					15	
					16	
DAY8						15
						16

TREATMENT: 10% U REPLICATE 1D	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	11					
DAY3		11				
DAY4		10				
DAY5				10		
DAY6					9	
DAY7					3	
DAY8						3
DAY9						3

TREATMENT: 10% U REPLICATE 1E	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	23					
DAY3		23				
DAY4			23			
DAY5				23		
DAY6				23		
DAY7					21	
DAY8					21	
DAY9						21

TREATMENT: 10% U REPLICATE 1F	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	20					
DAY3		19				
DAY4			19			
DAY5				18		
DAY6					18	
DAY7					18	
DAY8						18

TREATMENT: 10% U REPLICATE 2A	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	21					
	15					
DAY6					15	
					13	
DAY7					15	
					13	
DAY8						15
						13

TREATMENT: 10% U REPLICATE 2B	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	18					
	17					
DAY6					15	
					14	
DAY7					15	
					14	
DAY8						15
						14

TREATMENT: 10% U GASTRULA REPLICATE 20 TROCH VEL HIPPO H'LING N'NATE

DAY1 18  
15  
DAY6 18  
15  
DAY7 18  
15  
DAY8 18  
15

TREATMENT: 10% U GASTRULA REPLICATE 20 TROCH VEL HIPPO H'LING N'NATE

DAY1 16  
DAY3 15  
DAY4 13  
DAY5 4  
DAY6 DEAD

TREATMENT: 10% U GASTRULA REPLICATE 2E TROCH VEL HIPPO H'LING N'NATE

DAY1 33  
DAY3 16  
DAY4 13  
DAY5 13  
DAY6 5 7  
DAY7 12  
DAY8 11  
DAY9 11

TREATMENT: 10% U GASTRULA REPLICATE 2F TROCH VEL HIPPO H'LING N'NATE

DAY1 26  
DAY3 24  
DAY4 23  
DAY5 23  
DAY6 23  
DAY7 23  
DAY8 23  
DAY9 23

TREATMENT: 32% RP2 REPLICATE 1A GASTRULA TROCH VEL HIPPO H'LING N'NATE

DAY1 13  
DAY6 13  
DAY7 13  
DAY8 13  
DAY9 13  
DAY10 13  
DAY11 13

TREATMENT: 32% RP2 REPLICATE 1C GASTRULA TROCH VEL HIPPO H'LING N'NATE

DAY1 14  
DAY6 14  
DAY7 14  
DAY8 14

TREATMENT: 32% RP2 REPLICATE 1D GASTRULA TROCH VEL HIPPO H'LING N'NATE

DAY1 19  
DAY2 19  
DAY3 19  
DAY4 19  
DAY5 19  
DAY6 19  
DAY7 19  
DAY8 19

TREATMENT: 32% RP2 REPLICATE 1E GASTRULA TROCH VEL HIPPO H'LING N'NATE

DAY1 17  
DAY2 17  
DAY3 17  
DAY4 17  
DAY5 16  
DAY6 16  
DAY7 16  
DAY8 16  
DAY9 16  
DAY10 16

TREATMENT: 32% RP2 REPLICATE 1F GASTRULA TROCH VEL HIPPO H'LING N'NATE

DAY1 27  
DAY2 24  
DAY3 24  
DAY4 23  
DAY5 23  
DAY6 23  
DAY7 23

TREATMENT: 32% RP2 REPLICATE 2D GASTRULA TROCH VEL HIPPO H'LING N'NATE

DAY1 21  
DAY2 20  
DAY3 20  
DAY4 20  
DAY5 20  
DAY6 5 14  
DAY7 19  
DAY8 19  
DAY9 19

TREATMENT: 32% RP2 REPLICATE 2E GASTRULA TROCH VEL HIPPO H'LING N'NATE

DAY1 29  
DAY2 29  
DAY3 29  
DAY4 28  
DAY5 29  
DAY6 19  
DAY7 29  
DAY8 29

TREATMENT: 32% RP2 REPLICATE 2F GASTRULA TROCH VEL HIPPO H'LING N'NATE

DAY1 11  
DAY2 10  
DAY3 10  
DAY4 10  
DAY5 5 5  
DAY6 3  
DAY7 3  
DAY8 3  
DAY9 3

TREATMENT: 32% U GASTRULA REPLICATE 1A TROCH VEL HIPPO H'LING N'NATE

DAY1 19  
DAY4 18  
DAY5 18  
DAY6 18

TREATMENT: 32% U GASTRULA REPLICATE 1B TROCH VEL HIPPO H'LING N'NATE

DAY1 16  
DAY4 16  
DAY5 16  
DAY6 16

TREATMENT: 32% U GASTRULA REPLICATE 1C TROCH VEL HIPPO H'LING N'NATE

DAY1 18  
11  
DAY4 4  
18  
DAY5 4  
10  
DAY6 4  
6  
DAY7 4  
6  
DAY8 4  
6  
DAY9 4  
6  
DAY10 4  
5  
DAY11 4  
5



TREATMENT: 32% U		REPLICATE 1D	CONTROL LAID			
DAY	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	10					
DAY2	10					
DAY3		10				
DAY4			10			
DAY5					7	
DAY6					BREAKDOWN	
DAY7					DEAD	

TREATMENT: 32% U		REPLICATE 1E	CONTROL LAID			
DAY	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	18					
DAY2	17					
DAY3		17				
DAY4			13			
DAY5					9	
DAY6					BREAKDOWN	
DAY7					DEAD	

TREATMENT: 32% U		REPLICATE 1F	CONTROL LAID			
DAY	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	17					
DAY2	17					
DAY3		17				
DAY4			17			
DAY5					BREAKDOWN	
DAY6					BREAKDOWN	
DAY7					5	
DAY8					7	
DAY9						7

TREATMENT: 32% U		REPLICATE 2D	CONTROL LAID			
DAY	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	16					
DAY2	16					
DAY3		16				
DAY4				16		
DAY5					15	
DAY6					13	
DAY7					11	
DAY8						11

TREATMENT: 32% U		REPLICATE 2E	CONTROL LAID			
DAY	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	11					
DAY2						
DAY3		11				
DAY4			11			
DAY5				7		
DAY6					2	
DAY7					1	
DAY8					0	

TREATMENT: 32% U		REPLICATE 2F	CONTROL LAID			
DAY	GASTRULA	TROCH	VEL	HIPPO	H'LING	N'NATE
DAY1	17					
DAY2						
DAY3		17				
DAY4			11			
DAY5					7	
DAY6					7	
DAY7					6	
DAY8						6

# Appendix 8.c.1.

TRIAL #7 COMMENCED 1/5/91 RP2-URANIUM COMPARISON  
ENDPOINT: CONTROL REARED JUVENILE MORTALITY PAGE 1  
SPECIES: A. carinata 10 INDIVIDUALS PER SAMPLE

TREATMENT: 1% RP2	2 DAY EXPOSURE %MORT	4 DAY EXPOSURE %MORT
REP1A	100	70
REP1B	100	70
REP2A	80	80
REP2B	100	70
TREATMENT: 1% U	2 DAY EXPOSURE %MORT	4 DAY EXPOSURE %MORT
REP1A	100	100
REP1B	90	100
REP2A	100	100
REP2B	100	100
TREATMENT: 10% RP2	2 DAY EXPOSURE %MORT	4 DAY EXPOSURE %MORT
REP1A	100	40
REP1B	100	100
REP2A	70	80
REP2B	40	80
TREATMENT: 10% U	2 DAY EXPOSURE %MORT	4 DAY EXPOSURE %MORT
REP1A	100	100
REP1B	100	100
REP2A	100	100
REP2B	90	100
TREATMENT: 32% RP2	2 DAY EXPOSURE %MORT	4 DAY EXPOSURE %MORT
REP1A	40	100
REP1B	80	90
REP2A	20	60
REP2B	40	60
TREATMENT: 32% U	2 DAY EXPOSURE %MORT	4 DAY EXPOSURE %MORT
REP1A	100	100
REP1B	100	100
REP2A	100	100
REP2B	100	100

# Appendix 8.c.2.

TRIAL #7 COMMENCED 1/5/91 RP2-URANIUM COMPARISON  
ENDPOINT: CONTROL REARED JUVENILE MORTALITY PAGE 1  
SPECIES: A. cumingii 10 INDIVIDUALS PER SAMPLE

TREATMENT: 1% RP2	2 DAY EXPOSURE %MORT	4 DAY EXPOSURE %MORT
REP1A	0	40
REP1B	0	30
REP2A	30	30
REP2B	10	20
TREATMENT: 1% U	2 DAY EXPOSURE %MORT	4 DAY EXPOSURE %MORT
REP1A	10	30
REP1B	10	30
REP2A	0	10
REP2B	30	10
TREATMENT: 10% RP2	2 DAY EXPOSURE %MORT	4 DAY EXPOSURE %MORT
REP1A	30	20
REP1B	20	20
REP2A	10	10
REP2B	0	0
TREATMENT: 10% U	2 DAY EXPOSURE %MORT	4 DAY EXPOSURE %MORT
REP1A	20	60
REP1B	20	10
REP2A	30	60
REP2B	20	60
TREATMENT: 32% RP2	2 DAY EXPOSURE %MORT	4 DAY EXPOSURE %MORT
REP1A	10	20
REP1B	0	30
REP2A	60	50
REP2B	50	80
TREATMENT: 32% U	2 DAY EXPOSURE %MORT	4 DAY EXPOSURE %MORT
REP1A	70	100
REP1B	90	100
REP2A	80	100
REP2B	90	100

# Appendix 8.d.1.

TRIAL #7 COMMENCED 1/5/91 RP2-URANIUM COMPARISON  
ENDPOINT: JUVENILE MORTALITY EXPOSED EGG MASS PAGE 1  
SPECIES: A. carinata

TREATMENT: 1% RP2	LIVING	DEAD	%MORT
REP1A	19	0	0
REP1B	10	0	0
REP1C	6	5	45.5
REP2A	15	0	0
REP2B	15	0	0
REP2C	17	10	37
TREATMENT: 1% U	LIVING	DEAD	%MORT
REP1A	9	2	18.2
REP1B	11	1	9.1
REP1C	NOT SUIT.		
REP2A	INFECTED		
REP2B	NOT SUIT.		
REP2C	0	7	100
TREATMENT: 10% RP2	LIVING	DEAD	%MORT
REP1A	NONE LAID		
REP1B	NONE LAID		
REP1C	NONE LAID		
REP2A	NONE LAID		
REP2B	NONE LAID		
REP2C	14	14	50
TREATMENT: 10% U	LIVING	DEAD	%MORT
REP1A	NOT SUIT.		
REP1B	10	2	16.7
REP1C	3	9	75
REP2A	NOT SUIT.		
REP2B	NOT SUIT.		
REP2C	5	7	58.3
TREATMENT: 32% RP2	LIVING	DEAD	%MORT
REP1A	NONE LAID		
REP1B	14	1	6.7
REP1C	NOT SUIT.		
REP2A	NONE LAID		
REP2B	NOT SUIT.		
REP2C	NONE LAID		
TREATMENT: 32% U	LIVING	DEAD	%MORT
REP1A	NONE LAID		
REP1B	NONE LAID		
REP1C	NONE LAID		
REP2A	NOT SUIT.		
REP2B	NOT SUIT.		
REP2C	NOT SUIT.		
TREATMENT: 32% RP2	LIVING	DEAD	%MORT
REP1D	20	1	4.8
REP1E	12	2	14.3
REP1F	35	16	31.4
REP2D	18	4	18.2
REP2E	34	0	0
REP2F	26	4	13.3
TREATMENT: 32% U	LIVING	DEAD	%MORT
REP1D	0	19	100
REP1E	0	12	100
REP1F	0	23	100
REP2D	0	19	100
REP2E	0	20	100
REP2F	0	12	100

# Appendix 8.d.2.

TRIAL #7 COMMENCED 1/5/91 RP2-URANIUM COMPARISON  
ENDPOINT: JUVENILE MORTALITY EXPOSED EGG MASS PAGE 1  
SPECIES: A. cummingsii

TREATMENT: 1% RP2 LIVING	DEAD	SHORT
REP1A 26	4	11.3
REP1B 26	3	10.3
REP1C 21	1	4.5
REP2A 29	5	13.9
REP2B 16	2	11.1
REP2C 13	2	13.3

TREATMENT: 1% U LIVING	DEAD	SHORT
REP1A 10	0	0
REP1B 17	0	0
REP1C 27	0	0
REP2A 8	17	68
REP2B INFECTED		
REP2C INFECTED		

TREATMENT: 10% RP2 LIVING	DEAD	SHORT
REP1A 13	0	0
REP1B 14	2	12.5
REP1C 3	2	4.0
REP2A 20	9	31
REP2B 7	5	41.7
REP2C NOT SUIT.		

TREATMENT: 10% U LIVING	DEAD	SHORT
REP1A 0	22	100
REP1B 13	16	55.2
REP1C 18	10	35.7
REP2A 14	14	50
REP2B 13	16	55.2
REP2C 18	15	45.5

TREATMENT: 32% RP2 LIVING	DEAD	SHORT
REP1A 13	0	0
REP1B NONE LAID		
REP1C 12	2	14.3
REP2A NONE LAID		
REP2B NONE LAID		
REP2C NONE LAID		

TREATMENT: 32% U LIVING	DEAD	SHORT
REP1A 3	10	76.9
REP1B 0	9	100
REP1C 1	8	12.5
REP2A NONE LAID		
REP2B NONE LAID		
REP2C NONE LAID		

# Appendix 8.f.

TRIAL #7 COMMENCED 2/5/91 RP2-URANIUM COMPARISON  
PHYSICO-CHEMICAL DATA

DISSOLVED OXYGEN mg/L INSTRUMENT OVERHAUL

PH

	1%RP2	1%RP2	10%RP2	10%RP2	32%RP2	32%RP2	1%U	1%U	10%U	10%U	32%U	32
DAY1	6.63	6.63	6.25	6.4	7.05	7.17	5.48	5.72	7.09	6.91	6.74	6.9
DAY2	6.12	6.62	6.41	6.62	6.73	6.87	6.81	6.85	7.17	7.07	7.59	7.1
DAY3	6.05	6.09	6.02	6.27	6.3	6.55	6.21	6.38	6.8	6.66	7.09	6.8
DAY4	5.92	5.94	6.16	6.54	6.5	6.64	6.38	6.4	7.05	6.37	6.73	6.5
DAY5	6.22	6.41	6.25	6.38	6.62	6.77	6.23	6.28	6.25	6.28	7.33	7.1
DAY6	6.15	6.28	6.18	6.28	6.55	6.68	6.27	6.3	6.48	6.37	6.83	6.7
DAY7	6.12	6.38	6.3	6.37	6.43	6.78	6.37	6.45	6.62	6.47	7.11	7.0
DAY8	6.11	6.24	6.07	6.16	6.51	6.75	6.07	6.38	6.14	6.26	6.96	6.8
DAY9	6.09	6.28	6.2	6.26	6.94	6.91	6.5	6.8	6.53	6.54	7.2	7.0
DAY10	5.9	6.29	6.42	6.51	6.59	6.8	6.35	6.6	6.66	6.45	7.08	7.0
DAY11	6.21	6.5	6.4	6.48	7.07	7.03	6.62	6.39	6.48	6.55	7.07	6.7
DAY12	6.28	6.28	6.29	6.22	6.57	6.69	6.51	6.45	6.39	6.31	7	7.0
DAY13	6.17	6.19	6.37	6.4	6.81	6.75	6.19	6.16	6.41	6.35	6.86	6.7
DAY14	6.25	6.38	6.5	6.54	6.75	6.89	6.34	6.52	6.54	6.63	7.01	6.7
DAY15	6.04	6	6.36	6.37	6.61	6.71	7.01	6.72	6.95	6.65	6.94	6.5

TRIAL #7 COMMENCED 2/5/91 RP2-URANIUM COMPARISON  
PHYSICO-CHEMICAL DATA

CONDUCTIVITY	uS/cm/cm		TREATMENT									
	1%RP2	1%RP2	10%RP2	10%RP2	32%RP2	32%RP2	1%U	1%U	10%U	10%U	32%U	32%U
DAY1	39.2	44.9	170	176	450	453	26.7	23.9	25.8	23.8	32.2	34.7
DAY2	49.2	47.8	174	178	455	458	34.1	27.5	31	29.5	30.4	22.5
DAY3	39.7	47.8	171	185	458	460	34	20.7	23.7	22.6	24.7	23.8
DAY4	39.6	44.4	169	177	457	461	21.6	26.1	23.9	23.2	24.4	23.1
DAY5	42.1	40	172	176	457	461	23.7	23.9	22.1	26.3	28.9	24.7
DAY6	41	41.7	171	177	457	460	24.3	22.7	22.7	23.1	26.2	24.9
DAY7	40.9	43.1	175	177	467	464	23.1	27.2	20.6	23.5	20.7	20.1
DAY8	41.9	41.7	181	176	465	461	24.4	20.6	19.7	22.6	19.6	19.4
DAY9	44.3	41.1	174	181	466	466	23.3	20.6	19.6	24.8	19.7	19.9
DAY10	41.2	39.4	175	177	472	474	23.7	19.5	18.8	24.6	19.5	19.1
DAY11	40.3	39.9	173	186	468	464	24.4	18.7	18.9	29.6	19.2	18.9
DAY12	72.3	70.7	174	179	460	461	22.9	18.8	19.3	23.7	20.7	20
DAY13	42.4	42	171	172	474	475	21	19.3	19.9	19.7	21.5	19.8
DAY14	40.2	40.7	177	172	469	469	21.2	20.5	23.2	22.6	20.1	18.8
DAY15	36.2	35.5	166	168	463	456	23.4	17.7	19.8	20.6	18.1	19.3

# Appendix 8.e.1.

TRIAL #7 COMMENCED 1/5/91 RP2-URANIUM COMPARISON  
ENDPOINT: JUVENILE MORTALITY CONTROL EGG MASS PAGE 2  
SPECIES: A. cummingsii

TREATMENT: 10% RP2 LIVING	DEAD	SHORT
REP1D 13	5	27.8
REP1E 9	8	47.1
REP1F 13	3	18.8
REP2D BREAKDOWN		
REP2E BREAKDOWN		
REP2F BREAKDOWN		

TREATMENT: 10% U LIVING	DEAD	SHORT
REP1D 3	0	0
REP1E 16	5	23.8
REP1F 2	26	92.9
REP2D INFECTED		
REP2E 10	1	9.1
REP2F 19	4	17.4

TREATMENT: 32% RP2 LIVING	DEAD	SHORT
REP1D 16	3	15.8
REP1E 13	4	23.5
REP1F 20	3	13
REP2D NOT SUIT.		
REP2E 20	9	31
REP2F 3	1	0

TREATMENT: 32% U LIVING	DEAD	SHORT
REP1D BREAKDOWN		
REP1E BREAKDOWN		
REP1F 0	7	100
REP2D 0	7	100
REP2E BREAKDOWN		
REP2F 0	9	100

# Appendix 9.a.1

GENETIC VARIATION							
ENDPOINT: EMBRYONIC AND JUVENILE MORT							
PAGE 1							
SPECIES <i>A. carinata</i>							
SAMPLES FROM KNOWN PAIRS							
	DEV. TIME (DAYS)	EGG Nos.	Nos. HATCH	Nos. SURV.	%HATCH /LAID	%SURV. /HATCH	%SURV. /LAID
REP 1							
SAMPLE 1	8	20	7	4	35	57.1	16
2	8	10	10	8	100	80	80
3	8	9	9	9	100	100	100
REP 2							
SAMPLE 1	8	14	14	5	100	35.7	35.7
2	8	8	8	8	100	100	100
3	8	9	8	6	88.9	75	66.7
4	6	15	15	9	100	60	60
REP 3							
SAMPLE 1	7	22	22	17	100	77.3	77.3
2	7	9	9	4	100	44.4	44.4
3	7	25	25	22	100	88	88
REP 4							
SAMPLE 1	8	15	6	5	40	83.3	33.3
2	7	9	9	5	100	77.8	77.8
3	7	8	8	6	100	75	75
4	7	4	4	4	100	100	100

# Appendix 9.a.2

SPECIES <i>A. cummingii</i>							
	DEV. TIME (DAYS)	EGG Nos.	Nos. HATCH	Nos. SURV.	%HATCH /LAID	%SURV. /HATCH	%SURV. /LAID
REP 1							
SAMPLE 1	8	22	17	11	77.3	64.7	50
2	8	22	15	11	68.2	73.3	50
3	8	37	12	7	30.1	58.3	18.9
4	8	17	5	5	29.4	100	29.4
REP 2							
SAMPLE 1	7	48	22	20	45.8	90.9	41.7
2	7	28	28	21	100	75	75
3	7	32	22	19	68.8	86.4	59.4
REP 3							
SAMPLE 1	8	21	11	8	52.4	72.7	38.1
2	10	19	12	12	63.2	100	63.2
REP 4							
SAMPLE 1	8	25	11	9	44	81.8	36
2	7	16	16	13	100	43.3	43.3
3	9	22	18	17	81.8	94.4	77.3
REP 5							
SAMPLE 1	9	9	8	6	100	66.7	66.7
2	8	15	14	6	93.3	50	46.7
3	9	33	23	20	69.7	87	60.6
4	8	65	65	36	100	55.4	55.4

# Appendix 9.b.1

TRIAL CONTAINER/ALGAE TRIAL COMMENCED 2/5/91  
ENDPOINT: JUVENILE MORTALITY PAGE 1  
SPECIES *A. carinata*

TREATMENT: CONTROL WATER 20 NEONATES PER TREATMENT  
: LARGE/SMALL (STANDARD) VIALS  
: WITH/WITHOUT ALGAE

LARGE VIALS						
	+ALGAE LIVING	DEAD	%MORT	LIVING	-ALGAE DEAD	%MORT
REP1	5	15	75	8	12	60
REP2	7	13	65	7	13	65
REP3	11	9	45	12	8	40
REP4	2	18	90	3	17	85

SMALL VIALS						
	+ALGAE LIVING	DEAD	%MORT	LIVING	-ALGAE DEAD	%MORT
REP1	12	8	40	15	5	25
REP2	13	7	35	15	5	25
REP3	15	5	25	11	9	45
REP4	19	1	5	16	4	20

# Appendix 9.b.2

SPECIES *A. cummingii*

TREATMENT: CONTROL WATER 20 NEONATES PER TREATMENT  
: LARGE/SMALL (STANDARD) VIALS  
: WITH/WITHOUT ALGAE

LARGE VIALS						
	+ALGAE LIVING	DEAD	%MORT	LIVING	-ALGAE DEAD	%MORT
REP1	2	18	90	4	16	80
REP2	9	11	55	7	13	65
REP3	4	16	80	0	20	100
REP4	LOST			11	9	45

SMALL VIALS						
	+ALGAE LIVING	DEAD	%MORT	LIVING	-ALGAE DEAD	%MORT
REP1	1	19	95	7	13	65
REP2	11	9	45	20	0	0
REP3	17	3	15	15	5	25

## Appendix 10.a.

TRIAL #1 RESULTS OF URANIUM AND MANGANESE ANALYSIS  
DATE SUBMITTED 19/2/91

SAMPLE	URANIUM ug/L U	MANGANESE ug/L Mn
MAGELA CK WATER	1	6.5
0.3% RP2	6.1	12
1% RP2	16	18
3.2% RP2	53	37
3.2% RP2	48	38
10% RP2	170	90
32% RP2	520	180
BLANK	< 0.1	< 0.1

All samples were analysed using Scintrex Time Decay Flourimetric technique for the determination of uranium, and Graphite Furnace Atomic Absorption Spectrometric technique for the determination of manganese.

The 3.2% RP2 samples were randomly chosen replicates.

## Appendix 10.b.

TRIAL #2 RESULTS OF URANIUM AND MANGANESE ANALYSIS  
DATE SUBMITTED 28/2/91 COMMENCEMENT TRIAL#2

SAMPLE	URANIUM ug/L U	MANGANESE ug/L Mn
MAGELA CK WATER	5.7	13
1% RP2	21	22
3.2% RP2	62	41
10% RP2	180	90
32% RP2	510	170
32% RP2	550	170
100% RP2	1700	380
BLANK	< 0.1	< 0.1

All samples were analysed using Scintrex Time Decay Flourimetric technique for the determination of uranium, and Graphite Furnace Atomic Absorption Spectrometric technique for the determination of manganese.

The 32% RP2 samples were randomly chosen replicates.

## DATE SUBMITTED 28/2/91 TERMINATION TRIAL#2

SAMPLE	URANIUM ug/L U	MANGANESE ug/L Mn
MAGELA CK WATER	5	1.2
1% RP2	15	2.7
3.2% RP2	55	24
10% RP2	160	50
10% RP2	160	60
32% RP2	380	100
100% RP2	1400	440
BLANK	< 0.1	< 0.1

All samples were analysed using Scintrex Time Decay Flourimetric technique for the determination of uranium, and Graphite Furnace Atomic Absorption Spectrometric technique for the determination of manganese.

The 10% RP2 samples were randomly chosen replicates.

Appendix 10.c.

TRIAL #3 RESULTS OF URANIUM AND MANGANESE ANALYSIS  
DATE SUBMITTED 19/3/91 COMMENCEMENT TRIAL #3

SAMPLE	URANIUM ug/L U	MANGANESE ug/L Mn
MAGELA CK WATER	1.3	6.6
1% RP2	17	15
3.2% RP2	60	36
3.2% RP2	60	36
10% RP2	110	60
32% RP2	560	200
100% RP2	1600	460
BLANK	< 0.1	< 0.1

All samples were analysed using Scintrex Time Decay Flourimetric technique for the determination of uranium, and Graphite Furnace Atomic Absorption Spectrometric technique for the determination of manganese.

The 3.2% RP2 samples were randomly selected replicates.

Appendix 10.d.

TRIAL #5 RESULTS OF URANIUM AND MANGANESE ANALYSIS  
DATE SUBMITTED 28/3/91 COMMENCEMENT TRIAL #5

SAMPLE	URANIUM ug/L U	MANGANESE ug/L Mn
MAGELA CK WATER	0.2	1.4
1% RP2	11	2.3
3.2% RP2	37	3
10% RP2	160	56
10% RP2	180	46
32% RP2	420	310
100% RP2	1600	750
BLANK	< 0.1	< 0.1

All samples were analysed using Scintrex Time Decay Flourimetric technique for the determination of uranium, and Graphite Furnace Atomic Absorption Spectrometric technique for the determination of manganese.

The 10% RP2 samples were randomly selected replicates.

Appendix 10.d.

TRIAL #6 RESULTS OF URANIUM AND MANGANESE ANALYSIS		
DATE SUBMITTED	1/4/91	TRIAL #6
COMMENCEMENT	FIRST WATER LOAD	
SAMPLE	URANIUM ug/L U	MANGANESE ug/L Mn
MAGELA CK WATER	0.3	10
1% RP2	17	17
1% RP2	16	19
3.2% RP2	53	36
10% RP2	150	96
32% RP2	540	280
100% RP2	1700	860
BLANK	0.6	0.5

The 1% RP2 samples were randomly selected replicates.

DATE SUBMITTED	1/4/91	TRIAL #6
TERMINATION	FIRST WATER LOAD	
SAMPLE	URANIUM ug/L U	MANGANESE ug/L Mn
MAGELA CK WATER	2.7	110
1% RP2	32	110
1% RP2	140	640
10% RP2	670	890
32% RP2	1300	660
100% RP2	1800	600
BLANK	0.3	0.3

TRIAL #6 RESULTS OF URANIUM AND MANGANESE ANALYSIS		
DATE SUBMITTED	20/4/91	TRIAL #6
COMMENCEMENT	SECOND WATER LOAD	
SAMPLE	URANIUM ug/L U	MANGANESE ug/L Mn
MAGELA CK WATER	1.6	15
1% RP2	19	24
3.2% RP2	52	34
3.2% RP2	52	30
10% RP2	180	79
32% RP2	510	210
100% RP2	1600	920
BLANK	0.2	0.2

The 3.2% RP2 samples were randomly selected replicates.

DATE SUBMITTED	1/4/91	TRIAL #6
TERMINATION	SECOND WATER LOAD	
SAMPLE	URANIUM ug/L U	MANGANESE ug/L Mn
MAGELA CK WATER	0.9	1.6
1% RP2	16	8.4
3.2% RP2	50	19
10% RP2	170	57
10% RP2	180	58
32% RP2	540	180
100% RP2	1700	610
BLANK	0.1	0.1

TRIAL #6 RESULTS OF URANIUM AND MANGANESE ANALYSIS		
DATE SUBMITTED	25/4/91	TRIAL #6
Commencement	THIRD WATER LOAD	
SAMPLE	URANIUM ug/L U	MANGANESE ug/L Mn
MAGELA CK WATER	< 0.1	2.9
1% RP2	19	9.6
3.2% RP2	55	22
10% RP2	200	54
32% RP2	670	190
32% RP2	640	190
100% RP2	1900	690
BLANK	< .1	< .1

The 32% RP2 samples were randomly selected replicates.

DATE SUBMITTED	10/5/91	TRIAL #6
TERMINATION	THIRD WATER LOAD	
SAMPLE	URANIUM ug/L U	MANGANESE ug/L Mn
MAGELA CK WATER	1.4	3.5
1% RP2	37	56
3.2% RP2	72	42
3.2% RP2	70	33
10% RP2	220	84
32% RP2	730	230
100% RP2	2000	480
BLANK	0.2	< .1

All samples were analysed using Scintrex Time Decay Flourimetric technique for the determination of uranium, and Graphite Furnace Atomic Absorption Spectrometric technique for the determination of manganese.

The 10% RP2 samples were randomly selected replicates.

NOTE: SEDIMENTATION OBSERVED IN REPLICATES TAKEN

# Appendix 10.e.

TRIAL #7 RESULTS OF URANIUM AND MANGANESE ANALYSIS  
 DATE SUBMITTED 3/5/91 TRIAL #7  
 COMMENCEMENT

SAMPLE	URANIUM ug/L U	MANGANESE ug/L Mn
1% RP2	29	12
1% RP2	19	8.2
1% U	17	8.6
10% RP2	130	59
10% U	170	7.5
32% RP2	500	170
32% U	550	8
100% RP2	1300	540
100% U	1800	7.5
BLANK	< 0.1	< 0.1

The 1% RP2 samples were randomly selected replicates.

DATE SUBMITTED 31/5/91 TRIAL #7  
 TERMINATION

SAMPLE	URANIUM ug/L U	MANGANESE ug/L Mn
1% RP2	20	6.4
1% U	39	1.5
10% RP2	220	47
10% U	170	1.8
32% RP2	740	150
32% U	470	2
32% U	500	2.2
BLANK	12	< 0.1

All samples were analysed using Scintrex Time Decay Flourimetric technique for the determination of uranium, and Graphite Furnace Atomic Absorbption Spectrometric technique for the determination of manganese.

The 32% U samples were randomly selected replicates.



## Appendix 11.a.1.

## STATISTICAL TESTING

TEST: ANALYSIS OF VARIANCE  
 TRIAL: #1  
 ENDPOINT: JUVENILE MORTALITY

SPECIES: *A. carinata*

Source	dF	SS	MSS	F	P>F
Treatment	5	153.233	30.6466	0.29	0.9122
Error	18	1900.78	105.598		
Total	23	2054.01			

RESULT: NO SIGNIFICANT DIFFERENCE

SPECIES: *A. cummingii*

Source	dF	SS	MSS	F	P>F
Treatment	5	1299.85	259.970	1.12	0.3855
Error	18	4182.62	232.368		
Total	23	5482.47			

RESULT: NO SIGNIFICANT DIFFERENCE

SPECIES: GLYPTOPHYSA SP.

Source	dF	SS	MSS	F	P>F
Treatment	5	521.602	106.230	0.78	0.5757
Error	18	2447.16	135.953		
Total	23	2978.76			

RESULT: NO SIGNIFICANT DIFFERENCE

## Appendix 11.a.2.

## STATISTICAL TESTING

TEST: ANALYSIS OF VARIANCE  
 TRIAL: #1  
 ENDPOINT: DAYS TO HATCHING

SPECIES: *A. carinata*

Source	dF	SS	MSS	F	P>F
Treatment	5	1.7143	0.34286	1.87	0.1491
Error	18	2.2923	0.12735		
Total	23	5.0066			

RESULT: NO SIGNIFICANT DIFFERENCE

SPECIES: *A. cummingii*

Source	dF	SS	MSS	F	P>F
Treatment	5	0.95613	0.191228	0.73	0.6071
Error	18	4.68603	0.260335		
Total	23	5.64216			

RESULT: NO SIGNIFICANT DIFFERENCE

SPECIES: GLYPTOPHYSA SP.

Source	dF	SS	MSS	F	P>F
Treatment	5	1.0804	0.21608	0.45	0.8058
Error	18	3.5886	0.19937		
Total	23	9.6690			

RESULT: NO SIGNIFICANT DIFFERENCE

## Appendix 11.b.1

## STATISTICAL TESTING

TEST: ANALYSIS OF VARIANCE  
 TRIAL: #2  
 ENDPOINT: DAYS TO HATCHING - DEVELOPMENTAL RETARDATION

SPECIES: *A. cummingii*

Source	dF	SS	MSS	F	P>F
Treatment	5	6.8750	1.3750	3.19	0.308
Error	18	7.750	0.430556		
Total	23	14.6250			

RESULT: NO SIGNIFICANT DIFFERENCE

SPECIES: *GYRAULUS SP.*

Source	dF	SS	MSS	F	P>F
Treatment	5	42.8565	8.57197	31.67	0.0001
Error	18	4.87256	0.270203		
Total	23	47.7291			

RESULT: SIGNIFICANTLY DIFFERENT

MEANS: 5.4 / 5.81 / 5.2 / 5.5 / 5.54 / 9.05

SPECIES: *LYMNAEA SP.*

Source	dF	SS	MSS	F	P>F
Treatment	5	118.708	23.7417	100.55	0.0001
Error	18	4.250	0.236111		
Total	23	122.958			

RESULT: SIGNIFICANTLY DIFFERENT

MEANS: 7.5 / 7.5 / 7 / 7.25 / 7.25 / 13.25

## Appendix 11.b.2

## STATISTICAL TESTING

TEST: ANALYSIS OF VARIANCE  
 TRIAL: #2  
 ENDPOINT: EMBRYONIC MORTALITY

SPECIES: *A. cummingii*

Source	dF	SS	MSS	F	P>F
Treatment	5	627.383	125.476	0.67	0.6490
Error	18	3354.75	186.375		
Total	23	3982.13			

RESULT: NO SIGNIFICANT DIFFERENCE

SPECIES: *GYRAULUS SP.*

Source	dF	SS	MSS	F	P>F
Treatment	5	4060.98	812.197	2.38	0.082
Error	18	6150.32	341.685		
Total	23	10211.31			

RESULT: SIGNIFICANTLY DIFFERENT AT 10%

SPECIES: *LYMNAEA SP.*

Source	dF	SS	MSS	F	P>F
Treatment	5	469.194	93.8388	0.72	0.6196
Error	18	2358.75	131.042		
Total	23	2827.95			

RESULT: NO SIGNIFICANT DIFFERENCE

## Appendix 11.b.3

## STATISTICAL TESTING

TEST: DUNNETT'S COMPARISON TO CONTROL MEAN.  
TRIAL: #2  
ENDPOINT: DEVELOPMENTAL RETARDATION.

SPECIES: *Gyrodactylus* sp.

MEANS	CONT.L	1% RP2	3.2% RP2	10% RP2	32% RP2	100% RP2
	5.4	5.81	5.2	5.5	5.54	9.05

RANK	3.2%	CONT.	10%	32%	1%	100%
n = 7						

ANOVA EMSS = 0.2703

CONTRAST	DIFF.	S.E.	q	p	q'	CONCLUSION
10%	0.1	0.280	0.36	2	2.02	Do not reject.
32%	0.14	0.280	0.50	3	2.44	Do not reject.
1%	0.41	0.280	1.48	4	2.68	Do not reject.
100%	3.65	0.280	13.1	5	2.85	Reject.

NOEC 32% RP2  
LOEC 100% RP2.

SPECIES: *Lymnaea* sp.

MEANS	CONT.	1% RP2	3.2% RP2	10% RP2	32% RP2	100% RP2
	7.5	7.5	7	7.25	7.25	13.25

RANK	3.2%	10%/32%	CONT./1%	100%
Smallest n = 4				

ANOVA EMSS = 0.2361

CONTRAST	DIFF.	S.E.	q	p	q'	CONCLUSION
/100%	5.75	0.344	16.7	2	2.02	Reject.

NOEC 32% RP2  
LOEC 100% RP2.

## Appendix 11.c.2

## STATISTICAL TESTING

TEST: TWO WAY ANALYSIS OF VARIANCE  
TRIAL: #3  
ENDPOINT: EMBRYONIC MORTALITY - mortality to hatching stage, as hatching and as neonate.

SPECIES: *A. cummingii* zero values excluded

Source	dF	SS	MSS	F	P > F
Treatment	5	480.047	96.0094	1.41	0.2682
Error	18	1226.82	68.1563		
Total	16	355.119			
STAGE	1	3.62416	3.62416	0.21	0.7719

STAGE \* TREATMENT 8 72.0697 9.00871 0.23 0.9734  
RESULT: NO SIGNIFICANT DIFFERENCE BETWEEN TREATMENTS OR INTERACTION BETWEEN TREATMENTS AND STAGES

SPECIES: *A. cummingii* zero values excluded

Source	dF	SS	MSS	F	P > F
DEV.STAGE	2	331.339	165.670	5.6	0.0153
Error	15	444.136	29.6090		
Total	17	775.475			

RESULT: SIGNIFICANT DIFFERENCE IN MORTALITY BETWEEN STAGES  
MEANS: 3.1% / 1.7% / 0.0%

SPECIES: *A. carinata*

Source	dF	SS	MSS	F	P > F
Treatment	5	364.453	72.8909	1.79	0.131
STAGE	2	208.276	104.138	2.55	0.0872
INTERACTION	10	460.357	46.0357	1.13	0.3582

RESULT: NO SIGNIFICANT DIFFERENCE BETWEEN TREATMENTS OR STAGES, AND NO SIGNIFICANT INTERACTION BETWEEN TREATMENTS AND STAGES

## Appendix 11.c.1

## STATISTICAL TESTING

TEST: ANALYSIS OF VARIANCE  
TRIAL: #3  
ENDPOINT: DAYS TO HATCHING - DEVELOPMENTAL RETARDATION

SPECIES: *A. carinata*

Source	dF	SS	MSS	F	P > F
Treatment	5	16.8333	3.36667	6.73	0.0011
Error	18	9.0	0.50		
Total	23	25.8333			

RESULT: SIGNIFICANTLY DIFFERENT  
MEANS: 7.25 / 7.50 / 7.75 / 7.75 / 7.50 / 9.75

SPECIES: *A. cummingii*

Source	dF	SS	MSS	F	P > F
Treatment	5	6.20833	1.24167	3.31	0.027
Error	18	6.75	0.3750		
Total	23	12.9583			

RESULT: SIGNIFICANTLY DIFFERENT  
MEANS: 8.0 / 8.0 / 7.0 / 7.0 / 8.0 / 8.25

SPECIES:

Source	dF	SS	MSS	F	P > F
Treatment	5	26.0893	5.21786	5.2	0.0105
Error	18	22.3539	1.24188		
Total	23	48.4432			

RESULT: SIGNIFICANTLY DIFFERENT  
MEANS: 7.5 / 7.5 / 6.75 / 7.25 / 7.46 / 10.0

## STATISTICAL TESTING

TEST: TWO WAY ANALYSIS OF VARIANCE  
TRIAL: #3  
ENDPOINT: EMBRYONIC MORTALITY - mortality to hatching stage, as hatching and as neonate.

SPECIES: *GLYPTOPHYSA* SP.

Source	dF	SS	MSS	F	P > F
Treatment	5	998.492	199.690	0.46	0.8022
Error	18	7850.25	436.125		
Total	23	8848.70			
STAGE	2	954.650	477.325	2.28	0.1252
INTERACTION	5	1017.37	203.474	0.84	0.5397

RESULT: NO SIGNIFICANT DIFFERENCE BETWEEN TREATMENTS OR LIFE STAGES AND NO SIGNIFICANT INTERACTION BETWEEN TREATMENT AND STAGES.

## Appendix 11.c.3

## STATISTICAL TESTING

TEST: ANALYSIS OF VARIANCE  
 TRIAL: #3  
 ENDPOINT: JUVENILE MORTALITY

SPECIES: *A. carinata*

Source	dF	SS	MSS	F	P > F
Treatment	5	9140.92	1828.18	7.89	0.0004
Error	18	4170.17	231.676		
Total	23	13311.1			

RESULT: SIGNIFICANTLY DIFFERENT

MEANS: 16.5% / 24.5% / 45.1% / 10.9% / 33.6% / 96.1%

SPECIES: *A. cummingii*

Source	dF	SS	MSS	F	P > F
Treatment	5	10185.7	2037.15	11.89	0.0001
Error	18	5786.72	171.280		
Total	23	12205.4			

RESULT: SIGNIFICANTLY DIFFERENCE

MEANS: 22.5% / 16.4% / 11.4% / 5.3% / 9.1% / 92.7%

SPECIES: GLYPTOPHYSA SP.

Source	dF	SS	MSS	F	P > F
Treatment	5	6218.70	1243.74	3.4	0.017
Error	18	5786.72	332.595		
Total	23				

RESULT: SIGNIFICANTLY DIFFERENT

MEANS: 31.9% / 9.7% / 32.7% / 13.2% / 81.5% / 59.4%

## Appendix 11.c.4

## STATISTICAL TESTING

TEST: DUNNETT'S COMPARISON TO CONTROL MEAN.  
 TRIAL: #3  
 ENDPOINT: DEVELOPMENTAL RETARDATION - DAYS TO HATCH

SPECIES: *A. carinata*

MEANS	CON	1% RP2	3.2% RP2	10% RP2	32% RP2	100% RP2
	7.25	7.50	7.75	7.75	7.5	9.75

RANK	CON	1% RP2	3.2% RP2	10% RP2	32% RP2	100% RP2
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Smallest n=4 ANOVA EMSS=0.50

Ho: Xcon &gt; = Xa. H1: Xcon &lt; Xa.

CONTRAST DIFF. STD.ERR. |q| p q' CONCLUSION

CONTROL /

1% RP2 /

3.2% RP2 / 0.25 0.50 0.50 3 2.44 DO NOT REJECT Ho

3.2% RP2 /

10% RP2 0.50 0.50 1.0 5 2.85 DO NOT REJECT Ho

100% RP2 2.0 0.50 4.0 2.98 REJECT Ho

NOEC: 32% RP2

LOEC: 100% RP2

SPECIES: *A. cummingii*

MEANS	CON	1% RP2	3.2% RP2	10% RP2	32% RP2	100% RP2
	8	8	7	7	8	8.25

RANK	3.2% RP2	10% RP2	CON	1% RP2	32% RP2	100% RP2
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Smallest n=4 ANOVA EMSS=0.375

One-sided Ho: Ucon &gt; = Ua. H1: Ucon &lt; Ua. (Denoted by " ").

Two-sided Ho: Ucon - Ua = 0. H1: Ucon - Ua &lt; &gt; 0. (Denoted by " ").

CONTRAST DIFF. STD.ERR. |q| p q' CONCLUSION

CONTROL /

3.2% RP2 /

10% RP2 1.0 0.433 0.43 3 2.57 DO NOT REJECT Ho

100% RP2 0.25 0.433 0.57 2 2.44 DO NOT REJECT Ho

NOEC: 100% RP2

LOEC: UNDEFINED

## STATISTICAL TESTING

TEST: DUNNETT'S COMPARISON TO CONTROL MEAN.  
 TRIAL: #3  
 ENDPOINT: DEVELOPMENTAL RETARDATION - DAYS TO HATCH

SPECIES: GLYPTOPHYSA SP.

MEANS	CON	1% RP2	3.2% RP2	10% RP2	32% RP2	100% RP2
	7.5	7.5	6.75	7.25	7.46	10.0

RANK	3.2% RP2	10% RP2	32% RP2	CON	1% RP2	100% RP2
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Smallest n=4 ANOVA EMSS=1.24188

Ho: Xcon &gt; = Xa. H1: Xcon &lt; Xa.

CONTRAST DIFF. STD.ERR. |q| p q' CONCLUSION

CONTROL /

3.2% RP2 0.04 0.788 .05 2 2.57 DO NOT REJECT Ho

10% RP2 0.25 0.788 .32 3 3.03 DO NOT REJECT Ho

3.2% RP2 0.75 0.788 .95 4 3.29 DO NOT REJECT Ho

100% RP2 2.50 0.788 2.45 5 2.85 DO NOT REJECT Ho

NOEC: 100% RP2

LOEC: UNDEFINED

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