Australian Government Department of Agriculture, Water and the Environment

Aquatic animal diseases significant to Australia: identification field guide 5th edition

March 2020



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Foreword

The Australian Government Department of Agriculture, Water and the Environment is pleased to introduce the fifth edition of this field guide.

The field guide and its accompanying phone application aims to help people recognise diseases of significance to aquaculture and fisheries in Australia. This edition incorporates new and updated information gathered from an extensive review of the fourth edition. It now covers 53 aquatic animal diseases of significance to Australia that affect species of finfish, crustaceans, molluscs and amphibians.

Early recognition and reporting of suspected disease outbreaks is critical to protecting our unique aquatic biodiversity, and fisheries and aquaculture sectors. It can permit a rapid disease response to contain outbreaks, increasing the opportunity for disease eradication and thereby limiting potential impacts on aquatic animal industries and the environment. People who work closely with aquatic animals are usually the first to notice signs of a significant disease event. This field guide provides guidance for these people—aquaculture workers, commercial fishers, recreational fishers, quarantine staff, scientists, conservationists and students—to recognize significant aquatic animal diseases, should they occur.

Many people and institutions have contributed to the field guide, including fish health experts from industry, research organisations, state and territory governments, and government agencies of the Asia–Pacific region (including New Zealand), Canada, Denmark, Norway, the Netherlands, the United Kingdom and the United States of America.

Drawing extensively on experience and research activities in aquatic animal health management, both in Australia and abroad, the guide and its phone application complements the growing body of practical knowledge published for aquaculture and fisheries in Australia. On behalf of the Australian Government, I thank all contributors to this production for their efforts and commend this field guide to you.

Dr Mark Schipp Australian Chief Veterinary Officer

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Members of the Australian Government Department of Agriculture—Aquatic Animal Health Program coordinated the production of this fifth edition including Dr Cian Foster-Thorpe and Dr Ingo Ernst. Alistair Herfort is recognised as co-ordinator of the first three editions of the field guide.

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Introduction

This field guide provides ready access to information on the aquatic animal diseases significant to Australia. These diseases have potential to disrupt Australia's aquatic animal biodiversity, fisheries and aquaculture productivity, and international trade.

The diseases covered here are in <u>Australia's national list of reportable diseases of aquatic animals</u>, which includes those reportable through the regional Quarterly Aquatic Animal Disease reporting program (managed by the Network of Aquaculture Centres in Asia-Pacific, the Food and Agriculture Organization of the United Nations and World Organisation for Animal Health), as well as other diseases of national significance.

The field guide is aimed at fishery and aquaculture managers, their field staff, veterinary workers and students of aquatic animal health. The publication is also relevant to people in the seafood processing and retail industry, recreational and commercial fishers, and the general public. For people participating in national surveillance and monitoring for aquatic animal disease, the field guide is a valuable disease reference. For the casual reader, the field guide gives an informative and often graphic account of the diseases and organisms that threaten Australia's fisheries and expanding aquaculture industries.

The marine and freshwater environments in Australia are rich with many types of animals. The field guide focuses on fish, molluscs (such as oysters), crustaceans (such as prawns) and amphibians (such as frogs). It is not possible to list every disease for every species in a publication of this type. The emphasis is on infectious diseases found on Australia's National List of Reportable Diseases of Aquatic Animals, significant to our aquaculture industries, fisheries and environment.

Nature of disease

The diseases described in this field guide are caused by infectious agents—viruses, bacteria, fungi or parasites. Diseases with non-infectious causes are outside its scope. Non-infectious causes of disease in the aquatic environment are often attributable to the environment itself: morbidity and mortality can result from natural or human influenced events that lead to oxygen depletion, production of aquatic toxins, or changes in water temperature or salinity. However, disease will not occur simply because an infectious agent is present. The likelihood of disease being expressed is determined by the specific interactions between the host (the aquatic animal), the infectious agent or pathogen, and the environment (Figure 1).



Figure 1 Relationship between host, pathogen and the environment in disease outbreaks

Laboratory tests and sampling

Photographs of gross disease signs, such as those in this field guide, can help an investigator to create a preliminary list of possible disease agents (differential diagnosis list) for the case under investigation. However, although gross signs narrow the search for possible agents, they are not adequate for definitive diagnosis; representative samples from the diseased animal(s) and the environment in which they live need to be taken for analysis. Several publications offering information about sampling are available.

- The World Organisation for Animal Health (OIE) <u>Manual of diagnostic tests for aquatic animals</u> provides standard diagnostic methods for OIE-listed diseases.
- <u>Asia diagnostic guide to aquatic animal diseases</u> provides diagnostic guidelines for the pathogens and diseases listed in the Quarterly Aquatic Animal Disease Reporting System.
- The website of the Australian Government <u>Department of Agriculture</u>, <u>Water and the</u> <u>Environment</u> (DAWE) provides information on the management of inland water quality.

Reporting disease

Fishery and aquaculture industry managers, as well as farmers and their staff, should be aware of their responsibilities to rapidly report any suspicion of diseases on <u>Australia's national list of</u> reportable diseases of aquatic animals to local authorities.

To prepare for possible disease incursion, fishery and aquaculture industry managers should develop an emergency management plan, in consultation with farmers and appropriate extension staff.

If you identify signs of disease in a culture system, contact your aquatic animal health officer. If your observation is of wild aquatic animals, contact a wildlife or fisheries officer.

A national emergency animal disease watch hotline number has been established to assist early reporting of suspicious disease events. Call **1800 675 888** (free call and available 24 hours).

Current contact details for state and territory governments are provided in <u>appendix B</u>. Call to report your find or ask further questions on the observations you have made. You will be directed to an expert on diseases of aquatic animals within your state or territory.

Follow the directions and advice provided by the officer you contact. This field guide will help you find the information the officer needs.

Using the field guide

This field guide begins with coverage of the anatomy (section 1) of finfish, molluscs, crustaceans and amphibians, including images and illustrations. These are to help the reader describe lesions when reporting a suspected disease. It follows with descriptions for each infectious disease present in <u>Australia's national list of reportable diseases of aquatic animals</u>. These are presented alphabetically, and classified into infectious diseases affecting finfish, molluscs (such as oysters), crustaceans (such as prawns) and amphibians (such as frogs).

This field guide is available both as a PDF document and as a downloadable phone app. The field guide phone app (listed as the Aquatic disease field guide) can be downloaded free for iOS and android devices at the relevant iTunes or app store outlets. For more information, see the <u>Aquatic</u> <u>animal diseases significant to Australia: Identification field guide mobile application</u> introductory video.

In both the PDF document and phone app, each disease page describes the:

- signs of disease (at the farm/tank/pond level and gross and microscopic pathological signs)
- disease agent
- host species that carry the disease agent
- presence of the disease in Australia
- epidemiology of the disease
- other diseases in the field guide that may have similar signs
- sample collection and reporting of disease outbreaks.

The disease pages also include photographs of animals with gross signs of disease and/or histological images detailing the typical tissue changes present.

Diagnosing diseases of aquatic animals is a structured process, which begins with making detailed observations, then asking the right questions. The primary aim of this process is to create a broad list of possible diseases that may account for the observed signs. This list can be further refined by targeted questioning and diagnostic tests.

Questions to ask may include:

- Which species, age class, sizes and sex of animals are involved?
- What specifically have you observed—for example, behavioural changes, changes in feed intake (reduced or increased), changes in faecal output?
- Are there obvious gross lesions (such as colour changes, ulcers or spots)?
- Does disease result in morbidity only, or are there mortalities?
- What level of mortality has been observed?
- Do animals show signs of recovery?
- At what rate did you observe disease (sudden death or chronic progression of disease)?

- Is this the first occurrence of disease with this presentation in your facility?
- Are any neighbouring properties or facilities reporting diseases?
- Have there been recent introductions to the facility or any changes in practice?

After making your initial observations, go to the 'signs of disease' section of the disease entries. Look up the diseases listed that match your observations, and you will find photographs and further information to help you narrow the search for the cause of the disease. For example, you might have observed disease in a marine fish, but then find that the listed disease with similar presenting signs affects only freshwater species—in this way, you can eliminate the unlikely, and your list of differential diagnoses will be shorter. In all cases, however, laboratory tests would be required for a definitive diagnosis. Local authorities with responsibility for aquatic animal health can advise on further courses of action should you suspect any disease listed in the field guide.

Signs of disease

Diagnostic information based on disease signs at the farm, tank or pond level, and gross pathological signs (abnormalities that can be seen with the naked eye) are important. However, in most cases, it is impossible to arrive at a definitive diagnosis on the basis of these signs alone. Although some users will not have experience in histological examination, or the equipment required for such examination in the field, the field guide also lists microscopic pathological disease signs.

Mass mortality

A fish kill involving a range of species is more likely caused by an environmental problem, such as toxicity or oxygen depletion. Deaths limited to one species (where other species are also present) are more likely a result of an infectious agent.

Behavioural changes

All species of aquatic animals have characteristic protection, food-gathering and breeding behaviours. Abnormal behaviour, such as a decrease in feeding, could indicate stress from disease. Some behavioural changes can occur across groups of species, or even across different phyla. In molluscs, few behavioural differences are observed such as gaping or delayed closing (in oysters) and decreased feeding or adhesion to vertical surfaces (in abalone). Finfish often gather at water inlets or gasp for air at the surface if the water is depleted of oxygen. If irritated by skin parasites, they may scrape themselves on hard surfaces. Whirling or 'corkscrew' swimming could indicate disease from an infectious agent or aquatic toxins. In both finfish and crustaceans, gathering at the surface or pond edges can often be a sign of disease.

Gross signs

Gross pathological changes may indicate the presence of infectious disease. Careful observation and further laboratory investigation is required to make a definitive diagnosis because many of the signs are not singularly pathognomonic (characteristic for a specific disease) and may be common pathological changes associated with multiple infectious agents. The table below describes some of the more commonly observed visible signs and the groups of aquatic animals dealt with in this field guide for which these are most often observed.

| Sign | Finfish | Molluscs | Crustaceans | Amphibians |
|---|---------|----------|-------------|------------|
| Changes in the colour, texture and opacity of flesh | * | * | * | * |
| Tissue necrosis and lesions | * | * | * | * |
| Retraction of gill margins | na | * | na | na |
| Pustules | * | * | na | na |
| External spots | * | na | * | na |
| Changes in surface colour | * | na | * | * |
| Secondary fungal or bacterial growth | * | * | * | * |
| Deformities and tumours | * | na | * | * |
| Swollen or discoloured organs or faecal castes | * | * | * | * |
| White midgut line | na | na | * | na |
| Broken or damaged appendages | na | na | * | na |
| Erosion of shell | na | na | * | na |
| Lesions or ulcers on skin or gills | * | na | * | * |
| Haemorrhaging with associated anaemia | * | na | na | * |
| Granulomas | * | na | * | na |
| Exophthalmos (popeye) | * | na | na | na |
| Ascites (accumulation of fluid in peritoneal cavity) | * | na | na | * |
| Petechial haemorrhages (pinpoint bleeding in skin and mucous membranes) | * | na | na | * |
| Ecchymotic haemorrhages (bleeding or bruising beneath the skin or mucous membranes) | * | na | na | * |
| Excessive mucus on gills and skin | * | * | na | na |
| Dropsy (accumulation of fluid in body tissues) | * | na | na | * |
| Protrusion of scales | * | na | na | na |

Table 1 Potential gross signs of disease

* Potential gross sign of disease. **na** Sign is not applicable to the taxonomic grouping.

Host range

A list of species known to be susceptible to the infectious agent is provided. Species are further classified as either naturally susceptible (diseased animals have been identified in the wild) or experimentally susceptible. Lists of susceptible species reflect the information available at the time of publishing. However, with further understanding and sampling, it is expected that such lists will expand and/or require refining. Both common and scientific names for hosts are provided.

Presence in Australia

Information on the national distribution of diseases listed in the field guide is based on formal reporting through the regional Quarterly Aquatic Animal Disease reporting program. This program is managed by the Network of Aquaculture Centres in Asia–Pacific (NACA), Food and Agriculture Organisations of the United Nations (FAO) and the World Organisation for Animal Health (OIE). Australia has been an active participant since 1998.

Where a listed disease has been reported under the program to have been present, a map illustrating where it occurred in Australia is included. States or territories having reported disease are listed, and shown in orange in the distribution map. It is important to note that, although a map may identify a state or territory as having reported a disease, this implies neither that it is present at the time of publication nor that it occurs across the entire state or territory.

Readers should consult the <u>World Animal Health Information Database (WAHID) interface</u> for current information on global distribution of diseases outside Australia.

Exotic diseases

Diseases in this field guide described as exotic are those that do not occur in Australian aquatic animal populations. Diseases present in Australia may still be listed as exotic if they are subject to an ongoing emergency disease response to contain and eradicate the disease.

Endemic diseases

Endemic (enzootic) diseases are those that have established in Australian aquatic animal populations. They might be native to Australia or might have been introduced in the past.

Epidemiology

The field guide describes epidemiological factors that are important to each disease. The key to describing the epidemiology of a disease involves understanding the relationship between the infectious agent, the host/s and the environment. Factors relating to the infectious agent include its life cycle (direct transmission or a requirement for intermediate host stages), survival outside the host (carriage on fomites, passage in wastewater) and sensitivity to certain temperatures or salinities.

Host factors may include the susceptible species, ages, sexes and sizes involved. Environmental factors include seasonal and non-seasonal variations in temperature, food availability, salinity, available oxygen, species movement and exposure to different environments (for example, migrations or gathering for breeding). These factors can affect disease agent survivability and host immune competence.

Differential diagnosis

The list of similar diseases at the bottom of each disease page refers only to the diseases covered by this field guide. Gross signs observed might well be representative of a wider range of diseases not included here. Therefore, these diagnostic aids should not be used as a guide to a definitive diagnosis, but rather as a tool to help identify the diseases included in this field guide that most closely account for the observed gross signs. Further diagnostic testing will be required to confirm either presence or freedom from a suspected disease.

Appendix A: Scientific names

| Amphibians | Amphibians |
|-------------------------|----------------------------|
| Apistogramma spp. | Neurergus crocatus |
| Bufo marinus | Notophthalmus viridescens |
| Cichlidae | Onychodactylus japonicas |
| Cophixalus ornatus | Order Anura |
| Cyclorana alboguttata | Order Caudata |
| Cyclorana brevipes | Order Gymnophiona |
| Cynops cyanurus | Paramesotriton deloustali |
| Cynops ensicauda | Platyplectrum ornatus |
| Cynops orientalis | Pleurodeles waltl |
| Cynops pyrrhogaster | Pseudophryne coriacea |
| Euproctus platycephalus | Rhinella marina |
| Hydromantes strinatii | Salamandra algira |
| Hynobius nebulosus | Salamandra salamandra |
| Ichthyosaura alpestris | Salamandrella keyserlingii |
| Limnodynastes ornatus | Salamandrina perspicillata |
| Lissotriton italicus | Siren intermedia |
| Lissotriton vulgaris | Speleomantes spp. |
| Litoria caerulea | Taricha granulosa |
| Litoria inermis | Taudactylus acutirostris |
| Litoria rubella | Triturus cristatus |
| Litoria splendida | Tylototriton uyenoi |
| Mikrogeophagus ramirezi | Tylototriton vietnamensis |
| Mixophyes fasciolatus | Tylototriton ziegleri |

Crustaceans

| Acetes spp. |
|-----------------------------|
| Artemesia longinaris |
| Artemia salina |
| Astacopsis gouldi |
| Astacopsis spp. |
| Astacus astacus |
| Astacus leptodactylus |
| Astacus spp. |
| Austropotamobius pallipes |
| Austropotamobius torrentium |
| Callinectes arcuatus |

Crustaceans

Cambaroides japonicus Carcinus maenas Cherax destructor Cherax quadricarinatus Cherax spp. Eriocheir sinensis Euastacus kershawi Euastacus spp. Exopalaemon carinicauda Gammarus pulex

Geocherax spp.

| Crustaceans | Crustaceans |
|--|--------------------------------------|
| Hemigrapsus penicillatus | Penaeus (Fenneropenaeus) chinensis |
| Homarus americanus | Penaeus (Fenneropenaeus) indicus |
| Macrobrachium dayanum | Penaeus (Fenneropenaeus) merguiensis |
| Macrobrachium lanchesteri | Penaeus (Litopenaeus) occidentalis |
| Macrobrachium nipponense | Penaeus (Litopenaeus) schmitti |
| Macrobrachium rosenbergii | Penaeus (Litopenaeus) setiferus |
| Macrobrachium sintangene | Penaeus (Litopenaeus) stylirostris |
| Macrobrachium superbum | Penaeus (Litopenaeus) vannamei |
| Metapenaeus bennettae | Penaeus (Marsupenaeus) japonicus |
| Metapenaeus ensis | Penaeus esculentus |
| Orconectes spp. | Penaeus marginatus |
| Pacifastacus leniusculus | Penaeus monodon |
| Palaemon kadiakensis | Penaeus semisulcatus |
| Palaemon pugio | Portunus pelagicus |
| Palaemon serrifer | Portunus sanguinolentus |
| Palaemon styliferus | Potamon potamios |
| Palaemonetes kadiakensis | Procambarus clarkii |
| Paleamon pugio | Procambarus fallax. f. virginalis |
| Panulirus spp. | Scylla serrata |
| Penaeus (Farfantepenaeus) aztecus | Sesarma mederi |
| Penaeus (Farfantepenaeus) californiensis | Thalamita crenata |
| Penaeus (Farfantepenaeus) duorarum | Uca vocans |
| Penaeus (Farfantepenaeus) subtilis | |

Finfish—Non-salmonids

| Abramis brama | A |
|---------------------------|---|
| Acanthopagrus australis | A |
| Acanthopagrus latus | A |
| Acanthopagrus schlegelii | A |
| Acanthurus triostegus | Þ |
| Acipenser baerii | Þ |
| Acipenser gueldenstaedtii | Þ |
| Acipenser oxyrinchus | Þ |
| Acipenser transmontanus | Þ |
| Ambassis agassizii | A |
| Ambloplites rupestris | Þ |
| Ameiurus catus | Þ |
| Ameiurus melas | A |
| Ameiurus natalis | A |

Finfish—Non-salmonids

| Ameiurus nebulosus |
|----------------------|
| Ammodytes hexapterus |
| Ammodytes personatus |
| Ammodytes spp. |
| Ammodytes tobianus |
| Amniataba percoides |
| Anabas testudineus |
| Anarhichas lupus |
| Anarhichas minor |
| Anguilla anguilla |
| Anguilla australis |
| Anguilla japonica |
| Anguilla rostrata |
| Anguillidae spp. |

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| Finfish—Non-salmonids | Finfish—Non-salmonids |
|---------------------------|--|
| Anodontiglanis dahlia | Dorosoma cepedianum |
| Anoplopoma fimbria | Eigenmannia virescens |
| Aplocheilichthys normani | Enchelyopus cimbrius |
| Aplodinotus grunniens | Eopsetta grigorjewi |
| Argentina sphyraena | Epinephelus aeneus |
| Argyrosomus japonicus | Epinephelus akaara |
| Aristichthys nobilis | Epinephelus bruneus |
| Arius spp. | Epinephelus coioides |
| Astronotus ocellatus | Epinephelus fuscoguttatus |
| Atractoscion nobilis | Epinephelus fuscoguttatus × E. lanceolatus |
| Aulorhynchus flavidus | Epinephelus lanceolatus |
| Barbonymus schwanenfeldii | Epinephelus malabaricus |
| Bidyanus bidyanus | Epinephelus marginatus |
| Blicca bjoerkna | Epinephelus melanostigma |
| Branchiostegus japonicus | Epinephelus sexfasciatus |
| Carassius auratus | Epinephelus spp. |
| Carassius carassius | Epinephelus tauvina |
| Carassius sp. | Esox lucius |
| Chanos chanos | Esox masquinongy |
| Chelon auratus | Etroplus maculatus |
| Chrysophrys auratus | Evynnis japonica |
| Cinetodus froggatti | Evynnis tumifrons |
| Clarias batrachus | Exocoetus volitans |
| Clupea harengus | Fundulus heteroclitus |
| Clupea pallasii | Gadus chalcogramma |
| Cnidoglanis macrocephalus | Gadus macrocephalus |
| Colisa labiosa | Gadus microgadus |
| Coptodon zilli | Gadus morhua |
| Cottus gobio | Galaxias olidus |
| Cromileptes altivelis | Galaxiidae all species |
| Ctenolabrus rupestris | Gambusia affinis |
| Ctenopharyngodon idella | Gasterosteus aculeatus |
| Cymatogaster aggregata | Girella punctata |
| Cyprinidae | Girella tricuspidata |
| Cyprinus carpio | Glossamia aprion |
| Danio devario | Glossogobius giuris |
| Danio rerio | Glyptocephalus stelleri |
| Dicentrarchus labrax | Gymnocephalus cernua |

| Finfish—Non-salmonids | Finfish—Non-salmonids |
|------------------------------|--|
| Hexagrammos otakii | Maccullochella peelii |
| Hippoglossoides platessoides | Macquaria ambigua |
| Hippoglossus hippoglossus | Macquaria australasica |
| Hippoglossus stenolepis | Macquaria novemaculeata |
| Hoplobrotula armata | Macropodus opercularis |
| Hucho hucho | Melanogrammus aeglefinus |
| Hyperoplus lanceolatus | Melanotaenia splendida |
| Hypomesus pretiosus | Merlangius merlangus |
| Hypophthalmichthys molitrix | Merluccius productus |
| Hypophthalmichthys nobilis | Microgadus proximus |
| Hyporthodus septemfasciatus | Micromesistius poutassou |
| Ictalurus furcatus | Micropterus dolomieu |
| Ictalurus punctatus | Micropterus salmoides |
| Inimicus japonicus | Morone americana |
| Kurtus gulliveri | Morone chrysops |
| Labridae all species | Morone mississippiensis |
| Labrus bergylta | Morone saxatilis |
| Laetacara curviceps | Moxostoma anisurum |
| Lampetra fluviatilis | Moxostoma macrolepidotum |
| Larimichthys crocea | Mugil cephalus |
| Lateolabrax japonicus | Mugilidae, all species |
| Lateolabrax spp. | Mullus barbatus |
| Lates calcarifer | Nannoperca australis |
| Latris lineata | Nematalosa erebi |
| Leiopotherapon unicolor | Neoarius berneyi |
| Lepomis gibbosus | Neoarius leptaspis |
| Lepomis macrochirus | Neogobius melanostomus |
| Lethrinus haematopterus | Neosilurus ater |
| Lethrinus nebulosus | Notropis atherinoides |
| Leuciscus cephalus | Notropis hudsonius |
| Leuciscus idus | Noturus gyrinus |
| Leuciscus leuciscus | Oplegnathus fasciatus |
| Limanda limanda | Oplegnathus punctatus |
| Lota lota | Oreochromis aureus |
| Lutjanus argentimaculatus | Oreochromis niloticus |
| Lutjanus erythropterus | Oreochromis niloticus × O. aureus |
| Luxilus cornutus | Oreochromis niloticus × O. mossambicus |
| Maccullochella ikei | Oxyeleotris lineolatus |

| Finfish—Non-salmonids | Finfish—Non-salmonids |
|------------------------------|-------------------------------|
| Oxyeleotris marmorata | Pseudocaranx dentex |
| Oxyeleotris selheimi | Pseudochondrostoma polylepis |
| Pagrus auratus | Pseudopleuronectes americanus |
| Pagrus major | Pterapogon kauderni |
| Pampus argenteus | Pterophyllum scalare |
| Pangasianodon hypophthalmus | Puntius conchonius |
| Pangasius hypophthalmus | Rachycentron canadum |
| Paralichthyidae, all species | Reinhardtius hippoglossoides |
| Paralichthys olivaceus | Rhacochilus vacca |
| Parambassis gulliveri | Rhombosolea tapirina |
| Parapristipoma trilineatum | Rutilus rutilus |
| Parophrys vetulus | Sander lucioperca |
| Pelteobagrus fulvidraco | Sardinops sagax |
| Pelvicachromis pulcher | Sarotherodon galilaeus |
| Perca flavescens | Scardinius erythrophthalmus |
| Perca fluviatilis | Scatophagus argus |
| Percopsis omiscomaycus | Sciaenops ocellatus |
| Petromyzon marinus | Scleropages jardini |
| Phoxinus phoxinus | Scomber japonicus |
| Pimephales notatus | Scomberomorus niphonius |
| Pimephales promelas | Scophthalmus maximus |
| Platax orbicularis | Scortum barcoo |
| Platichthys flesus | Sebastes schlegelii |
| Platycephalus fuscus | Sebastes spp. |
| Platycephalus indicus | Seriola dumerili |
| Plecoglossus altivelis | Seriola hippos |
| Plectorhinchus cinctus | Seriola lalandi |
| Plectranthias yamakawai | Seriola quinqueradiata |
| Plectropomus leopardus | Sillago ciliata |
| Plectropomus maculatus | Sillago maculata |
| Pleuronectes platessa | Silurus asotus |
| Poecilia latipinna | Silurus glanis |
| Poecilia reticulata | Siniperca chuatsi |
| Poecilia sphenops | Solea senegalensis |
| Pollachius virens | Solea solea |
| Pomatoschistus minutus | Sparus aurata |
| Pomoxis nigromaculatus | Sprattus sprattus |
| Pristiapogon exostigma | Stephanolepis cirrhifer |

| Finfish—Non-salmonids | Finfish—Non-salmonids |
|------------------------|--|
| Strongylura krefftii | Trichogaster lalius |
| Takifugu rubripes | Trichogaster leerii |
| Tandanus tropicanus | Trichogaster microlepis |
| Thaleichthys pacificus | Trichogaster trichopterus |
| Thunnus thynnus | Trisopterus esmarkii |
| Tilapia zilli | Trisopterus minutus |
| Tinca tinca | Tristramella simonis |
| Toxotes chatareus | Umbrina cirrosa |
| Toxotes jaculatrix | Verasper moseri |
| Toxotes lorentzi | Verasper variegatus |
| Trachinotus blochii | Xiphophorus helleri |
| Trachinotus falcatus | Xiphophorus maculatus |
| Trachurus japonicus | Zoarces viviparous |
| Trichiurus lepturus | |
| Finfish—Salmonids | Finfish—Salmonids |
| Constant and the | On any the second secon |

| Coregonus artedi | Oncorhynchus rhodurus | |
|----------------------------------|--------------------------------------|--|
| Coregonus clupeaformis | Oncorhynchus spp. | |
| Coregonus muksun | Oncorhynchus tshawytscha | |
| Coregonus peled | Prosopium williamsoni | |
| Coregonus spp. | Salmo labrax | |
| Oncorhynchus aguabonita | Salmo salar | |
| Oncorhynchus clarkii | Salmo trutta | |
| Oncorhynchus gilae | Salmonidae all species | |
| Oncorhynchus gorbuscha | Salvelinus alpinus | |
| Oncorhynchus keta | Salvelinus confluentus | |
| Oncorhynchus kisutch | Salvelinus fontinalis | |
| Oncorhynchus masou | Salvelinus leucomaenis | |
| Oncorhynchus mykiss | Salvelinus namaycush | |
| Oncorhynchus mykiss × O. kisutch | Salvelinus namaycush × S. fontinalis | |
| Oncorhynchus nerka | Thymallus thymallus | |

Molluscs

| Anadara trapezia | | |
|-------------------------|--|--|
| Anodonta cygnea | | |
| Argopecten gibbus | | |
| Austrovenus stutchburyi | | |
| Balanus spp. | | |
| Barbatia novaezelandiae | | |

Molluscs

Cerastoderma edule Chamelea gallina Chelonibia spp. Octolasmis spp. Crassostrea angulata Crassostrea ariakensis

| Molluscs | Molluscs | |
|----------------------------------|-------------------------------------|--|
| Crassostrea corteziensis | Ostrea denselammellosa | |
| Crassostrea gigas | Ostrea edulis | |
| Crassostrea nippona | Ostrea equestris | |
| Crassostrea rhizophorae | Ostrea lurida | |
| Crassostrea sikamea | Ostrea puelchana | |
| Crassostrea virginica | Ostrea stentina | |
| Dendostrea sandvicensis | Paphies australis | |
| Haliotis conicopora | Patinopecten yessoensis | |
| Haliotis corrugata | Pecten maximus | |
| Haliotis cracherodii | Pecten novaezelandiae | |
| Haliotis cyclobates | Perna canaliculus | |
| Haliotis discus hannai | Pinctada fucata | |
| Haliotis diversicolor | Pinctada margaritifera | |
| Haliotis diversicolor supertexta | Pinctada martensii | |
| Haliotis fulgens | Pinctada maxima | |
| Haliotis iris | Pinctada sugillata | |
| Haliotis laevigata | Pitar prostrata | |
| Haliotis rubra | Protothaca jedoensis | |
| Haliotis rubra x laevigata | Pyrazus spp. | |
| Haliotis rufescens | Batillaria spp. | |
| Haliotis scalaris | Ruditapes (Venerupis) philippinarum | |
| Haliotis sorenseni | Ruditapes decussatus | |
| Haliotis tuberculata | Saccostrea cucullata | |
| Haliotis wallalensis | Saccostrea glomerata | |
| Katelysia rhytiphora | Solen marginatus | |
| Macoma balthica | Trichomya hirsuta | |
| Macomona liliana | Tridacna crocea | |
| Mya arenaria | Tridacna gigas | |
| Mytilus edulis | Tridacna maxima | |
| Mytilus galloprovincialis | Venerupis aurea | |
| Ostrea angasi | Venerupis corrugata | |
| Ostrea chilensis | Venerupis philippinarum | |
| Ostrea conchaphila | Xenostrobus securis | |

Other

Ardea cinerea Argulus foliaceus Caligus coryphaenae

Other

Caligus elongatus Callibaetis spp. Lepeophtheirus salmonis

Department of Agriculture, Water and the Environment

Aquatic animal diseases significant to Australia: identification field guide 5th edition

Other

Nephtys australiensis

Nereis spp.

Paracartia grani

| Other |
|-------|
|-------|

Piscicola spp.

Salmincola spp.

Tubifex tubifex

Appendix B: Reporting suspected disease

If you see any unusual symptoms in wild or farmed aquatic animals, play it safe and **report it immediately**. Don't worry about how insignificant it may be—small signs may be an early indication of a serious disease problem.

National 24 hour emergency animal disease watch hotline: 1800 675 888 (freecall)

Each state or territory can also be contacted if you suspect a disease. Table 2 shows the state and territory government agencies responsible for aquatic health, and provides points of contact.

| State or territory | Government agency | Internet site | State telephone |
|---------------------------------|--|----------------------------|---|
| Australian Capital Territory | Environment Planning and Sustainable Development Directorate | www.environment.act.gov.au | 132 281 |
| New South Wales | Department of Primary Industries | www.dpi.nsw.gov.au | 1800 043 536 (fishwatch) |
| Northern Territory | Department of Environment and Natural Resources | <u>nt.gov.au</u> | 1800 891 136 (fishwatch) 0413 381 094 |
| Queensland | Department of Agriculture and Fisheries | www.daf.qld.gov.au | 13 25 23 or 07 3404 6999 |
| South Australia | Department of Primary Industries and Regions SA | www.pir.sa.gov.au | 1800 065 522 (fishwatch 24 hours) |
| Tasmania | Department of Primary Industries, Parks, Water and Environment | www.dpipwe.tas.gov.au | 1300 368 550 |
| Victoria | Department of Jobs, Precincts and Regions | agriculture.vic.gov.au | 136 186 |
| Western Australia | Department of Primary Industries and Regional Development | www.fish.wa.gov.au | 1800 815 507 (fishwatch) |

Table 2 State and territory government contact details for reporting aquatic animal disease

Appendix C: Further reading

These hyperlinks were correct at the time of publication.

General

CABI Invasive Species Compendium (search by disease)

CEFAS International Database on Aquatic Animal Diseases

Department of Agriculture, Water and the Environment <u>AQUAVETPLAN disease strategy manuals</u>

World Organisation for Animal Health Manual of diagnostic tests for aquatic animals

Finfish

Biosecurity Australia 2014, Importation of freshwater ornamental fish: review of biosecurity risks associated with gourami iridovirus and related viruses—provisional final import risk analysis report

Camus, AC 2004, Channel catfish virus disease, Southern Regional Aquaculture Centre

Elwell LC, Stromberg KE, Ryce EK, Bartholomew JL 2009, 'Whirling disease in the United States: a summary of progress and research and management 2009', *Trout Unlimited*

Fu X, Li N, Liu L, Lin Q, Wang F, Lai Y, Jiang H, Pan H, Shi C, Wu S 2011, 'Genotype and host range analysis of infectious spleen and kidney necrosis virus (ISKNV)', *Virus Gene*

Glenn RA, Taylor PW, Pelton EH, Gutenberger SK, Ahrens MA, Marchant LM, Hanson KC 2015, <u>Genetic evidence of vertical transmission and cycling of 'Yersinia ruckeri' in hatchery-origin fall</u> <u>chinook salmon 'Oncorhynchus tshawytscha'</u>, *Journal of Fish and Wildlife Management*.

Hazeri M, Hassan MD, Abba Y, Omar AR, Allaudin ZN, Soltani M, Hamdan RH, Mohamad NF 2017, <u>Molecular characterisation of grouper iridovirus isolates from Peninsular Malaysia</u>, *Journal Veterinar Malaysia*

Jansen MD and Mohan CV 2017, *Tilapia lake virus (TiLV): Literature review*, Penang, Malaysia: CGIAR Research Program on Fish Agri-Food Systems

Kumar G, Menanteau-Ledouble S, Saleh M, El-Matbouli M 2015, <u>'Yersinia ruckeri', the causative</u> agent of enteric red mouth disease in fish, *Veterinary Research*.

Ma H, Peng C, Su Y, Feng J, Guo Z 2016, 'Isolation of a Ranavirus-type grouper iridovirus in mainland China and comparison of its pathogenicity with that of a Megalocytivirus-type grouper iridovirus', *Aquaculture*

Menanteau-Ledouble S, Kumar G, Saleh M, El-Matbouli M 2016, <u>'Aeromonas salmonicida': updates</u> on an old acquaintance, Diseases of Aquatic Organisms

Network of Aquaculture Centres in Asia–Pacific Epizootic ulcerative syndrome: disease card

Network of Aquaculture Centres in Asia–Pacific Grouper iridoviral disease: disease card

Network of Aquaculture Centres in Asia-Pacific <u>Tilapia Lake Virus (TiLV)—A novel orthomyxo-like viru:</u> <u>disease card</u> Whittington RJ, Djordjevic SP, Carson J, Callinan RB 1995, <u>Restriction endonuclease analysis of</u> <u>atypical 'Aeromonas salmonicida' isolates from goldfish 'Carassius auratus', silver perch 'Bidyanus</u> <u>bidyanus', and greenback flounder 'Rhombosolea tapirina' in Australia</u>, *Diseases of Aquatic Organisms*

Molluscs

Australia and New Zealand standard diagnostic procedure Bonamiosis in Australian 'Ostrea angasi'

European Union Reference Laboratory for Molluscs Diseases <u>'Marteilioides chungmuensis'</u>

European Union Reference Laboratory for Molluscs Diseases 'Mikrocytos mackini'

Fisheries and Oceans Canada <u>'Marteilioides chungmuensis' of oysters</u>

Fisheries and Oceans Canada Gill disease of Portuguese oyster

Fisheries and Oceans Canada Oyster velar virus disease

Crustaceans

Network of Aquaculture Centres in Asia-Pacific <u>'Monodon' slow growth syndrome and Laem-Singh</u> virus retinopathy: disease card

Network of Aquaculture Centres in Asia-Pacific Acute hepatopancreatic necrosis: disease card

Network of Aquaculture Centres in Asia-Pacific <u>Hepatopancreatic microsporidosis caused by infection</u> with 'Enterocytooon hepatopenaei': disease card

Poornima, M, Seetang-Nun, Y, Alavandi, SV & Dayal, J 2012, <u>Laem-Singh virus: A probable etiological</u> <u>agent associated with Monodon slow growth syndrome in farmed black tiger shrimp</u> <u>('Penaeus monodon')</u>, *Indian Journal of Virology*, vol. 23.

Amphibians

Department of the Environment and Energy <u>Infection of amphibians with chytrid fungus resulting in</u> <u>chytridiomycosis (2016)</u>

European Food Safety Authority <u>Scientific and technical assistance concerning the survival</u>, establishment and spread of 'Batrachochytrium salamandrivorans' (Bsal) in the EU

Imperial College London Global Bd mapping project

Wildlife Health Australia Ranaviral disease in wild Australian amphibians

World Organisation for Animal Health, 'Batrachochytrium salamandrivorans' disease card