



Diarrheagenic *Escherichia coli* – Enrichment and isolation of *E. coli* serotype O157:H7 from Foods – FDA BAM Chapter 4A(K)

SCOPE

This method, updated in June 2016, is applicable to the analysis of food in general and is suitable for the enumeration and isolation of *E. coli* O157 from raw ground beef and trim. The approach recommended here allows the qualitative determination of *E. coli* O157 in raw ground beef and trim.

PRINCIPLES

Enrichment and isolation of *E. coli* O157 can be broken down into the following steps:

- **Enrichment¹**
A 25 g sample is diluted in 225 mL of mBPWp and incubated at $37 \pm 0.5^{\circ}\text{C}$ for 5 h. One mL of ACV supplement is then added and incubation continued at $42 \pm 1^{\circ}\text{C}$ static overnight (18-24 h). Positive controls are to be used as detailed in BAM Chapter 4A.
- **Real-time PCR Screening**
Lab must screen samples using Real-time PCR on enriched samples. The Real-time PCR protocol must be as per BAM Chapter 4A and is performed after immuno-magnetic separation. PCR negative samples are regarded as negative. PCR positive samples require cultural confirmation.
- **Isolation**
E. coli O157 is isolated by diluting the immuno-magnetic separated sample in Butterfield's phosphate buffer and spread plating 0.05 mL in duplicate onto TC-SMAC plates and one chromogenic agar (Rainbow Agar O157 or R&F *E. coli* O157:H7 agar). Plates are incubated at $37 \pm 1^{\circ}\text{C}$ for 18-24 h. Suspected colonies are confirmed using latex agglutination (Remel kit). All typical colonies are streaked onto TSAYE plates and incubated at 35°C for 18-24 h
- **Confirmation**
E. coli O157 is confirmed by indole production, lack of β -glucuronidase activity and serological tests. Presence of Shiga toxin or Shiga toxin genes or genetic confirmation of H7, confirms the presence of *E. coli* O157:H7.

¹ Enrichment with the following modification; must use the IMS option and a sample size of 325 g for ground beef, analysed as five separate 65 g portions, 1:10 dilution

CHECKLIST

| | | |
|---------------------|--|-------|
| Enrichment | Is the sample enriched in mBPWp? | _____ |
| | Is enrichment at $37 \pm 0.5^{\circ}\text{C}$ initially for 5 h? | _____ |
| | Is 1 mL ACV supplement added and incubation continued at $42 \pm 1^{\circ}\text{C}$ for 18-24 h? | _____ |
| | Is a positive control run with each batch of samples analysed? | _____ |
| | Are reference cultures inoculated into primary enrichment broth at a level of 10 to 100 cells? | _____ |
| Separation | Is <i>E. coli</i> O157 separated and concentrated from the enrichment broth using IMS? | _____ |
| | Is a screen Real-time PCR assay performed on IMS broth samples? | _____ |
| Isolation | Is IMS sample subcultured in duplicate onto TC-SMAC and onto one chromogenic agar plate? | _____ |
| | Are plates incubated at $37 \pm 1^{\circ}\text{C}$ for 18-24 h? | _____ |
| | Are suspect colonies streaked onto TSAYE and incubated at 35°C for 18 to 24 h. | _____ |
| Confirmation | Is <i>E. coli</i> O157 confirmed by: | _____ |
| | Indole production? | _____ |
| | β -glucuronidase activity? | _____ |
| | Agglutination of <i>E. coli</i> O157 antiserum? | _____ |
| | Are tests for toxin or toxin genes carried out? | _____ |